# **Crystallization Optimizing of Cefradine**

Lei Du\*,1 and Wenbo Luo<sup>2</sup>

**Abstract:** Cefradine crystallization was studied under different conditions. The optimal conditions for crystallization were: dissolving temperature, 15°C; initial temperature in crystallization, 30°C; cooling temperature in crystallization, 0°C; and adding 1,2-propanediol as adjuvant agent with a volume ratio 0.2 of water solution. The yield of cefradine crystallization is about 92% under these conditions. The purity of cefradine crystals is over 98% under the same conditions. The average size of cefradine crystal is 500 μm. The content of cefalexin in crystal is reduced to only 0.36%.

**Keywords:** Cefradine, crystal, crystal seed, optimization.

#### 1. INTRODUCTION

Cefradine (cephradine, cephalosporin IV, sefril), 7-[a-D-(cyclohexa-1,4-dienyl)- glycyl-amino]-3-methyl-3-cephem-4-carboxylic acid ( $C_{16}H_{19}N_3O_4S$ ), is a first-generation cephalosporin, originally isolated in 1948 [1]. It exhibits broad-spectrum antibacterial activity against gram-positive and gram-negative microorganisms by inhibiting bacterial cell wall synthesis. Cefradine is useful for treatment of infections of the urinary and respiratory tracts, skin, and soft tissues. However, the solubility of cefradine in most solvents is low, resulting in limited bioavailability and/or erratic absorption [2]. The crystallization mainly includes precipitate crystallization, dilution crystallization and reaction crystallization. The crystallization of cefradine belongs to the reaction crystallization which is usually hard to control because of its high reaction rate [3-15]. Therefore, the aim of this study was to optimize the crystallization process to increase the yield and purity of cefradine crystals.

# 2. EXPERIMENTAL

#### 2.1. Materials

Commercial crude cefradine was purchased from BETA Inc. (NCPC, China). Standard cefradine and cefalexin were obtained from Sigma. All the other chemicals were of analytical reagent grade; the solutions were prepared with double-distilled water.

# 2.2. Preparation of Cefradine Solution and Crystal Seed

Five (5.00) g of cefradine was added in 0.25% NaHSO<sub>3</sub> (W/V) solution and dissolved completely by slowly adding 12 M HCl while stirring. The solution pH value was then adjusted to 2.8–2.9 by adding dimethylamine/ethylenediamine (1:2, V/V). Next, 5% of the solution volume was removed and the remained was reserved; we continued stirring this solution at 25°C and used it to seed nucleation after ultrasonic wave radiation (30 kHz, 70 kW/cm²) for 10–20 seconds.

# 2.3. Crystallization of Cefradine

After adding the seed crystals to the reserved solution, the pH value of resulting solution was adjusted to 3.0–3.1 by adding dimethylamine/ethylenediamine (1:2, V/V). The solution was kept for 0.5 h under the higher temperature, and 1,2-propanediol at 0.2 ratio by volume was added to the mixture. The pH value of the mixture was then adjusted to 4.6–4.7 by addition of dimethylamine/ethylenediamine (1:2, V/V). The mixture was cooled and left standing for 0.5 h. The crystals were washed twice with 95% ethanol and leached twice with absolute ethyl alcohol. After vacuum filtering, the crystals of cefradine were dried in a vacuum at 30°C for 1–2 h.

#### 2.4. High-Performance Liquid Chromatography (HPLC)

The HPLC system used was Agilent 1100 (Agilent, USA). The chromatographic system consisted of a polymeric reversed-phase PLRP-S HPLC column (1503 4.6 mm I.D., 5 mm; Polymer Labs., Church Stretton, UK) with an in-line 2-mm pre-filter. Analyses were performed at ambient temperature with a mobile phase of 10.5% (v/v) acetonitrile in 20 mM ammonium dihydrogen orthophosphate (pH 2.75) at a flow-rate of 1 mL/min. The UV detection wavelength used was 260 nm and sample aliquots of 100 mL were injected into the chromatographic system.

# 2.5. Fourier Transforms Infrared Spectroscopy (FTIR)

FTIR spectra were recorded with a Bruker Vector 33 spectrometer in the 500–4000 cm<sup>-1</sup> range at a resolution of 2 cm<sup>-1</sup>; 32 scans were run. Samples were diluted at 1% concentration with KBr mixing powder and pressed to obtain self-supporting disks.

# 2.6. X-Ray Diffraction Studies (XRD)

X-ray diffraction analysis was performed using the D/max-IIIA (SHIMADZU Inc., Japan) to detect the physical characteristics and crystallinity of the cefradine. The measuring unit consisted of a rotating anode in transmission technique with the following specifications: Cu  $K\alpha_1$  radiation generated at 30 mA and 40 kV. The scanning speed was  $10^{\circ}/\text{min}$ , from  $5^{\circ}$  to  $55^{\circ}$ , with a step size of  $0.02^{\circ}$ .

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#### 3. RESULTS AND DISCUSSIONS

# 3.1. The Effect of Dissolving Temperature on Cefradine Crystallization

The process of dissolving cefradine is the reaction of cefradine with HCl, which is obviously influenced by different temperatures (Fig. 1). Both the crystal yield and the purity of cefradine decrease rapidly with increasing dissolving temperature above 15°C. However, the content of cefalexin in solution increases rapidly as temperature rises above 15°C (Fig. 2). The results show that cefalexin is the main product of the side reaction [16].

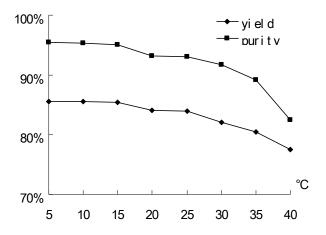


Fig. (1). The effect of dissolving temperature on cefradine crystallization.

# 3.2. Initial and Cooling Temperature for Crystallization

In the crystallization step, the initial and final temperatures were regulated to optimize the crystallizing process. Raising the initial temperature results in a noticeable increase in average crystal size (Fig. 3). Conversely, crystal purity declines greatly as the temperature

rises. These results show that a higher initial temperature accelerates not only the speed of crystallization of cefradine, but also that of the side reaction in the system. In addition, the growth rate of crystal size is lower when the solution temperature is above 30°C because the generation of new seed crystals offsets the increased reaction speed.

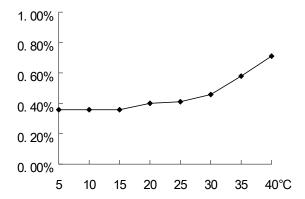


Fig. (2). The content of cefalexin at different dissolving temperatures.

In the crystallization process, a cooling strategy is often used to reduce the solubility of a molecule and to improve the crystal yield. Moreover, the lower temperature can suppress the oxidation reaction of cefradine in solution. A slight increase in crystal yield, and in crystal purity as well, was observed as the cooling temperature of cefradine crystallization was lowered (Fig. 4).

## 3.3. Optimize Agent of Crystal Feature

It is often useful to add specific agents to optimize the process of crystallization. The agents are used to increase the solubility of the molecule or to slow the speed of crystallization by changing thermodynamic and dynamic parameters in the crystalline system. In this study,

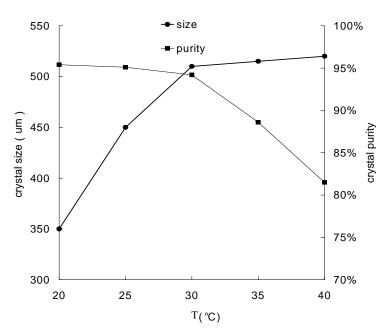


Fig. (3). The effect of initial crystal temperature on cefradine crystallization.

Conventional organic solvents with arginine was compared, which is often mixed with cefradine in commodity products. It is found that 1,2-propanediol displays the best performance and can be used as an adjuvant in cefradine crystallization (Table 1).

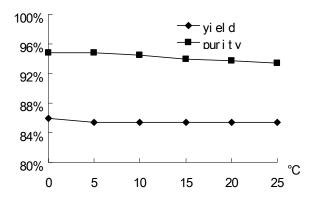


Fig. (4). The effect of cooling temperature on cefradine crystallization.

Comparison of the Agents for Optimizing Cefradine Table 1. **Crystal Structure** 

	Crystal Structure	Average Size (µm)	Yield (%)	Purity (%)	Content of Cefalexin (%)
alcohol	rhabditiform	350	85.0	95.2	0.39
acetonitrile	aciculiform	300	84.6	95.3	0.38
isopropanol	slender	330	84.5	96.1	0.38
propyl alcohol	slender	350	84.6	94.8	0.39
butanol	slender	350	84.6	94.8	0.39
methanol	flake	300	83.5	91.6	0.42
aether	aciculiform	280	84.2	94.5	0.42
acetone	aciculiform	260	84.7	95.1	0.38
1,2-propanediol	clavate	380	84.9	96.4	0.38
arginine	aciculiform	300	84.2	94.0	0.38

### 3.4. Optimization of Cefradine Crystallization

According to the previous results, the optimal conditions for cefradine crystallization are: dissolving temperature, 15°C; initial temperature, 30°C; cooling temperature, 0°C; and addition of a 0.2 ratio by volume of 1,2-propanediol as the adjuvant reagent (Table 2). Compared with the control, the crystal yield increased from 89.4% to 92.3%. The crystal purity also increased from 90.7% to 98.2%. The average size of a cefradine crystal increased from 220 µm to 500 µm (Fig. 5). The content of cefalexin in the crystals decreased from 2.1% to 0.36%.

Through a comparison of the characteristics of a cefradine crystal using FTIR and XRD, it can be concluded that there is no apparent difference in either the composition or the crystal structure between the cefradine crystals

Table 2. The Result of Cefradine Crystallizing in Optimizing Condition

	Control	Optimizing Condition
Crystal yield (%)	89.4	92.3
Crystal purity (%)	90.7	98.2
Average size of crystal (µm)	220	500
Content of cefalexin (%, w/w)	2.1	0.36
Water content (%, w/w)	3.9	3.7

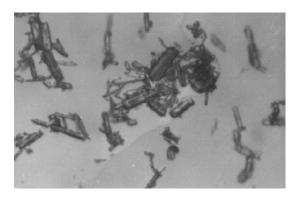




Fig. (5). Crystal photo of cefradine crystal (top: commercial crude powers, bottom: optimizing crystal. 100×).

obtained from the optimized condition in our experiments with the standard commercially available cefradine crystal (Figs. 6-8) we used as a starting material.

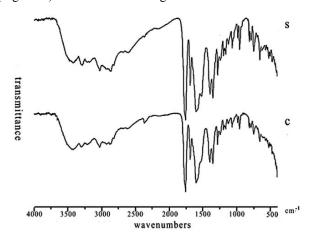


Fig. (6). FTIR spectra of cefradine under optimized crystallization conditions (s) vs Standard (c).

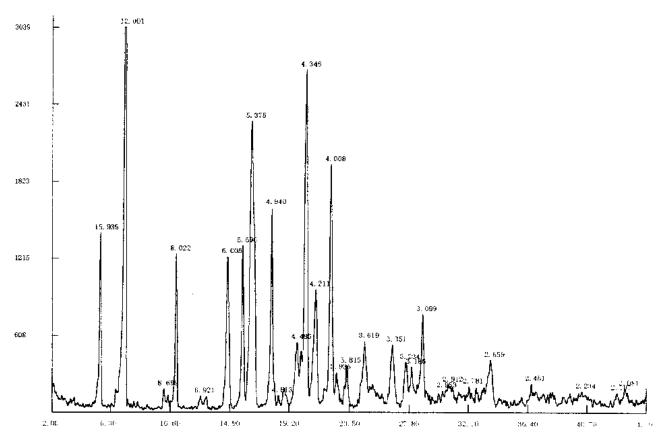


Fig. (7). X-rays diffraction of a cefradine crystal produced under optimized conditions.

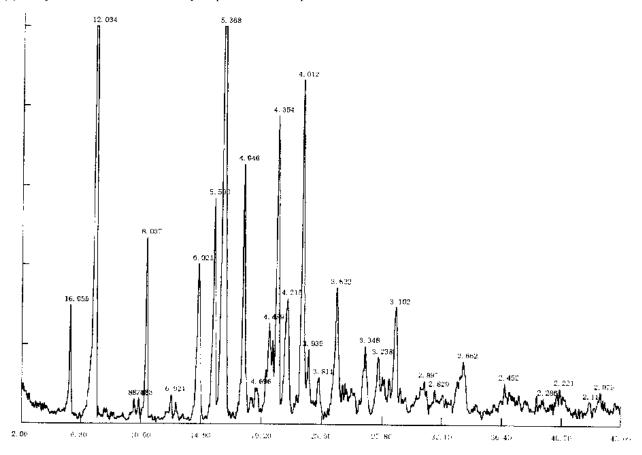


Fig. (8). X-ray diffraction pattern of a standard cefradine crystal.

#### 4. CONCLUSIONS

In this paper, we discuss different conditions for cefradine crystallization, including dissolving temperature, initial and cooling temperature for crystallization, and adjuvant agents for optimizing crystal characteristics. The optimal conditions for cefradine crystallization are: dissolving temperature, 15°C; initial temperature for crystallization. 30°C: cooling temperature crystallization, 0°C; and addition of 1,2-propanediol at a 0.2 ratio by volume as the adjuvant agent for crystal characteristics. The yield of cefradine crystals is about 92% under these conditions, and the purity of the obtained cefradine crystals is over 98%. The average size of a cefradine crystal approaches 500 um, and the content of cefalex in in the crystals is reduced to only 0.36%.

#### REFERENCES

- Joseph, E.D.; Harold, E.A. A new class of semisynthetic penicillins [1] and cephalosporins derived from D-2-(1,4-cyclohexa- dienyl) glycine. J. Med. Chem., 1971, 14(2), 117-119.
- Jie, Z.; Shen, Z.; Yang, Y.; Chen, J.F. Preparation and [2] characterization of uniform nanosized cephradine by combination of reactive precipitation and liquid anti-solvent precipitation under high gravity environment. Int. J. Pharm., 2005, 301, 286-293.
- Frank, L.W.; Joseph, E.D.; Georges, G.B.; Jack, B. Alpha-amino-[3] cyclohexadienylalkylene- -penicillins and cephalosporins. U.S. Pat., US3485819, 1969.

- Herman, B.B.; Treuner, U. 2-(Thiocarbonylamino)acetamidocepha-[4] losporanic acid compounds. U.S. Pat., US3862941, 1975.
- [5] Diassi, P.A.; Atwal, M.S. Process for the preparation of 7-(D-2amino-2(1,4-cyclo-hexadienyl) acetamido) desacetoxycephalosporanic acid and 7-(D-2-amino-2-(1,4-cyclohexadienyl) acetamido) cephalosporanic acid. U.S. Pat., 3912726, 1975.
- [6] Robinson, C.A. Intermediates for preparing cephalosporins and methods of production. U.S. Pat., US3965098, 1976.
- [7] Faarup, P. Method of preparing a sparingly soluble complex of cephalexin. U.S. Pat., US4003896, 1977.
- [8] Broggi, R.; Falciani, M. Process for preparing cephalosporines. U.S. Pat., US4139702, 1979.
- Bouzard, D.; Weber, A.; Stemer, J. Process for the preparation of [9] the crystalline monohydrate of 7-[D-α-aα-(p-hydroxyphenyl)acetamido]-3-methyl-3-cephem-4-ca rboxylic acid. U.S. Pat., US4160 863, 1979.
- [10] Walker, D.; Silvestri, H.H.; Sapino C.; Johnson, D.A. Production of cephalosporins. U.S. Pat., US4223135, 1980.
- [11] Palomo-coll, A.; Palomo-coll, A.L. Process for the preparation of solutions of 7-aminocephalosporanic acids. U.S. Pat., US4405782,
- [12] Meseguer, J.D.; Codes, R.B.; Ciriza, S.A. Stable cephradine hydrate. U.S. Pat., US5278157, 1994.
- [13] Diago, J.; Ludescher, J. Beta lactam production. U.S. Pat., US5719276, 1998.
- Diago, J.; Ludescher, J. Processes for the production of 6-α-[14] aminoacyl-penicillin and 7-.alpha. U.S. Pat., US5840885, 1998.
- [15] Centellas, V.; Diago, J.; Ludescher, J. Silylation process. U.S. Patent, US5998610, 1999.
- Christel, H.; Eugène, R.; Valère, B.; Josiane, T.; Martine, P.; Roger, B.; Gerard, J.; Jos, H. Synthesis of potential impurities of [16] cefalexin and cefradine. Archiv. Pharm., 1994, 327(4), 215-219.

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