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## RESEARCH ARTICLE

### Temporal Variation in Antibiotic Resistance of *Acinetobacter baumannii* in a Teaching Hospital in Tunisia: Correlation with Antimicrobial Consumption

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#### Abstract:

#### Background:

To investigate the potential correlation between the rates of antimicrobial drug consumption and the prevalence of antimicrobial resistance among clinical *Acinetobacter baumannii* recovered in a tertiary care hospital in Tunisia.

#### Methods:

The microbiological and epidemiological profiles of *A. baumannii* infections at the Hospital Sahloul, Sousse, were investigated between 2001 to 2004 and 2012 to 2015 along with the consumption record of broad-spectrum antibiotics.

#### Results:

Our data showed that extensively drug-resistant *A. baumannii* (XDRAb) isolates increased from 11.2% to 30.5% between 2012 and 2015 and disseminated endemically for a long time. Furthermore, we evidenced a drastic increase of carbapenem-resistant *A. baumannii* isolates from 29.5% in 2001 up to 88.6% in 2015 (612/691). This rise could be paralleled with a significant increase in antibiotic consumption over the last 15 years, especially with the sharp increase in the annual consumption of imipenem ( $r = 0.816$  and  $p < 10^{-3}$ ). A noteworthy correlation between carbapenem use and resistance rate ( $r = 0.778$ ,  $p < 0.001$ ) was evidenced.

#### Conclusion:

Feedback of these data to clinicians and decision-makers in the local setting was crucial to promote the rational use of antimicrobials and to raise awareness to strictly implement hygiene measures to limit the spread of these XDRAb isolates, to prevent colonization and subsequent infection.

**Keywords:** Antimicrobial resistance, Antimicrobial consumption, MDR-Ab, XDR-Ab, DDD/1000 patient-day, Imipenem.

#### Article History

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## 1. INTRODUCTION

*Acinetobacter baumannii* has emerged as one of the most

troublesome pathogens for health care settings worldwide [1]. The World Health Organization (WHO) declared it as one of the most serious ESKAPE organisms in public health [2]. They have been involved in a variety of Health Care Associated Infections (HCAIs), associated with extremely high crude mortality rates mainly among patients in intensive care units

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(ICUs).<sup>1,3</sup> Its clinical significance, has been propelled by its capacity to tolerate desiccation and its remarkable propensity to accumulate enzymatic and non-enzymatic resistance mechanisms [1]. The combination of several of these resistance mechanisms, especially those conferring resistance to carbapenems, will limit the therapeutic options for severe infections. Carbapenem-resistant *A. baumannii* (CRAb) infections are usually resistant to all antibiotics, except colistin [3]. In the present study we (i) describe the microbiological and epidemiological characteristics of *A. baumannii* isolates identified in the clinical microbiology laboratory in a Tunisian teaching hospital between 2012-2015; (ii) present the trend of the antibiotic resistance rate and correlate them with the antibiotic consumption over a 15-year-period.

## 2. MATERIALS AND METHODS

### 2.1. Clinical Settings, Study Design and Bacterial Isolates

A descriptive study was performed at the Sahloul Hospital, Sousse, Tunisia, a 629-bed teaching hospital, over a four-year period, from 2012 to 2015. *A. baumannii* isolates were identified through the clinical microbiology laboratory data base. Duplicate isolates were excluded, based on antibiotic patterns and time period. Isolates were considered different when complying the following criteria one major and three minor differences in antibiotic patterns and a 30-day time period between two isolates. According to these criteria, a total of 691 isolates were retained and recovered from 573 hospitalized patients.

For further statistical analysis, we retrieved data concerning the resistance features from period ranged between 2001 and 2004 in order to highlight the global evolution of the antibiotic resistance. We considered the period between 2001 and 2004 as period 1 (270 isolates) and that between 2012 and 2015 as period 2 (691 isolates).

### 2.2. Microbiological Methods

The isolates were identified by conventional methods and using the Vitek 2 automatic system (BioMérieux, Tunis, Tunisia). Antimicrobial susceptibility of *A. baumannii* isolates was assessed by a disc diffusion method based on MH agar which was performed as recommended by the European Committee on Antimicrobial Susceptibility Testing (EUCAST, 2015, www.eucast.org).

### 2.3. Definitions

According to the work of a group of international experts following definitions were used in the medical literature to characterize the different patterns of resistance found in healthcare-associated, antimicrobial resistant bacteria [4]:

**MDR** (for Multidrug-Resistant) was defined as acquired non-susceptibility to at least one agent in three or more antimicrobial categories listed in the worksheet for categorizing isolates.

**XDR** (for Extensively Drug-Resistant) was defined as non-susceptible to all but two or fewer antimicrobial categories (*i.e.* bacterial isolates remain susceptible to only one or two

categories listed in the worksheet for categorizing isolates).

**PDR** (for Pandrug-Resistant) was defined as non-susceptible to all agents in all antimicrobial categories listed in the worksheet for categorizing isolates.

### 2.4. Antibiotic Consumption

Data regarding antibiotic consumption and the occupancy from 2001 to 2015 were collected from the Pharmacy department and statistics office of the hospital, respectively. We focused on broad-spectrum antibiotics, used to treat severe infections with Multidrug resistant *A. baumannii* isolates (MDRAb). The extracted data were converted to DDD is defined daily dose per 1,000 inpatient-days used according to the Anatomic Therapeutic Chemical classification system defined by WHO Collaborating Centre for Drug Statistics Methodology according to the ATC/DDD Index 2017 ([https://www.whocc.no/atc\\_ddd\\_index/](https://www.whocc.no/atc_ddd_index/)).

### 2.5. Statistical Analysis

Descriptive statistics was used for all the studied variables. A chi-squared test was used to compare susceptibility rates between period 1 and 2. A *P*-value (*P*) below 0.05 was deemed statistically significant. A linear regression was applied to assess local trends in the consumption of antibacterial agents (*P* ≤ 0.05 considered for statistical significance). Normality was tested using the Shapiro Wilk test. For the correlation analysis, the nonparametric Spearman correlation coefficient was used. Statistical analyses were performed using SPSS 17.0 Statistical package (SPSS Inc, Chicago, USA).

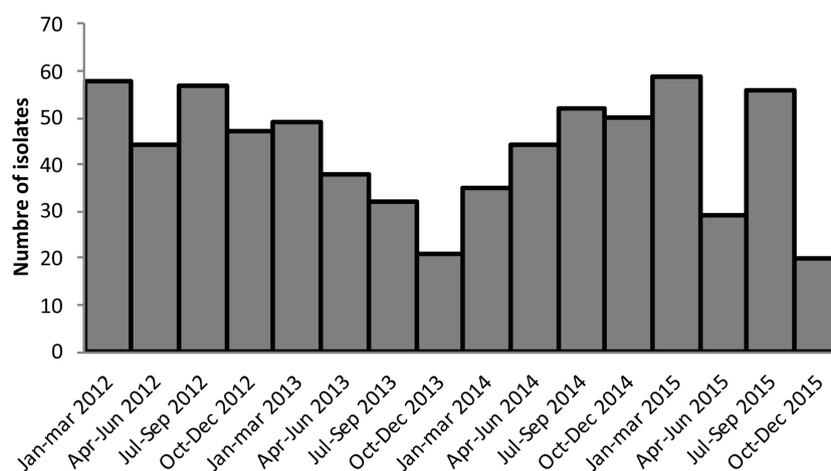
## 3. RESULTS AND DISCUSSION

### 3.1. Demographic and Microbiological Data

A total of 691 *A. baumannii* were collected over four years in a teaching hospital in Sousse-Tunisia. The majority 536/691 (77.6%) of the isolates were recovered from patients hospitalized in the Intensive Care Units (ICU). The predominant clinical samples that were positive for *A. baumannii* were blood 264/691 (38.2%), urines 149/691 (21.6%), respiratory tract 78/691 (11.3%) and biopsies, urethral samples, articular fluids, ascites, pleural, sinusual, and synovial fluids 200/691 (28.9%).

The clinical epidemiologic data obtained by retrospective chart review clearly revealed the endemicity of *A. baumannii* over a 2012-2015 4-year period, as shown by the epidemic curve (Fig. 1). In fact, *A. baumannii* can assemble and modulate a host of antimicrobial resistance mechanisms to survive the selective pressure they encounter in the hospital environment, providing them with a strong ecological advantage [1].

Antimicrobial susceptibility testing results revealed a high rate of resistance to most antimicrobials tested, exceeding 80% in most cases. Carbapenem resistance was observed in 612 (88.6%) *A. baumannii* isolates. Only four antibiotics still revealed some activity (> 50% susceptibility) against these *A. baumannii* isolates: Co-trimoxazole (Sulphamethoxazole/trimethoprim), Netilmicin, rifampicin and colistin (Table 1, Table S1).



**Fig. (1).** Epidemic curve over four years (2012-2015): an overview of the epidemiological dissemination of *A. baumannii* both in its epidemic and endemic forms in the local setting.

**Table 1.** Antibiotic resistance rate (%) of *A. baumannii* isolates between 2001–2004 (period 1) and 2012–2015 (period 2) evaluated in this study.

Antibiotic	Period 1 2001-2004	Period 2 2012-2015	P-value
TIC	64.6	93	<10 <sup>6*</sup>
PIP	85.75	91	0.019*
CAZ	90	89.4	0.796
IMP	40	88.6	<10 <sup>6*</sup>
GM	71	85.2	<10 <sup>6*</sup>
TM	29	62	<10 <sup>6*</sup>
NET	24.5	40.6	<10 <sup>4*</sup>
AN	79	79.2	0.925
OFX	88	91.8	0.083
CIP	88	91.7	0.083
SXT	89	51.5	<10 <sup>6*</sup>
RA	91.6	26.3	<10 <sup>6*</sup>
COL	0	0.3	--
<b>Total</b>	<b>270</b>	<b>691</b>	

\*p significant at the 0.05 level

TIC: Ticarcillin, PIP: Piperacillin, TZP: Piperacillin / tazobactam, CAZ: Ceftazidime, IMP: Imipenem, GM: Gentamicin, TM: Tobramycin, NET: Netelmicin, AN: Amikacin, OFX: Ofloxacin, CIP: Ciprofloxacin, SXT: Sulphamethoxazole / trimethoprim, RA: Rifampicin, COL: Colistin.

According to the definitions cited below 69.6% (481/691) were MDR*Ab* and 22.6% (156/691) were XDR *A. baumannii* (XDR*Ab*) (Table S2). The prevalence of XDR*Ab* significantly increased throughout the duration of the study ( $p < 10^{-3}$ ). The rate of the MDR*Ab* and XDR*Ab* isolates in the ICU is significantly higher as compared to the general hospital setting showing 83% and 79% ( $p < 10^{-3}$ ) respectively. There were 42 isolates, which remained susceptible only to colistin (27% of the total XDR*Ab*). Only two isolates from 2012 were colistin resistant with MICs > 128mg/L.

In the last decade, the empirical therapy for treatment of *A. baumannii* infections includes the aminoglycosides, fluoroquinolones, and carbapenems owing to their efficiency and safety against this organism [3, 5]. Our data clearly show an increasing trend of resistance to these antibiotics over the time

course of this study, with a resistance rate of 88%. The expanded surveillance network, which encompasses about ten teaching Tunisian hospitals (LART: L'AntibioRésistance en Tunisie) [6] reported 75.7% in 2014. The same high rate was reported in Europe, with generally higher resistance percentages observed in countries in the east and south of Europe than in the north. The combined resistance to all three groups ranged from 83% to 90% in Greece and Italy [7]

To highlight the evolution of resistance locally, we compared susceptibility rates between the 2001-2004 (period 1) and 2012–2015 (period 2) periods. The most remarkable finding of our study was a statistically significant increase of imipenem resistance from 40% (period 2001-2004) to 88.6% (period 2012-2015) ( $p < 10^{-6}$ ) (Table 1). Similarly, a significant increase in the rate of resistance ( $p < 0.05$ ) for all the aminoglycosides was observed. Whilst, there was a remarkable decrease ( $p < 10^{-6}$ ) in sulfamethoxazole and rifampin resistance rate. Only colistin remained constantly efficient against *A. baumannii* isolates during the study period.

Already, in 2006, the multicenter pilot survey of resistant bacteria in the Mediterranean area brings up the fact that imipenem-resistant *A. baumannii* seems to be a particularly prevalent organism in Tunisia with 60% of resistance to imipenem especially in ICU [8]. The same alarming findings were reported by Thabet *et al.* [9] in Tunisia, where the resistance rate to imipenem raised from 63.9% between 2005-2008 to 89.3% between 2008-2011 in the intensive care burn unit and a teaching hospital in the capital. The national survey (LART) [6] reported the rates to imipenem ranging from 44.9% in 2008 to 86.9% in 2014. The European Antimicrobial Resistance Surveillance Network (EARS-Net) report the same high rate in Greece (88-95%) and Italy (87–92%) [7]. Resistance to carbapenems is still lower in Spain (64.4%) in 2014 [7].

Globally, data reported from studies all over the world show that with the exception of polymyxins and tigecyclin, high rates of resistance to all classes of antibiotics have been observed [1, 10, 11]. Indeed, tigecyclin is a broad-spectrum modified minocycline derivative considered as therapeutic alternative showing excellent *in vitro* activity against MDR*Ab*

Table 2. Trends in antimicrobial consumption at university hospital of Sahloul, Sousse, Tunisia, 2001 to 2015.

Antibiotic	Antibiotic Consumption (DDD/1000 patient-days) by Year															Correlation		
	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	2011	2012	2013	2014	2015	r	p	R2
CAZ	3.8	3.2	2.8	1	3	6	4	3.6	3	3.5	4	7	3.5	7	5.5	0.574	0.025	0.330
IMP	12.9	14.5	12.6	13	22	27	27.5	25.6	19	24	25.5	23	29	27	32.5	0.816	<10 <sup>3</sup>	0.665
AN	9	9	8	4	33	10	11	8	6.6	7.3	9	8	8.5	10	7	-0.17	0.539	0.030
GM	43.5	45.2	42.5	14	33	27	28	24	23	19.5	19	16	19	23	23.5	0.71	0.003	0.502
LVX	0	0.1	2	1	3.5	5	4.5	2	4	3	2	3	4.5	3.5	3.2	0.56	0.030	0.314
CIP	17.9	23	22	11	34.5	37	34	36.5	35.7	39	33	44.5	31.5	41	17	0.46	0.085	0.211
FOS	1.3	3.4	4.5	2	5	2.6	4	7	6.5	6.3	7	5	10.5	8	8	0.834	<10 <sup>3</sup>	0.696
SXT	7.6	5.8	5	1.5	4	3.5	2	4.5	3.5	3	3	2.5	3	4	6	-0.3	0.280	0.089
RA	53	53.8	53	23	23	45	46	38.6	40	24.5	35.5	41	44	49	50	-0.05	0.865	0.002
COL	0.1	0.1	0.1	0.1	0.1	0.1	0.3	0.2	0.3	0.2	0.3	1.3	1.3	2	2	0.821	<10 <sup>3</sup>	0.675

DDD: defined daily dose; r: Spearman's rho; p: p-value \* Statistically significant association ( $r > 0.72$ ,  $P < 0.05$ ); CI, 95% confidence interval

CAZ: Cefazidime, IMP: Imipenem, AN: Amikacin, GM: Gentamicin, LVX: Levofloxacin, CIP: Ciprofloxacin, FOS: Fosfomycin, SXT: Sulphamethoxazole / trimethoprim, RA: Rifampicin, COL: Colistin.

[12]. Nevertheless, we've noted in our series a high rate of resistance ranged from 61.1% to 89.4% and this value is alarming as compared to worldwide studies results. Mendes *et al.* [11] conducted a large assessment of tigecyclin activity tested against 5127 *Acinetobacter spp.* They noted a slight increase in tigecyclin resistance rates occurred in Europe (from 1.3-1.5% in 2005-2006, to 3.2-5.8% in 2007- 2008) and Latin America (from 0.4-0.9% in 2005-2006, to 5.3-6.9% in 2007-2008).

The decrease of resistance of co-trimoxazole was remarkable reaching 50% in periods 2. These rates were lower than several studies where resistance rate to co-trimoxazol within XDRAb were ranged from 97.9% to 100% [13]. We also noted in our study a significant correlation between resistance to co-trimoxazole and phenotype. It shows that resistance for non-MDRAb isolates was 5.5% versus 40.2% for the MDRAb and 91.7% for XDRAb isolates.

\*Correlation is significant at the 0.05 In this study, colistin has shown a good *in vitro* activity against *A. baumannii* isolates. Only two isolates from 2012 were colistin resistant with MICs > 128mg/ml. This is a noteworthy finding since colistin has been revisited as the last defense line against MDR Gram-negative bacilli [16].

The patterns of susceptibility to rifampin in our series reflected a good potency against MDRAb, even though it is not recommended to use rifampin in monotherapy because of the rapid development of resistance. This effect can be blunted by the association with imipenem or sulbactam/ampicillin or colistin [14, 15].

### 3.2. Trends in Antimicrobial Consumption

To allow comparison of our findings with the previous study we focused only on studies using the same measurement unit (*i.e.*, DDD per 1 000 inhabitants and per day) for reporting antimicrobial consumption. The most prominent finding during the study period was a significant increase in the annual DDD/1000 patient-days of IMP ( $r = 0.816$  and  $p < 10^{-3}$  and a remarkable 20-fold increase in the colistin consumption ( $r = 0.816$  and  $p < 10^{-3}$ ) (Table 2, Table S3, Fig. S1). Our findings are in line with the ESAC-Net results [17] between 2008 and

2012 that showed a significantly increasing trend in the consumption of imipenem in seven countries (Denmark, Estonia, France, Ireland, Norway, Slovenia and Sweden).

A remarkable 20-fold increase of colistin consumption ( $r = 0.821$ ,  $p < 10^{-3}$ ) was noted. Indeed, the dramatic rise of XDR strains may further explain the increase in consumption of so-called "last resort" antimicrobial agent. However, the ESAC-NET reported that there is no significant change at European country level in polymyxin consumption during 2010–2014, but it increased significantly in three individual countries (Denmark, Hungary and Italy) [17].

### 3.3. Correlation Between Antimicrobial Consumption and Resistance Rate

Unlike several published studies [18, 19], only imipenem consumption and resistance to this subgroup showed a strong positive correlation ( $r > 0.72$  and  $P < 0.05$ ) (Figure S1). In fact, carbapenems remained for a long time the only active beta-lactam against MDRAb; this has led to an increase in their use not only for documented infections but also for empirical treatment of acquired hospital infections especially in ICU setting [3, 5]. Our finding support a recent study by Munoz-Price *et al.* [20] who demonstrated that the risk for acquisition of carbapenem resistant *A. baumannii* (CRAb) quadrupled with carbapenem exposure and that every additional carbapenem DDD increased the chance of acquiring CRAb by 5.1%. Wide-ranging approach has yielded varying results worldwide, but it is generally assumed that the selective pressure due to the excessive use over the last 50 years of antimicrobials has a significant upshot on bacterial resistance.

### CONCLUSION

The escalating problem of antimicrobial resistance in our health care setting is the consequence of the overuse and misuse of antimicrobials leading to increasing selection pressure, and the likely spread of antimicrobial-resistant microorganisms, as recently shown for *A. baumannii* [21]. Feedback of these data to clinicians and decision-makers at the

hospital were crucial to promote the rational use of antimicrobials and to improve compliance with standard and transmission-based precautions and other infection prevention strategies.

#### ETHICS APPROVAL AND CONSENT TO PARTICIPATE

Not applicable.

#### HUMAN AND ANIMAL RIGHTS

No animals/humans were used for studies that are the basis of this research.

#### CONSENT FOR PUBLICATION

Not applicable.

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#### CONFLICTS OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

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#### SUPPLEMENTARY MATERIAL

Supplementary material is available on the publishers website along with the published article.

#### REFERENCES

- [1] Peleg AY, Seifert H, Paterson DL. *Acinetobacter baumannii*: Emergence of a successful pathogen. *Clin Microbiol Rev* 2008; 21(3): 538-82. [http://dx.doi.org/10.1128/CMR.00058-07] [PMID: 18625687]
- [2] Boucher HW, Talbot GH, Bradley JS, et al. Bad bugs, no drugs: No ESCAPE! An update from the Infectious Diseases Society of America. *Clin Infect Dis* 2009; 48(1): 1-12. [http://dx.doi.org/10.1086/595011] [PMID: 19035777]
- [3] Doi Y, Murray G, Peleg A. *Acinetobacter baumannii*: Evolution of antimicrobial resistance: Treatment options. *Semin Respir Crit Care Med* 2015; 36: 85-98.

- [4] Magiorakos A-P, Srinivasan A, Carey RB, et al. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clin Microbiol Infect* 2012; 18(3): 268-81. [http://dx.doi.org/10.1111/j.1469-0691.2011.03570.x] [PMID: 21793988]
- [5] Maragakis LL, Perl TM. *Acinetobacter baumannii*: Epidemiology, antimicrobial resistance, and treatment options. *Clin Infect Dis* 2008; 46(8): 1254-63. [http://dx.doi.org/10.1086/529198] [PMID: 18444865]
- [6] Boutiba I, et al. <https://www.infectiologie.org.tn/resistance.php>
- [7] [http://ecdc.europa.eu/en/healthtopics/antimicrobial-resistance-and-consumption/antimicrobial\\_resistance/EARS-Net/Pages/EARS-Net.aspx](http://ecdc.europa.eu/en/healthtopics/antimicrobial-resistance-and-consumption/antimicrobial_resistance/EARS-Net/Pages/EARS-Net.aspx) n.d.
- [8] Marazian K, Fendri C, Missoum MFK, et al. Multicenter pilot survey of resistant bacteria in the Mediterranean area. *Eur J Clin Microbiol Infect Dis* 2006; 25(5): 340-3. [http://dx.doi.org/10.1007/s10096-006-0125-z] [PMID: 16601956]
- [9] Thabet L, Zoghalmi A, Boukadida J, Ghanem A, Messadi AA. [Comparative study of antibiotic resistance in bacteria isolated from burned patients during two periods (2005-2008, 2008-2011) and in two hospitals (Hospital Aziza Othmana, Trauma and Burn Center)]. *Tunis Med* 2013; 91(2): 134-8. [PMID: 23526277]
- [10] Kempf M, Rolain J-M. Emergence of resistance to carbapenems in *Acinetobacter baumannii* in Europe: clinical impact and therapeutic options. *Int J Antimicrob Agents* 2012; 39(2): 105-14. [http://dx.doi.org/10.1016/j.ijantimicag.2011.10.004] [PMID: 22113193]
- [11] Mendes RE, Farrell DJ, Sader HS, Jones RN. Comprehensive assessment of tigecycline activity tested against a worldwide collection of *Acinetobacter* spp. (2005-2009). *Diagn Microbiol Infect Dis* 2010; 68(3): 307-11. [http://dx.doi.org/10.1016/j.diagmicrobio.2010.07.003] [PMID: 20955916]
- [12] Pournaras S, Koumaki V, Gennimata V, Kouskouni E, Tsakris A. *in vitro* Activity of tigecycline against *acinetobacter baumannii*: Global epidemiology and resistance mechanisms. In: Donelli G, Ed. *Adv Microbiol Infect Dis Public Health*. Cham: Springer International Publishing 2015; 897: pp. 1-14. [http://dx.doi.org/10.1007/5584\_2015\_5001]
- [13] Kuo SC, Chang SC, Wang HY, et al. Emergence of extensively drug-resistant *Acinetobacter baumannii* complex over 10 years: nationwide data from the Taiwan Surveillance of Antimicrobial Resistance (TSAR) program. *BMC Infect Dis* 2012; 12: 200. [http://dx.doi.org/10.1186/1471-2334-12-200] [PMID: 22929085]
- [14] Pachón-Ibáñez ME, Fernández-Cuenca F, Docobo-Pérez F, Pachón J, Pascual A. Prevention of rifampicin resistance in *Acinetobacter baumannii* in an experimental pneumonia murine model, using rifampicin associated with imipenem or sulbactam. *J Antimicrob Chemother* 2006; 58(3): 689-92. [http://dx.doi.org/10.1093/jac/dkl303] [PMID: 16870647]
- [15] Gauthier TP. Editorial commentary: rifampicin plus colistin in the era of extensively drug-resistant *Acinetobacter baumannii* infections. *Clin Infect Dis* 2013; 57(3): 359-61. [http://dx.doi.org/10.1093/cid/cit262] [PMID: 23616496]
- [16] Gales AC, Jones RN, Sader HS. Contemporary activity of colistin and polymyxin B against a worldwide collection of Gram-negative pathogens: results from the SENTRY Antimicrobial Surveillance Program (2006-09). *J Antimicrob Chemother* 2011; 66(9): 2070-4. [http://dx.doi.org/10.1093/jac/dkr239] [PMID: 21715434]
- [17] <http://ecdc.europa.eu/en/healthtopics/antimicrobial-resistance-and-consumption/antimicrobial-consumption/ESAC-Net/Pages/ESAC-Net.aspx> n.d.
- [18] Hsueh P-R, Chen W-H, Luh K-T. Relationships between antimicrobial use and antimicrobial resistance in Gram-negative bacteria causing nosocomial infections from 1991-2003 at a university hospital in Taiwan. *Int J Antimicrob Agents* 2005; 26(6): 463-72. [http://dx.doi.org/10.1016/j.ijantimicag.2005.08.016] [PMID: 16280243]
- [19] Loeffler JM, Garbino J, Lew D, Harbarth S, Rohner P. Antibiotic consumption, bacterial resistance and their correlation in a Swiss university hospital and its adult intensive care units. *Scand J Infect Dis* 2003; 35(11-12): 843-50. [http://dx.doi.org/10.1080/00365540310016646] [PMID: 14723360]
- [20] Munoz-Price LS, Rosa R, Castro JG, et al. Evaluating the impact of

antibiotic exposures as time-dependent variables on the acquisition of carbapenem-resistant *Acinetobacter baumannii*. *Crit Care Med* 2016; 44(10): e949-56. [http://dx.doi.org/10.1097/CCM.0000000000001848] [PMID: 27167999]

[21] Jaidane N, Naas T, Oueslati S, *et al.* Whole-genome sequencing of NDM-1-producing ST85 *Acinetobacter baumannii* isolates from Tunisia. *Int J Antimicrob Agents* 2018; 52(6): 916-21. [http://dx.doi.org/10.1016/j.ijantimicag.2018.05.017] [PMID: 29857033]

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