



The Open AIDS Journal

Content list available at: www.benthamopen.com/TOAIDJ/

DOI: 10.2174/1874613601711010024



RESEARCH ARTICLE

Plasma IP-10 Concentrations Correlate Positively with Viraemia and Inversely with CD4 Counts in Untreated HIV Infection

Kudakwashe Mhandire^{1,4,*}, §, Tommy Mlambo^{2,*}, Lynn Sodai Zijenah², Kerina Duri², Kudzaishe Mateveke⁶, Mqondisi Tshabalala², Doreen Zvipo Mhandire^{1,4}, Cuthbert Musarurwa¹, Petronella Taonga Wekare³, Lovemore Ronald Mazengera², Hilda Tendisa Matarira¹ and Babill Stray-Pedersen^{4,5}

¹Department of Chemical Pathology, University of Zimbabwe, Harare, Zimbabwe

²Department of Immunology, University of Zimbabwe, Harare, Zimbabwe

³Medical Laboratory Sciences, University of Zimbabwe, College of Health Sciences, Harare, Zimbabwe

⁴Letten Foundation Research House, Harare, Zimbabwe

⁵Institute of Clinical Medicine, University of Oslo and Womens' Clinic, Rikshospitalet, University Hospital, Oslo, Norway

⁶Research Support Centre, University of Zimbabwe, College of Health Sciences, Harare, Zimbabwe

Received: November 21, 2016

Revised: January 31, 2017

Accepted: February 21, 2017

Abstract:

Background:

Chronic immune activation is a feature of HIV infection associated with accelerated HIV disease progression. There is conflicting data on the association of biomarkers of immune activation with traditional markers of HIV disease progression; CD4 counts and viral load (VL).

Objective:

The study aimed to determine the association of biomarkers of immune activation; interferon (IFN)- γ -induced protein 10 (IP-10) and soluble cluster of differentiation 14 (sCD14) in chronic HIV infection with traditional markers of HIV disease progression.

Methods:

We collected demographic data, enumerated CD4 counts and quantified VL in 183 antiretroviral therapy (ART)-naive adults with chronic HIV infection. Plasma concentrations of IP-10 and sCD14 were quantified in the ART-naive adults with chronic HIV infection and 75 HIV-uninfected controls.

Results:

IP-10 concentrations were significantly higher in the HIV-infected group (median; 257.40pg/ml, IQR; 174.08-376.32) than in the HIV-uninfected (median; 86.19pg/ml, IQR; 67.70-116.39) ($P < 0.001$). Similarly, sCD14 concentrations were significantly higher in the HIV-infected (median; 1.45 μ g/ml, IQR; 1.02-2.16) group than in the controls (median; 0.89 μ g/ml, IQR; 0.74-1.18) ($P < 0.001$). High \log_{10} IP-10 concentrations were positively correlated with high \log_{10} viral loads (Spearman's correlation coefficient [R]=0.21, $P = 0.003$) and inversely correlated with low CD4 counts ($R = -0.19$, $P = 0.011$). In contrast, \log_{10} sCD14 was not significantly associated with either \log_{10} viral loads ($R = 0.03$, $P = 0.707$) nor CD4 count ($R = -0.04$, $P = 0.568$).

* These authors contributed equally

§ Address correspondence to this author at the Department of Chemical Pathology, College of Health Sciences, University of Zimbabwe, Mazowe Street, Harare; Cell: +263 773 578 505; Emails: mhandiremhandire@gmail.com; doreenduri@gmail.com

Conclusion:

We conclude that plasma sCD14 and IP-10 were elevated in the HIV-infected patients compared to HIV-uninfected individuals possibly due to on-going immune activation. In addition, plasma high concentrations of IP-10 but not sCD14 concentrations are associated with high VL and low CD4 count.

Keywords: Immune activation, sCD14, IP-10, CD4 count, Viral load, Zimbabwean.

INTRODUCTION

Chronic immune activation is a feature of HIV infection that is associated with accelerated HIV disease progression [1]. Chronic immune activation is the persistent stimulation of the immune system characterized by elevation of markers of cell activation, cytokines, acute phase proteins and pathological changes in composition of immune cells [2]. HIV replication, co-infections, depletion of gut mucosal integrity and consequent microbial translocation are among the main mechanisms of immune activation in HIV infection [3]. In non-human primates that do not progress to AIDS despite high simian immunodeficiency virus (SIV) load, immune activation that follows infection is transient and often successfully reduced to pre-infection levels without intervention [2]. In contrast, immune activation tends to be chronic among non-human primates SIV and human HIV disease progressors, suggesting that immune activation may be the major vehicle behind disease progression. However, the dynamics of immune activation in HIV infection are not yet fully understood.

ART down-regulates immune activation but does not eradicate it [4]. Thus, immune activation is reported to persist despite successful viral suppression [5] leading to immune depletion and non-AIDS complications such as cardiovascular disease and kidney disease [6, 7]. Efforts to identify biomarkers that correlate with immunological, virological and clinical disease progression have been informative but contradictory [2, 8]. Among the plasma biomarkers documented to date, IP-10, a chemokine secreted by leukocytes, monocytes, epithelia, stromal cells and keratinocytes following induction by IFN- γ [9], and sCD14, a cell receptor released from monocytes stimulated by lipopolysaccharides during microbial translocation [8], hold promise for use as markers of immune activation and possibly disease progression. However, results from studies comparing IP-10 and sCD14 levels in the HIV- infected individuals and uninfected individuals are contradictory [1, 7, 10 - 12]. Research findings on the association between IP-10 and sCD14 concentrations and VL and/or CD4 counts are also controversial [1, 10, 12 - 14]. Persistence of immune activation despite viral suppression increases the risk of immunological depletion and development of non-AIDS events, hence monitoring of immune activation may be important in HIV management.

In this study, we described differences in the markers of immune activation, IP-10 and sCD14 between ART-naïve HIV-infected and HIV-uninfected adult Zimbabweans. In addition, we correlate IP-10 and sCD14 concentrations with VL and CD4 count to identify their role in HIV disease progression. ART-naïve HIV-infected individuals present us with an opportunity to determine the interaction between immune activation and virological/immunological status. We found both IP-10 and sCD14 elevated in the HIV-infected group compared to HIV-uninfected controls. However, only high concentrations of IP-10 were positively correlated with high VL and inversely correlated with low CD4 count.

METHODS**Study Participants**

HIV-infected, ART naïve Black Zimbabweans aged ≥ 18 years with CD4 count >350 cells/ μ L were enrolled into the Immunological and Virological Investigations of HIV-infected individuals with CD4 counts >350 cells/ μ L (IVI HIV) Study at the Parirenyatwa Groups of Hospitals Opportunistic Infections Clinic in Harare, Zimbabwe. Written informed consent was obtained from each participant. The study was conducted from 2010 to 2013. During this period, ART was provided only to patients with CD4 counts <350 cells/ μ L under the Zimbabwe National ART Program. At enrollment, a questionnaire was administered to collect demographic data and medical history, whole blood was collected in ethylene diaminetetraacetic acid (EDTA) anti-coagulant for CD4 counts and plasma isolation within six hours of collection. The plasma was immediately stored at -80°C until further use. Plasma HIV RNA load (VL) was quantified using the archived plasma collected at enrolment. The median time since the diagnosis of HIV infection in the cohort was 6.3 years, hence the patients were considered as chronically HIV-infected.

In this cross sectional study, we quantified IP-10 and sCD14 in the 183 HIV-infected individuals using archived plasma collected at enrolment. We then determined the association of the concentrations of these two biomarkers with

viral load and CD4 counts. We also collected demographic information and EDTA whole blood from 75 unmatched HIV-uninfected Black Zimbabwean blood donors aged ≥ 18 years at the National Blood Transfusion Services (NBTS) of Zimbabwe in Harare who provided written informed consent to participate in the study. The study was approved by the Medical Research Council of Zimbabwe (MRCZ/A/2016).

Measurement of Plasma IP-10 and sCD14 Concentrations

IP10 and sCD14 concentrations were measured in archived plasma from HIV-infected cases and HIV-uninfected controls using Quantikine[®] enzyme linked immune-sorbent assays (R&D Systems, Minneapolis, USA) following the manufacturer's instructions. Each assay was run in duplicate. The lower detection limits of the assays were 0.125 μ g/mL and 1.67pg/mL for sCD14 and IP-10, respectively.

Measurement of CD4 Counts and Plasma Viral Load

CD4+ T lymphocytes were enumerated in EDTA anti coagulated whole blood within 6 hours of blood collection using the Partec platform (Sysmex, Germany) the following manufacturer's instructions. HIV RNA copies were estimated in archived plasma using the Cavid Exavir[™] viral load version 3 assay (Cavidi AB, Uppsala Science Park, Sweden) following the manufacturer's instruction. The VL assay has a lower detection limit of 200 copies/ml.

Statistical Analyses

Statistical analyses were done using Stata version 13.0 (StataCorp, Texas, USA) and GraphPad Prism 7 (GraphPad Software Inc, Carlsifornia, USA). Demographic and clinical characteristics were compared between HIV-infected and HIV-uninfected groups using the Student's T-test for parametric variables, Mann-Whitney sum rank for non-parametric variables, and Chi-squared test or Fisher's exact tests for categorical data. A value of $P < 0.05$ was considered statistically significant. P-values were adjusted for multivariate comparisons using Bonferroni correction method. For correlation analysis, a p-value < 0.025 was considered significant. Spearman's correlation coefficient (R) was used to infer association between biomarkers and traditional markers of HIV disease progression (VL and CD4 count). A multivariate linear regression analysis was used to predict the effect of age, gender, weight and co-morbidities on IP-10 and sCD14 concentrations in the HIV-infected group and that of age, gender and weight in the HIV-uninfected group.

RESULTS

Demographic and Clinical Characteristics of the Participants

The demographic characteristics of the participants are shown in Table (1). The mean weight of the HIV-infected participants was significantly lower than that of the HIV-uninfected participants. Females were overrepresented in the HIV-infected group than in the uninfected group. Mean age was not significantly different between the HIV-infected and uninfected participants. Twenty-six (14%) participants had VL ≤ 200 cpm; 55 (30%) had VL > 200 cpm but ≤ 1000 cpm, 64 (35%) > 1000 cpm but ≤ 10000 cpm whilst the remaining 38 (21%) had VL > 10000 cpm. The median CD4 count was 435 cells/ μ l, IQR: 388-556. One hundred and twenty-four (68%) participants had CD4 counts < 500 cells/ μ l (median: 407 cells/ μ l, IQR: 309-496) while 59 (32%) had ≥ 500 cells/ μ l (median: 613 cells/ μ l, IQR: 503-923). Fifty-one (28%) of the HIV-infected individuals had at least one self reported co-morbidity, of whom 15 (30%), had two or more co-morbidities.

Table 1. Demographic and clinical characteristic of participants.

Characteristic	HIV-infected (n=183)	HIV-uninfected (n=75)	p-value
Female n (%)	136 (0.74)	29 (0.39)	$< 0.001^*$
Mean age in years \pm SD	34.7 \pm 9.1	36.9 \pm 11.8	0.08 ^a
Mean weight in kg \pm SD	66.8 \pm 0.98	75.6 \pm 9.8	$< 0.001^a$
Median CD4 count in cells/ μ l (IQR)	435 (388-556)	NA	NA
Median plasma viral load in copies/mL (IQR)	1444 (519-7858)	NA	NA
≥ 1 current co-morbidities	51(0.28)	NA	NA

SD: standard deviation; IQR: interquartile range; NA: non-applicable; *Chi-squared test; ^aStudent's T-test.

Plasma IP-10 and sCD14 Concentrations in HIV-Infected and HIV-uninfected Groups

We compared IP-10 and sCD14 plasma concentrations between HIV-infected cases and HIV-uninfected controls to

determine association between the concentrations of these biomarkers with HIV status Fig. (1). IP-10 plasma concentrations were significantly higher in the HIV-infected group (median; 257.40pg/ml, IQR; 174.08-376.32) than in the HIV-uninfected (median; 86.19pg/ml, IQR; 67.70-116.39) ($P<0.001$). Similarly, sCD14 plasma concentrations were significantly higher in the HIV-infected (median; 1.45 μ g/ml, IQR; 1.02-2.16) group than the HIV-uninfected (median 0.89 μ g/ml, IQR; 0.74-1.18) ($P<0.001$). There was no significant correlation between IP-10 and sCD14 concentrations in either the HIV-infected ($R = 0.09$, $P=0.248$) or the HIV-uninfected groups ($R= 0.05$, $P=0.662$). We used a linear regression analysis to account the possible effect of age, gender and weight between the HIV-infected and uninfected groups on IP-10 and sCD14 concentrations. Age, gender and weight were not significantly associated with either IP-10 or sCD14 plasma concentrations in either the HIV-infected or HIV-uninfected groups (Supplementary Table 1). Also, having an HIV co-morbidity was not associated with either IP-10 or sCD14 concentrations.

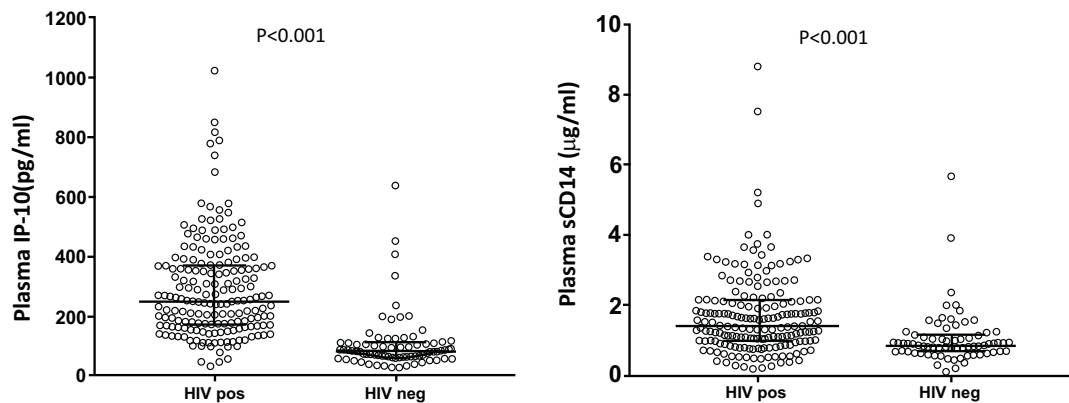


Fig. (1). Comparison of plasma IP-10 and sCD14 levels between HIV-infected and HIV-uninfected groups. Mann-Whitney rank sum test was used to compare difference between medians of the two groups. Both IP-10 and sCD14 were significantly higher in the HIV-infected group than the HIV-uninfected group.

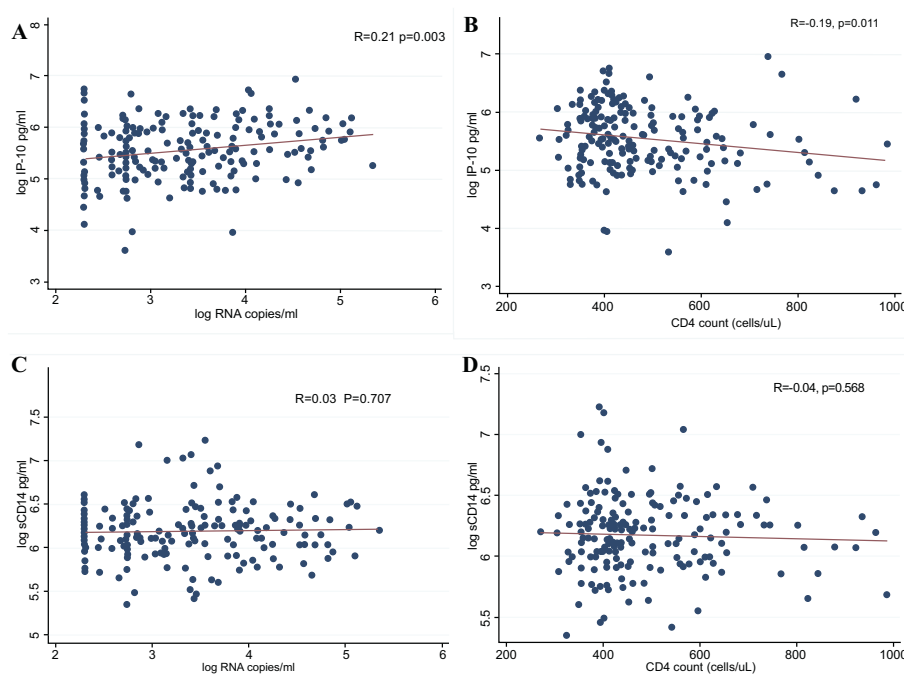


Fig. (2). Correlation of IP-10 and sCD14 with viral loads and CD4 counts. Spearman's correlation coefficient was used to determine strength of association between biomarkers (\log_{10} IP-10 and \log_{10} sCD14) and markers of disease progression (\log_{10} viral loads and CD4 counts). \log_{10} IP-10 levels positively correlate with \log_{10} viral load (A) and negatively with CD4 count (B). CD14 levels were not significantly associated with neither \log_{10} viral load (C) nor CD4 count (D).

Correlation of IP-10 and sCD14 with VL and CD4 Counts.

We then determined the correlation between immune activation biomarkers (IP-10 and sCD14) and traditional markers of HIV disease progression (VL and CD4 counts). As shown in Fig. (2), high log₁₀ IP-10 concentrations were positively correlated with high log₁₀ viral loads (R=0.21, P=0.003) and inversely correlated with low CD4 counts (R=-0.19, P=0.011). However, log₁₀ sCD14 was not associated with either log₁₀ viral load (R=0.03, P=0.707) or CD4 count (R=-0.04, P=0.568).

Supplementary Table 1. Summary of simple linear regression analyses of demographic variables predicting plasma IP-10 and sCD14 concentrations in the HIV-infected and HIV-uninfected groups.

	Coefficient	Standard Error	P-value
HIV-infected			
<i>For Log IP-10</i>			
Gender	0.153	0.101	0.131
Age	0.005	0.005	0.250
Weight	0.002	0.003	0.554
Co-morbidity	-0.078	0.098	0.423
<i>For log sCD14</i>			
Gender	0.074	0.056	0.196
Age	0.002	0.002	0.575
Weight	0.002	0.003	0.947
Co-morbidity	-0.003	0.055	0.963
HIV-uninfected			
<i>For Log IP-10</i>			
Gender	-0.098	0.148	0.508
Age	-0.007	0.008	0.318
Weight	0.005	0.009	0.594
<i>For log sCD14</i>			
Gender	-0.074	0.057	0.196
Age	-0.001	0.002	0.575
Weight	0.001	0.003	0.947

DISCUSSION

Immune activation is an important determinant of HIV disease course. Here, we characterized biomarkers of immune activation (IP-10 and sCD14) in HIV-infected patients and HIV-uninfected individuals in Zimbabwe. The plasma IP-10 and sCD14 concentrations were elevated in the HIV-infected group compared to the HIV-uninfected group. Among the HIV-infected individuals, high plasma IP-10 concentrations were positively correlated with high VL and inversely correlated with low CD4 count. Surprisingly, there was no significant correlation of plasma sCD14 with either plasma VL or CD4 count. Our findings support the existence of chronic immune activation in chronic HIV-infection as previously described [2, 15] and the positive correlation of high IP-10 concentrations with high VL and inverse correlation with low CD4 count. To our knowledge, this is the first study to determine IP-10 and sCD14 plasma concentrations as biomarkers of immune activation and their association with traditional markers of HIV disease progression among Zimbabweans infected predominantly with HIV-1 subtype C.

Our findings are consistent with earlier reports of significant elevation of plasma IP-10 [1, 10, 14, 16] and sCD14 [7, 16, 17] in HIV-infected adults compared to HIV-uninfected controls. IP-10 is among the earliest cytokines to become elevated following infection with HIV [18]. Plasmacytoid dendritic cells (pDC) recruited to a site of infection produce IFNs that induce IP-10 production [14] by a range of cells that includes leukocytes, monocytes and neutrophils [13]. While this is an active mechanism for IP-10 production in acute HIV infection stage, our study participants had chronic HIV infection. In chronic HIV infection, prolonged production of IFN- γ in lymphoid organs is responsible for sustained IP-10 elevation [19]. Despite marked reduction in plasma IP-10 from acute HIV infection stage concentrations, chronic HIV infection concentrations remain higher than pre-infection concentrations even in patients commenced on ART [20]. IP-10 is a ligand of the CXCR3 receptor which is not known to function as a co-receptor for HIV [21], however, CXCR3+ CD4+ T lymphocytes show high HIV infection rate [22]. Furthermore, IP-10 increases

susceptibility of nascent naïve CD4+ T lymphocytes to HIV infection [23], hence IP-10 enhances the establishment of HIV reservoirs [12].

Elevated concentrations of sCD14 in HIV-infected compared to HIV-uninfected individuals are largely attributed to monocyte activation by bacterial lipopolysaccharides (LPS) leaking into circulation through compromised gut mucosa, a process known as microbial translocation [8]. CD14 molecules are expressed on monocytes where they act as co-receptors for LPS and other microbial components [24]. When LPS bind CD14 receptors, monocytes are activated to release CD14 in a soluble form, sCD14 [24] hence the rise in plasma sCD14. Higher concentrations of sCD14 in HIV-infected individuals compared to the HIV-uninfected have also been reported in other studies [1, 25, 26]. Microbial translocation as determined by elevated sCD14 has been linked with increased mortality among HIV-infected individuals [25] and individuals on haemodialysis [27]. Importantly, sCD14 concentrations are also elevated in hepatitis C virus infection [28], tuberculosis [29], inflammatory disorders such as rheumatoid arthritis and systemic lupus erythematosus [30] and in HIV associated accelerated aging [31]. The diversity of conditions in which sCD14 levels are reported elevated perhaps limits the specificity of the biomarker to any of them thereby limiting its utility as a diagnostic or prognostic marker. Interestingly, although, high levels of sCD14 were mostly associated with poor prognosis in these conditions. In the current study, we did not observe any association with traditional markers of HIV disease progression.

In sharp contrast, we found IP-10 concentrations positively and negatively correlated to VL and CD4 counts, respectively. These findings corroborate reports from earlier studies [1, 8 - 11]. Whilst our study focused on IP-10 and sCD14 because of their link to HIV status and markers of disease progression described in literature, some studies have profiled between 15 and 28 proteins and singled out elevated IP-10 concentrations to positively correlate with high VL and inversely correlate with low CD4 counts [1, 10, 12 - 14]. Determination of IP-10 concentrations may have a prognostic value in early HIV infection as elevated IP-10 concentrations have been reported to predict disease progression and up to 30% variation in CD4 count set point [10, 19]. However, it remains unclear whether measurement of plasma IP-10 may be useful in HIV patient management during the acute or chronic stages of infection. In chronic HIV infection, immune activation leads to poor prognosis and eventual death [15, 32] hence we speculate that biomarkers such as IP-10 whose elevation may favor viral replication may guide anti-immune activation treatment in future.

IP-10 stimulates viral replication in infected cells [21] consequently increasing viraemia. In the vaginal mucosa, an important site in HIV pathogenesis, IP-10 is associated with increased HIV shedding [33] that facilitates infection of new targets. Persistent elevation of IP-10 has been linked to immunological treatment failure among patients on cART despite suppressed viraemia [20]. Immune activation therefore threatens the gains of ART hence treatment options aimed at reducing immune activation have been explored, but with mixed results [15]. Importantly, though statistically significant, the correlations between IP-10 and VL and CD4 counts in our study were fairly weak hence our findings need to be interpreted with care. Our cross sectional approach may have precluded investigations on the influence of IP-10 and sCD14 on HIV disease progression. A longitudinal study involving multiple time point measurements of IP-10 and sCD14 against VL and CD4 counts from acute infection stage to chronic stage is therefore recommended. However, accurate timing of HIV infection remains a challenge because patients tend to seek treatment when they have reached the chronic stage. Furthermore, differences between HIV-infected patients and voluntary blood donors may have introduced some selection bias and other undetectable confounding to our findings.

CONCLUSION

IP-10 and sCD14 concentrations were elevated in the HIV-infected patients compared to HIV-uninfected individuals possibly due to on-going immune activation. In addition, plasma high concentrations of IP-10 but not sCD14 concentrations were associated with high VL and low CD4 count. Thus, IP-10 secretion may be associated with HIV pathogenesis and immune depletion. Prospective studies with larger sample sizes are required to establish causation.

CONFLICT OF INTEREST

The author confirms that this article content has no conflict of interest.

ACKNOWLEDGEMENTS

We would like to thank the Letten Foundation Research House, Norway and the European and Developing

Countries Clinical Trials Partnership for funding. We are also indebted to the Parirenyatwa Groups of Hospital, University of Zimbabwe College of Health Sciences and National Blood Service Zimbabwe for facilitating the research activities. Finally, special thanks to our study participants.

REFERENCES

- [1] Kamat A, Misra V, Cassol E, *et al.* A plasma biomarker signature of immune activation in HIV patients on antiretroviral therapy. *PLoS One* 2012; 7(2): e30881. [<http://dx.doi.org/10.1371/journal.pone.0030881>] [PMID: 22363505]
- [2] Paiardini M, Müller-Trutwin M. HIV-associated chronic immune activation. *Immunol Rev* 2013; 254(1): 78-101. [<http://dx.doi.org/10.1111/imr.12079>] [PMID: 23772616]
- [3] Rajasuriar R, Khoury G, Kamarulzaman A, French MA, Cameron PU, Lewin SR. Persistent immune activation in chronic HIV infection: do any interventions work? *AIDS* 2013; 27(8): 1199-208. [<http://dx.doi.org/10.1097/QAD.0b013e32835ecb8b>] [PMID: 23324661]
- [4] Lederman MM, Calabrese L, Funderburg NT, *et al.* Immunologic failure despite suppressive antiretroviral therapy is related to activation and turnover of memory CD4 cells. *J Infect Dis* 2011; 204(8): 1217-26. [<http://dx.doi.org/10.1093/infdis/jir507>] [PMID: 21917895]
- [5] French MA, King MS, Tschampa JM, da Silva BA, Landay AL. Serum immune activation markers are persistently increased in patients with HIV infection after 6 years of antiretroviral therapy despite suppression of viral replication and reconstitution of CD4+ T cells. *J Infect Dis* 2009; 200(8): 1212-5. [<http://dx.doi.org/10.1086/605890>] [PMID: 19728788]
- [6] Poesen R, Ramezani A, Claes K, *et al.* Associations of soluble CD14 and endotoxin with mortality, cardiovascular disease, and progression of kidney disease among patients with CKD. *Clin J Am Soc Nephrol* 2015; 10(9): 1525-33. [<http://dx.doi.org/10.2215/CJN.03100315>] [PMID: 26153127]
- [7] Castley A, Williams L, James I, Guelfi G, Berry C, Nolan D. Plasma CXCL10, sCD163 and sCD14 levels have distinct associations with antiretroviral treatment and cardiovascular disease risk factors. *PLoS One* 2016; 11(6): e0158169. [<http://dx.doi.org/10.1371/journal.pone.0158169>] [PMID: 27355513]
- [8] Leeansyah E, Malone DF, Anthony DD, Sandberg JK. Soluble biomarkers of HIV transmission, disease progression and comorbidities. *Curr Opin HIV AIDS* 2013; 8(2): 117-24. [<http://dx.doi.org/10.1097/COH.0b013e32835e7134>] [PMID: 23274365]
- [9] Liu M, Guo S, Hibbert JM, *et al.* CXCL10/IP-10 in infectious diseases pathogenesis and potential therapeutic implications. *Cytokine Growth Factor Rev* 2011; 22(3): 121-30. [PMID: 21802343]
- [10] Liovat AS, Rey-Cuillé MA, Lécuroux C, *et al.* Acute plasma biomarkers of T cell activation set-point levels and of disease progression in HIV-1 infection. *PLoS One* 2012; 7(10): e46143. [<http://dx.doi.org/10.1371/journal.pone.0046143>] [PMID: 23056251]
- [11] Castley A, Berry C, French M, Fernandez S, Krueger R, Nolan D. Elevated plasma soluble CD14 and skewed CD16+ monocyte distribution persist despite normalisation of soluble CD163 and CXCL10 by effective HIV therapy: a changing paradigm for routine HIV laboratory monitoring? *PLoS One* 2014; 9(12): e115226. [<http://dx.doi.org/10.1371/journal.pone.0115226>] [PMID: 25544986]
- [12] Ploquin MJ, Madec Y, Casrouge A, *et al.* Elevated basal pre-infection CXCL10 in plasma and in the small intestine after infection are associated with more rapid HIV/SIV disease onset. *PLoS Pathog* 2016; 12(8): e1005774. [<http://dx.doi.org/10.1371/journal.ppat.1005774>] [PMID: 27509048]
- [13] Jiao Y, Zhang T, Wang R, *et al.* Plasma IP-10 is associated with rapid disease progression in early HIV-1 infection. *Viral Immunol* 2012; 25(4): 333-7. [<http://dx.doi.org/10.1089/vim.2012.0011>] [PMID: 22788418]
- [14] Lee S, Chung YS, Yoon CH, *et al.* Interferon-inducible protein 10 (IP-10) is associated with viremia of early HIV-1 infection in Korean patients. *J Med Virol* 2015; 87(5): 782-9. [<http://dx.doi.org/10.1002/jmv.24026>] [PMID: 25678246]
- [15] Hatano H. Immune activation and HIV persistence: considerations for novel therapeutic interventions. *Curr Opin HIV AIDS* 2013; 8(3): 211-6. [<http://dx.doi.org/10.1097/COH.0b013e32835f9788>] [PMID: 23454864]
- [16] Makhubele TG, Steel HC, Anderson R, van Dyk G, Theron AJ, Rossouw TM. Systemic immune activation profiles of HIV-1 subtype C-infected children and their mothers. *Mediators Inflamm* 2016; 2016: 1-7.
- [17] Krebs SJ, Slike BM, Sithinamsuwan P, *et al.* Sex differences in soluble markers vary before and after the initiation of antiretroviral therapy in chronically HIV-infected individuals. *AIDS* 2016; 30(10): 1533-42. [<http://dx.doi.org/10.1097/QAD.0000000000001096>] [PMID: 26990631]
- [18] Stacey AR, Norris PJ, Qin L, *et al.* Induction of a striking systemic cytokine cascade prior to peak viremia in acute human immunodeficiency

- virus type 1 infection, in contrast to more modest and delayed responses in acute hepatitis B and C virus infections. *J Virol* 2009; 83(8): 3719-33.
[<http://dx.doi.org/10.1128/JVI.01844-08>] [PMID: 19176632]
- [19] Noel N, Lerolle N, Lécuroux C, *et al.* Immunologic and virologic progression in HIV controllers: the role of viral Blips and immune activation in the ANRS CO21 CODEX study. *PLoS One* 2015; 10(7): e0131922.
[<http://dx.doi.org/10.1371/journal.pone.0131922>] [PMID: 26146823]
- [20] Stylianou E, Aukrust P, Bendtzen K, Müller F, Frøland SS. Interferons and interferon (IFN)-inducible protein 10 during highly active anti-retroviral therapy (HAART)-possible immunosuppressive role of IFN-alpha in HIV infection. *Clin Exp Immunol* 2000; 119(3): 479-85.
[<http://dx.doi.org/10.1046/j.1365-2249.2000.01144.x>] [PMID: 10691920]
- [21] Lane BR, King SR, Bock PJ, Strieter RM, Coffey MJ, Markovitz DM. The C-X-C chemokine IP-10 stimulates HIV-1 replication. *Virology* 2003; 307(1): 122-34.
[[http://dx.doi.org/10.1016/S0042-6822\(02\)00045-4](http://dx.doi.org/10.1016/S0042-6822(02)00045-4)] [PMID: 12667820]
- [22] Gosselin A, Monteiro P, Chomont N, *et al.* Peripheral blood CCR4+CCR6+ and CXCR3+CCR6+CD4+ T cells are highly permissive to HIV-1 infection. *J Immunol* 2010; 184(3): 1604-16.
[<http://dx.doi.org/10.4049/jimmunol.0903058>] [PMID: 20042588]
- [23] Cameron PU, Saleh S, Sallmann G, *et al.* Establishment of HIV-1 latency in resting CD4+ T cells depends on chemokine-induced changes in the actin cytoskeleton. *Proc Natl Acad Sci USA* 2010; 107(39): 16934-9.
[<http://dx.doi.org/10.1073/pnas.1002894107>] [PMID: 20837531]
- [24] Shive CL, Jiang W, Anthony DD, Lederman MM. Soluble CD14 is a nonspecific marker of monocyte activation. *AIDS* 2015; 29(10): 1263-5.
[<http://dx.doi.org/10.1097/QAD.0000000000000735>] [PMID: 26035325]
- [25] Sandler NG, Wand H, Roque A, *et al.* Plasma levels of soluble CD14 independently predict mortality in HIV infection. *J Infect Dis* 2011; 203(6): 780-90. [<http://dx.doi.org/10.1093/infdis/jiq118>]. [PMID: 21252259].
[<http://dx.doi.org/10.1093/infdis/jiq118>] [PMID: 21252259]
- [26] Méndez-Lagares G, Romero-Sánchez MC, Ruiz-Mateos E, *et al.* Long-term suppressive combined antiretroviral treatment does not normalize the serum level of soluble CD14. *J Infect Dis* 2013; 207(8): 1221-5.
[<http://dx.doi.org/10.1093/infdis/jit025>] [PMID: 23322858]
- [27] Raj DS, Shah VO, Rambod M, Kovcsy CP, Kalantar-Zadeh K. Association of soluble endotoxin receptor CD14 and mortality among patients undergoing hemodialysis. *Am J Kidney Dis* 2009; 54(6): 1062-71.
[<http://dx.doi.org/10.1053/j.ajkd.2009.06.028>] [PMID: 19699018]
- [28] Medhat E, Salama H, Fouad H, *et al.* Serum soluble CD14 in Egyptian patients with chronic hepatitis C: its relationship to disease progression and response to treatment. *J Interferon Cytokine Res* 2015; 35(7): 563-8.
[<http://dx.doi.org/10.1089/jir.2014.0155>] [PMID: 25811314]
- [29] Alavi-Naini R, Salimi S, Sharifi-Mood B, Davoodikia AA, Moody B, Naghavi A. Association between the CD14 gene C-159T polymorphism and serum soluble CD14 with pulmonary tuberculosis. *Int J Tuberc Lung Dis* 2012; 16(10): 1383-7.
[<http://dx.doi.org/10.5588/ijtld.11.0827>] [PMID: 23107636]
- [30] Mikuls TR, LeVan TD, Sayles H, *et al.* Soluble CD14 and CD14 polymorphisms in rheumatoid arthritis. *J Rheumatol* 2011; 38(12): 2509-16.
[<http://dx.doi.org/10.3899/jrheum.110378>] [PMID: 21921097]
- [31] Jiménez VC, Wit FW, Joerink M, *et al.* T-cell activation independently associates with immune senescence in HIV-infected recipients of long-term antiretroviral treatment. *J Infect Dis* 2016; 214(2): 216-25.
[<http://dx.doi.org/10.1093/infdis/jiw146>] [PMID: 27073222]
- [32] Klatt NR, Chomont N, Douek DC, Deeks SG. Immune activation and HIV persistence: implications for curative approaches to HIV infection. *Immunol Rev* 2013; 254(1): 326-42.
[<http://dx.doi.org/10.1111/imr.12065>] [PMID: 23772629]
- [33] Blish CA, McClelland RS, Richardson BA, *et al.* Genital inflammation predicts HIV-1 shedding independent of plasma viral load and systemic inflammation. *J Acquir Immune Defic Syndr* 2012; 61(4): 436-40.
[<http://dx.doi.org/10.1097/QAI.0b013e31826c2edd>] [PMID: 22878424]