

In Vitro Study of Antimicrobial Activity of *Acalypha Indica* Linn. Extract

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Abstract: The objective for this research undertaken was to evaluate *Acalypha indica* extracts for their antimicrobial potential. Fresh plant samples were extracted via maceration in increasing polarity using petroleum ether, chloroform and methanol. Antimicrobial activity of the extracts was assessed using disc diffusion assay and minimum inhibitory concentration. It was observed that the crude methanol leaves, stem bark and chloroform stem bark extracts exhibit antimicrobial activity towards both *Staphylococcus aureus* bacteria and *Candida albicans* fungi. The inhibition zones recorded in millimeters by the chloroform stem bark extracts were 9.57±0.51 (200 mg/mL), 8.33±0.58 (100 mg/mL) and 7.33±0.58 (50 mg/mL) while methanol stem bark extract recorded inhibition zones at 13.67±0.58 (200 mg/mL), 10.00±1.00 (100 mg/mL) and 9.33±0.58 (50 mg/mL) against *S. aureus*. Antifungal activity was also detected against *C. albicans* as the chloroform stem bark extract produced inhibition zones of 14.33±0.58 (200 mg/mL), 13.33±0.58 (100 mg/mL) and 12.67±0.58 (50 mg/mL) while the methanol stem bark extract resulted in inhibition zones of 15.33±0.58 (200 mg/mL), 14.33±0.58 (100 mg/mL) and 13.0±0.00 (50 mg/mL). In the case of crude methanol leaves extract, the extract produced inhibition zones of 19±0.58 (200 mg/mL), 13±1.00 (100 mg/mL) and 10±1.00 (50 mg/mL) against *S. aureus* while against *C. albicans*, the extract recorded inhibition zones of 15±0.58 (200 mg/mL), 14±0.00 (100 mg/mL) and 13±0.00 (50 mg/mL). The chloroform stem bark crude extract developed a medium MIC value of 0.938 mg/mL while both methanol crude extracts produced a strong MIC value of 0.469 mg/mL during the minimum inhibition concentration screening against *Candida albicans*.

Keywords: Euphorbiaceae, *Acalypha indica*, Antimicrobial.

INTRODUCTION

The derivation of plants to produce drugs is basically bound to a principle which describes the potential of breaking plants down, isolating their active components and producing powerful drugs in standard form. Thus, to design drugs from medicinal plants of interest, it is crucial to research certain aspects of the plant itself. One of these areas is pharmacognosy. As such, the *Acalypha* is one of the genres that show a great potential in the world of scientific advancement due to its promising chemical and biological results.

Acalypha indica Linn. (*Euphorbiaceae*) also known as 'kucing galak' is widely distributed throughout tropical Africa and South Africa, India and Sri Lanka, as well as Yemen and Pakistan. It is a monoecious plant with a weedy nature, annual to sometimes short-lived perennial herb that can grow up to 1.5 to 2.5 m tall [1]. *A. indica* are popularly utilized as herbal medicine in the Indian Ocean islands as well as in India for its expectorant properties. The main parts that are usually exploited in medicine are the roots, leaf, stalk and flowers. The plants are emetic, expectorant, laxative and diuretic. It is useful in bronchitis, pneumonia

and pulmonary tuberculosis. *A. indica* also contains several alkaloids as well as hydrocyanic acid which can be deadly in the wrong dose, but young shoots are also eaten as vegetable [1].

The plant extract is considered to have antidiabetic in accordance to a research involving normal rats and alloxan induced diabetic rats [2]. *A. indica* is also known to possess respiratory effect on experimental animals such as bronchodilation and bronchial hyperreactivity. The end result of this particular study was that it has beneficial effect in asthma [3]. Another potential attribute of the *A. indica* is that it has antioxidant effect and should be implemented in a diet to control diseases where free radicals are involved.

MATERIALS AND METHODS

Preparation of Extracts

The *Acalypha indica* plant was collected from Tasik Chini, Pahang. It was authenticated by a plant botanist from Universiti Kebangsaan Malaysia. About 5 kg of finely powdered dry stem bark and leaves were soaked with petroleum ether, chloroform and methanol. The soaking process was repeated three times for each solvent. The solvent extracts were filtered and evaporated under vacuum at 55°C to yield the respective crude extract. The crude extracts were transferred into sample bottles and kept in refrigerator prior to use.

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Table 1. Antimicrobial Activity of Different Crude Solvent Extracts of *Acalypha Indica*

Extracts	Concentration (mg/ml)	Inhibition Zone (mm)				
		<i>S.a</i>	<i>P.a</i>	<i>E.c</i>	<i>C.a</i>	<i>T.t</i>
Chloroform (stem bark)	200	9.57 ± 0.51	-	-	14.33 ± 0.58	-
	100	8.33 ± 0.58	-	-	13.33 ± 0.58	-
	50	7.33 ± 0.58	-	-	12.67 ± 0.58	-
Methanol (stem bark)	200	13.67 ± 0.58	-	-	15.33 ± 0.58	-
	100	10.00 ± 1.00	-	-	14.33 ± 0.58	-
	50	9.33 ± 0.58	-	-	13.00 ± 0.00	-
Petroleum ether (stem bark)	200	-	-	-	-	-
	100	-	-	-	-	-
	50	-	-	-	-	-
Methanol (leaves)	200	19.00 ± 0.58	-	-	15.00 ± 0.58	-
	100	13.00 ± 1.00	-	-	14.00 ± 0.00	-
	50	10.00 ± 1.00	-	-	13.00 ± 0.00	-
Nystatin		-	-	-	25.00	32.00
Tetracycline		-	32.00	-	-	-
Ampicillin		18.00	-	25.00	-	-

No activity; S.a, *Staphylococcus aureus*; P.a, *Pseudomonas aeruginosa*; E.c, *Escherichia coli*; C.a, *Candida albicans*; T.t, *Trichophyton tonsurans*

Antimicrobial Screening

Microorganisms

The microorganisms used in this present study were bacteria (*Staphylococcus aureus*, *Pseudomonas aeruginosa* ATCC 27853, *Escherichia coli* ATCC 25922) and fungi (*Candida albicans*, *Trichophyton tonsurans* ATCC 28942). Bacteria and fungi were cultured maintained on Mueller Hinton Agar (Merck, Germany) and Sabouraud's dextrose agar, SDA (Difco, USA) respectively at 25°C.

Preparation of Standardized Microbial Suspension

Stock cultures were prepared and maintained at 4°C in Mueller Hinton Broth (MHB) and Sabouraud Dextrose Broth (SDB) cultures. Standardized microbial inoculums was adjusted to 0.5 McFarland and then diluted to 1 x 10⁸ CFU/mL [4].

Disk Diffusion Method

An antimicrobial assay was performed by using the disc diffusion agar method [5]. Petri dishes were first inoculated with microbes by pipetting the microbial suspension onto the agar. The standardized microbial suspension was applied onto the solidified agars by using sterile cotton swabs and allowed to dry for 10 minutes. Crude extract impregnated discs were aseptically transferred on the inoculated agar plates and left to be incubated for 24 hrs to 7 days. The clear zones of inhibition around the test crude extract disc were measured for any indication of antimicrobial activity. Nystatin, ampicillin and tetracycline impregnated discs were used as standard reference or positive controls and the

solvents were used as negative controls. All assays were carried out in triplicate.

Minimum Inhibitory Concentration (MIC)

The MIC determination of the crude extracts was referred and modified from [4, 6]. 30 µL samples were diluted by two fold serial dilution with its suitable solvent in the wells of microtiter plate. 170 µL of the prepared microbial culture was added to each well to give a final volume of 200 µL with final concentrations of each well ranging from 50 mg/mL to 0.024 mg/mL. The microtitre plates were then incubated at 30°C to 37°C for 24 to 48 hours, with their upper surface covered and sealed with parafilms. After incubation, 20 µL of MTT solution was pipette into the wells for indication of growth. The lowest concentration that did not show any visible growth was recorded as the MIC of that extract for the tested microbial species. All the MIC experimentations were performed in triplicate.

RESULTS AND DISCUSSION

Antimicrobial Activity

The results for the antimicrobial activity test of different crude solvent extracts of *A. indica* are displayed in Table 1. The growth of both *Staphylococcus aureus*, and *Candida albicans* can be seen to be inhibited by chloroform stem bark, methanol stem bark and leaves extract in a concentration dependent manner but not inhibited by the petroleum stem bark extract. While *Pseudomonas aeruginosa*, *Escherichia coli* and *Tricophyton tonsurans* were shown to be resistant towards all type of extracts that was studied in this research. *C. albicans* seems to be more susceptible

Table 2. MIC Determination of *Acalypha Indica* Active Crude Extracts Against *Candida Albicans*

Extracts	MIC (mg/mL)
Chloroform (stembark)	0.938
Methanol (stembark)	0.469
Methanol (leaves)	0.469

MIC: Minimum Inhibitory Concentration

towards *A. indica* stembark when compared to *S. aureus* while the *S. aureus* was more susceptible towards the leaves extract when compared to *C. albicans*. All microbes were susceptible to its respective positive control. Both *S. aureus* and *E.coli* were susceptible to ampicillin while *P. aeruginosa* was susceptible to its positive control, tetracycline. In the terms of the dermatophytes group, both *C. albicans* and *T. tonsurans* were susceptible to nystatin.

According to the study, all tested microorganisms were resistant towards petroleum ether crude extract of *A. indica* stembark which could indicate that the extract does not contain active compounds that could be responsible for antimicrobial activities. It is also noted that all Gram-negative bacteria were more resistant towards all the extracts tested compared to the Gram-positive bacteria. According to a study conducted by [7], *A. indica* extracts produced active results against all the Gram-positive bacteria tested while one of the Gram-negative bacteria, *Pseudomonas aeruginosa* was only susceptible towards the extracts at a higher concentration. This result could be attributed to the difference in wall compositions that exist in both Gram-positive and Gram-negative bacteria. While the Gram-negative bacteria possess wall that consists of lipopolysaccharide layer along with proteins and phospholipids that may impede the entry of active compounds of *A. indica* crude extracts, the Gram-positive bacteria contains a very active area of cell metabolism called periplasmic space that carry many digestive enzymes and transport proteins which could attribute to the susceptibility of the microorganisms.

In the aspect of its antifungal properties, the plants extract prove to be active against fungi *Candida albicans* in increasing concentration compared to *Tricophyton tonsurans*. Previous antifungal studies conducted by [8, 9] proved that methanol extract of *A. indica* possess antifungal activity against *C. albicans*. However, their studies involve the use of the whole plant and not a specific part of the plant. Another study from [10] has also showed that the methanol leaves extract of *A. indica* is most active against *C. albicans* compared to other extracts tested as it resulted in the highest inhibition zones against fungi. *C. albicans* was also proven to be susceptible towards chloroform and ethanol extracts compared to other fungi in a study conducted by [11].

Minimum Inhibitory Concentration

As it is showed in Table 2, both methanol stembark and leaves extract recorded a strong MIC value of 0.469 mg/mL in which the growth of *C. albicans* was inhibited while the chloroform stembark extract was recorded to possess a medium MIC value of 0.938 mg/mL.

The antimicrobial activity could be attributed to the presence of alkaloids, tannins and saponins in the *A. indica*

leaves extract according to [12]. This was also supported by [13] with additional compounds that were discovered such as steroids, cardiac glycosides and phenols. One study by [14] suggested that the antibacterial activity of *Acalypha indica* leaves extract could be attributed to the active compounds of alkaloids and tannins. According to a study conducted by [15], the presence of bioactive compounds such as alkaloids, tannins, steroids, saponins, flavanoids, glycosides and phenolic compounds was also detected during its phytochemical testing.

CONCLUSION

In conclusion, the present study indicates that the crude extract of *A. indica* stembark and leaves possess some antimicrobial activities against certain pathogenic microbes. The extracts of methanol and chloroform proved to be a good antifungal agent against *Candida albicans*. Nevertheless, future studies in regards to its bioactive compound should be done in order to identify the compound that is responsible for its antimicrobial activities.

CONFLICT OF INTEREST

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