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RESEARCH ARTICLE

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The Common Ancestor of Deinococcus Species was Rod-Shaped

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Abstract:

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Background:

The genus Deinococcus consists of species in rod-shape (Bacilli) and spherical shape (Cocci).

Objective:

In this study, we aimed to determine whether the common ancestor of *Deinococcus* species was rod-shaped or spherical.

Methods:

We compared the homologs of the proteins related to the rod-shape in bacteria (MreB, MreC, MreD, MrdA, RodA, and RodZ) in various Deinococcus species and Thermus thermophilus.

Results:

The phylogenetic trees based on each protein and the homologs reflected the evolutionary relationships of the species, indicating that the Horizontal transfer of the genes did not occur during the Deinococcus evolution.

Conclusion:

The ancestor of the genus Deinococcus was rod-shaped, and the spherical forms appeared when the rod-shaped formation system was lost during evolution and diversification within the genus.

Keywords: Deinococcus, Bacilli, Cocci, Thermus thermophilus, Phylogenetic trees, Evolution.

1. INTRODUCTION

In 1981, the genus Deinococcus was defined as cocci [1]. The radiation-resistant spherical bacterium Deinococcus radiodurans has a unique lipid composition, *i.e.*, glycolipids and glycophospholipids have been identified, but phosphatidylethanolamine and phosphatidylglycerol, which are common in other bacteria, are absent [2 - 4]. This unique character is observed in this bacterium [5]. In 1987, Deinobacter grandis was reported as a radiation-resistant rod-shaped bacterium [6]. In 1997, D. grandis was transferred to the genus Deinococcus based on 16S rDNA sequence comparison [7]. At present, the genus *Deinococcus* consists of both bacilli and cocci species. Evolutionarily, the order Deinococcales is closely related to the order Thermales [8]. These two orders constitute the phylum Deinococcus-Thermus. The purpose of this study was to elucidate whether the common ancestor of the Deinococcus species was spherical or rod-shaped on the basis of molecular evolutionary analyses. Although the last common ancestor of bacteria has been thought to be rod-shaped [9], the rod-shaped related proteins (MreB, MreC, MreD, MrdA, RodA, and RodZ) have been recently reported [10 - 13]. In spherical bacterium Staphylococcus aureus, MreC and MreD are not essential for cell viability and do not affect cell morphology [14]. If the genes coding for those six proteins had been inherited during the evolution of Deinococcus species, their common ancestor would be rod-shaped. If those genes had been

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acquired by horizontal transfer, the common ancestor would probably be spherical.

2. MATERIALS AND METHODS

In this study, we used the complete genome sequences of eleven species, and the almost complete genome sequences of three species (D. ficus, D. grandis, and D. hopiensis) of Deinococcus (Table 1). Homologs of Thermus thermophilus were used as out-groups. Orthologous protein sequence comparison is a powerful tool that enables a more accurate phylogenetic evaluation than that based on 16S and/or 23S rRNA sequence comparison [15]. Most orthologous proteins are ribosomal proteins [16]. Thus, in order to understand the evolutionary relationships among *Deinococcus* species, a phylogenetic tree was constructed using their ribosomal protein sequences. The amino acid sequences of the ribosomal proteins- S1, S2, S3, S4, S5, S6, S7, S8, S9, S10, S11, S12, S13, S14, S15, S16, S17, S18, S19, and S20 of the small subunit, and L1, L2, L3, L4, L5, L6, L7/L12, L9, L10, L11, L13, L14, L15, L16, L17, L18, L19, L20, L21, L22, L23, L24, L25, L27, L28, L29, L30, L31, L32, L33, L34, L35, and L36 of the large subunit, were used for the tree construction. We identified proteins, homologous to each of the six proteins, which are related to rod-shape in bacteria, namely MreB, MreC, MreD, MrdA, RodA, and RodZ, in each of the fifteen bacterial species considered in the study. Homologous proteins were selected on the basis of BLASTP search in NCBI. Multiple sequence alignment using MUSCLE [17] and phylogenetic tree construction in Maximum Likelihood (ML) method using MEGA software, version 6 were performed [18]. The Le_Gascuel_2008 (LG) model [19] was selected as the best model for each ML analysis. The nearest neighbor interchange was used as the heuristic approach for the ML method for inferring the phylogenetic tree. The γ -distribution rate was considered, and the number of discrete γ -categories was five. Bootstrap analysis was performed with 1000 replicates.

| Organism | Shape | Genome GC% | Assembly level (NCBI Accession Number) | MreB Homolog | MreC Homolog | MreD Homolog | MrdA(PBP2) Homolog | RodA Homolog | RodZ Homolog |
|--------------------------------|------------------------|---------------|---|-----------------|-----------------|-----------------|-----------------------|-----------------|-----------------|
| Deinococcus actinosclerus | rod/spherical | 70.6 | complete (CP013910.1, CP029774.1) | present | present | present | present | present | present |
| Deinococcus deserti | rod | 63.0 | complete (CP001114.1) | present | present | present | present | present | present |
| Deinococcus ficus | rod | 69.9 | 6 scaffolds (ATTJ00000000.1) | present | present | present | present | present | present |
| Deinococcus geothermalis | spherical | 66.6 | complete (CP000359.1) | lack | lack | lack | lack | lack | lack |
| Deinococcus gobiensis | spherical | 69.2 | complete (CP002191.1) | lack | lack | lack | lack | lack | lack |
| Deinococcus grandis | rod | 69.9 | 7 scaffolds (BCMS0000000.1) | present | present | lack | present | present | present |
| Deinococcus hopiensis | spherical | 64.9 | 11 scaffolds (FWWU00000000.1) | lack | lack | lack | lack | lack | lack |
| Deinococcus maricopensis | rod | 69.8 | complete (CP002454.1) | present | present | lack | present | present | present |
| Deinococcus peraridilitoris | short rod/spherical | 63.7 | complete (CP003382.1) | present | present | present | present | present | present |
| Deinococcus proteolyticus | spherical | 65.7 | complete (CP002536.1) | lack | lack | lack | lack | lack | lack |
| Deinococcus puniceus | spherical | 62.6 | complete (CP011387.1) | present | present | present | present | present | present |
| Deinococcus radiodurans | spherical | 66.7 | complete (AE000513.1, AE001825.1, CP015081.1, CP015082.1) | lack | lack | lack | lack | lack | lack |
| Deinococcus soli | short rod | 70.2 | complete (CP011389.1) | present | present | lack | present | present | present |
| Deinococcus swuensis | spherical | 67.4 | complete (CP010028.1) | present | present | present | lack | present | lack |

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3. RESULTS AND DISCUSSION

The ribosomal protein sequence comparison produced more accurate phylogenetic trees than the rRNA sequence comparison (Fig. 1). The phylogenetic tree based on the ribosomal protein sequences showed that neither bacilli nor

cocci formed a monophyletic lineage; both the spherical and rod-shaped species were scattered in the tree (Fig. 1).



Fig. (1). Phylogenetic tree showing evolutionary relationships among fourteen species of *Deinococcus* and *Thermus thermophilus*, inferred from the comparison of ribosomal protein sequences, and 16S and 23S rRNA sequences.

Phylogenetic tree based on ribosomal proteins: The evolutionary history was inferred using the maximum likelihood method based on the Le_Gascuel_2008 model [19]. The tree with the highest log likelihood (-63291.1582) is shown. The percentage of trees in which the associated taxa were clustered together is shown next to the branches. Initial tree(s) for heuristic search were obtained by applying the neighbor-joining method to a matrix of pair wise distances estimated using a JTT model [20]. A discrete γ -distribution was used to model evolutionary rate differences among sites (5 categories (+G, parameter = 0.4545)). The tree was drawn in scale, with branch lengths measured in the number of substitutions per site. The analysis involved 15 amino acid sequences. All positions containing gaps and missing data were eliminated. There were totally 6904 positions in the final set of data. Evolutionary analyses were conducted using MEGA 6 software [18].

Phylogenetic tree based on rRNAs: The evolutionary history was inferred using the maximum likelihood method based on the Tamura-Nei model [21]. The tree with the highest log likelihood (-19723.9210) is shown. The percentage of trees in which the associated taxa were clustered together is shown next to the branches. Initial tree(s) for heuristic search were obtained by applying the neighbor-joining method to a matrix of pair wise distances estimated using the maximum composite likelihood (MCL) approach. A discrete γ -distribution was used to model evolutionary rate differences among sites (5 categories (+G, parameter = 0.1283)). The tree was drawn in scale, with branch lengths measured in the number of substitutions per site. The analysis involved 15 nucleotide sequences. All positions containing gaps and missing data were eliminated. There were totally 4160 positions in the final set of data. Evolutionary analyses were conducted using MEGA 6 software [18].

The results revealed that most of the rod-shape related genes were not only distributed in *bacilli*, but also in *cocci*. For example, the spherical *D. puniceus* was shown to have all of the six rod-shape related genes (Table 1). It is uncertain whether all of the six genes are expressed and function in *D. puniceus*. On the other hand, the rod-shaped *D. grandis* and *D. maricopensis* lacked *mreD* homologs (Table 1). In addition, *D. actinosclerus* and *D. peraridilitoris*,

which can become bacilli or cocci, showed to have all six rod-shape related genes (Table 1).

Among eleven complete genomes of *Deinococcus* analyzed, *D. actinosclerus*, *D. deserti*, *D. peraridilitoris*, and *D. puniceus* had all the six rod-shape related genes (Table 1). Homologs of three genes among the six, *mreC*, *mreD*, and *mrdA*, formed a conserved gene cluster, which were found conserved in *T. thermophilus* also (Fig. 2). Although *mrdA* and *rodA* were found to be clustered in *Escherichia coli* [22, 23], *mrdA* homolog was not clustered with *rodA* homolog, but with *mreC* and *mreD* homologs in *Deinococcus* and *T. thermophilus* (Fig. 2). Homologs of the other three genes *mreB*, *rodA*, and *rodZ* were scattered in the genome of all the species (Fig. 2). Although *mreB*, *mreC*, and *mreD* were found clustered in *E. coli* [24], *mreB* homolog was not clustered with *mreC* and *mreD* homologs in *Deinococcus* and *T. thermophilus* (Fig. 2). These results indicate that horizontal transfer of the gene cluster did not occur.

| D. actinosclerus | mrdA mreD mre | C mreB | rodZ | rodA |
|--------------------|---------------|--------------|-----------|-------------|
| D. deserti | rodZ ro ← | odA mreE | 3 mrdA | A mreD mreC |
| D. peraridilitoris | mreCmreDmrd | A rodA | mreB ▲ | rodZ |
| D. puniceus | mrdA mreD mr | eC mreB | rodZ | rodA ◀ |
| T. thermophilus | rodZ n | nreCmreDmrdA | rodA | mreB |

Fig. (2). Locations of homologs of *mreB*, *mreC*, *mreD*, *mrdA*, *rodA*, and *rodZ* on genomes of *Deinococcus actinosclerus*, *D. deserti*, *D. peraridilitoris*, *D. puniceus*, *E. coli* and *T. thermophilus*.

Phylogenetic relationships inferred on the basis of rod-shape related genes showed that phylogenetic clusters were conserved. *D. actinosclerus*, *D. grandis*, and *D. soli* were found to be clustered when analyzed for their *mreB*, *mreC*, *rodA*, and *rodZ* homologs; *D. deserti* and *D. ficus* were clustered for their *mreB*, *mreC*, *mreD*, and *mrdA* homologs (Fig. 3). In addition, all phylogenetic trees deduced in this way indicated that *D. maricopensis* and *D. peraridilitoris* had diverged prior to the separation of other *Deinococcus* species (Fig. 3), which is in agreement with the evolution of *Deinococcus* species, elucidated from the ribosomal protein sequence comparison (Fig. 1).

The evolutionary history was inferred using the maximum likelihood method based on the Le_Gascuel_2008 model [19]. Phylogenetic trees based on rod-shape related proteins: The tree with the highest log likelihood (-2103.0678 in MreB, -2842.1661 in MreC, -1771.9295 in MreD, -6392.3576 in MrdA, -3330.3680 in RodA, and -3760.3540 in RodZ) is shown. The percentage of trees in which the associated taxa were clustered together is shown next to the branches. Initial tree(s) for the heuristic search were obtained by applying the neighbor-joining method to a matrix of pair wise distances estimated using a JTT model. A discrete γ -distribution was used to model evolutionary rate differences among sites (5 categories (+G, parameter = 0.5099 in MreB, 0.9195 in MreC, 1.1260 in MreD, 0.6919 in MrdA, 0.9724 in RodA, and 1.0505 in RodZ)). The tree is drawn in scale, with branch lengths measured in the number of substitutions per site. All positions containing gaps and missing data were eliminated. There were totally 345, 240, 152, 550, 342, and 270 positions of MreB, MreC, MreD, MrdA, RodA, and RodZ respectively, in the final set of data. Evolutionary analyses were conducted using MEGA 6 software [18].

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Fig. (3). Maximum likelihood trees based on rod-shape related proteins.

CONCLUSION

Our results showed that each of the rod-shaped related genes had been inherited in most of rod-shaped species of *Deinococcus* during the evolution of *Deinococcus* species. Thus, the common ancestor of *Deinococcus* species was rod-shaped. Major gene loss had occurred four times during the evolution of *Deinococcus* with respect to the rod-shape related genes, which led to the generation of *cocci* in the same genus (Fig. 4).



Fig. (4). Model of deletions of rod-shape related genes in *Deinococcus*. **a**: *mreB* homolog deletion, **b**: *mreC* homolog deletion, **c**: *mreD* homolog deletion, **d**: *mrdA* homolog deletion, **e**: *rodA* homolog deletion, **f**: *rodZ* homolog deletion.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

Not applicable.

HUMAN AND ANIMAL RIGHTS

No animals/humans were used for studies that are the basis of this research.

CONSENT FOR PUBLICATION

Not applicable.

CONFLICTS OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

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REFERENCES

- Brooks BW, Murray RGE. Nomenclature for "*Micrococcus radiodurans*" and other radiation-resistant cocci: Deinococcaceae fam. nov. and *Deinococcus* gen. nov., including five species. Int J Syst Bacteriol 1981; 31(3): 353-60. [http://dx.doi.org/10.1099/00207713-31-3-353]
- [2] Anderson R, Hansen K. Structure of a novel phosphoglycolipid from *Deinococcus radiodurans*. J Biol Chem 1985; 260(22): 12219-23.
 [PMID: 4044593]
- Huang Y, Anderson R. Structure of a novel glucosamine-containing phosphoglycolipid from *Deinococcus radiodurans*. J Biol Chem 1989; 264(31): 18667-72.
 [PMID: 2808391]
- Sohlenkamp C, Geiger O. Bacterial membrane lipids: Diversity in structures and pathways. FEMS Microbiol Rev 2016; 40(1): 133-59. [http://dx.doi.org/10.1093/femsre/fuv008] [PMID: 25862689]
- [5] Nishino K, Morita Y, Takahashi S, et al. Enlargement of Deinococcus grandis spheroplasts requires Mg²⁺ or Ca². Microbiology 2018. [http://dx.doi.org/10.1099/mic.0.000716] [PMID: 30222092]
- [6] Oyaizu H, Stackebrandt E, Schleifer KH, et al. A radiation-resistant rod-shaped bacterium, *Deinobacter grandis* gen. nov., sp. nov., with peptidoglycan containing ornithine. Int J Syst Bacteriol 1987; 37(1): 62-7. [http://dx.doi.org/10.1099/00207713-37-1-62]
- [7] Rainey FA, Nobre MF, Schumann P, Stackebrandt E, da Costa MS. Phylogenetic diversity of the deinococci as determined by 16S ribosomal DNA sequence comparison. Int J Syst Bacteriol 1997; 47(2): 510-4.
 [http://dx.doi.org/10.1099/00207713-47-2-510] [PMID: 9103641]
- [8] Ho J, Adeolu M, Khadka B, Gupta RS. Identification of distinctive molecular traits that are characteristic of the phylum "Deinococcus-Thermus" and distinguish its main constituent groups. Syst Appl Microbiol 2016; 39(7): 453-63. [http://dx.doi.org/10.1016/j.syapm.2016.07.003] [PMID: 27506333]
- [9] Siefert JL, Fox GE. Phylogenetic mapping of bacterial morphology. Microbiology 1998; 144(Pt 10): 2803-8. [http://dx.doi.org/10.1099/00221287-144-10-2803] [PMID: 9802021]
- [10] Shiomi D, Sakai M, Niki H. Determination of bacterial rod shape by a novel cytoskeletal membrane protein. EMBO J 2008; 27(23): 3081-91. [http://dx.doi.org/10.1038/emboj.2008.234] [PMID: 19008860]
- Bendezú FO, de Boer PA. Conditional lethality, division defects, membrane involution, and endocytosis in *mre* and *mrd* shape mutants of *Escherichia coli*. J Bacteriol 2008; 190(5): 1792-811.
 [http://dx.doi.org/10.1128/JB.01322-07] [PMID: 17993535]
- [12] Bendezú FO, Hale CA, Bernhardt TG, de Boer PA. RodZ (YfgA) is required for proper assembly of the MreB actin cytoskeleton and cell shape in *E. coli*. EMBO J 2009; 28(3): 193-204. [http://dx.doi.org/10.1038/emboj.2008.264] [PMID: 19078962]
- Shiomi D, Toyoda A, Aizu T, *et al.* Mutations in cell elongation genes *mreB*, *mrdA* and *mrdB* suppress the shape defect of RodZ-deficient cells. Mol Microbiol 2013; 87(5): 1029-44.
 [http://dx.doi.org/10.1111/mmi.12148] [PMID: 23301723]
- Tavares AC, Fernandes PB, Carballido-López R, Pinho MG. MreC and MreD proteins are not required for growth of *Staphylococcus aureus*. PLoS One 2015; 10(10): e0140523.
 [http://dx.doi.org/10.1371/journal.pone.0140523] [PMID: 26470021]
- [15] Oshima K, Nishida H. Phylogenetic relationships among mycoplasmas based on the whole genomic information. J Mol Evol 2007; 65(3): 249-58.

[http://dx.doi.org/10.1007/s00239-007-9010-3] [PMID: 17687503]

- [16] Nishida H, Beppu T, Ueda K. Whole-genome comparison clarifies close phylogenetic relationships between the phyla Dictyoglomi and Thermotogae. Genomics 2011; 98(5): 370-5.
 [http://dx.doi.org/10.1016/j.ygeno.2011.08.001] [PMID: 21851855]
- [17] Edgar RC. MUSCLE: multiple sequence alignment with high accuracy and high throughput. Nucleic Acids Res 2004; 32(5): 1792-7. [http://dx.doi.org/10.1093/nar/gkh340] [PMID: 15034147]
- [18] Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. MEGA6: Molecular evolutionary genetics analysis version 6.0. Mol Biol Evol 2013; 30(12): 2725-9.
 [http://dx.doi.org/10.1093/molbev/mst197] [PMID: 24132122]

- [19] Le SQ, Gascuel O. An improved general amino acid replacement matrix. Mol Biol Evol 2008; 25(7): 1307-20. [http://dx.doi.org/10.1093/molbev/msn067] [PMID: 18367465]
- [20] Jones DT, Taylor WR, Thornton JM. The rapid generation of mutation data matrices from protein sequences. Comput Appl Biosci 1992; 8(3): 275-82.
 [PMID: 1633570]
- [21] Tamura K, Nei M. Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. Mol Biol Evol 1993; 10(3): 512-26. [PMID: 8336541]
- [22] Tamaki S, Matsuzawa H, Matsuhashi M. Cluster of *mrdA* and *mrdB* genes responsible for the rod shape and mecillinam sensitivity of *Escherichia coli*. J Bacteriol 1980; 141(1): 52-7.
 [PMID: 6243629]
- [23] Spratt BG, Boyd A, Stoker N. Defective and plaque-forming lambda transducing bacteriophage carrying penicillin-binding protein-cell shape genes: genetic and physical mapping and identification of gene products from the *lip-dacA-rodA-pbpA-leuS* region of the *Escherichia coli* chromosome. J Bacteriol 1980; 143(2): 569-81. [PMID: 6451612]
- Wachi M, Doi M, Okada Y, Matsuhashi M. New *mre* genes *mreC* and *mreD*, responsible for formation of the rod shape of *Escherichia coli* cells. J Bacteriol 1989; 171(12): 6511-6.
 [http://dx.doi.org/10.1128/jb.171.12.6511-6516.1989] [PMID: 2687239]

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