

Dynamics of a Free-Living Nitrogen-Fixing Bacteria Population Lacking of Competitive Advantage Towards an Antagonistic Population

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Abstract: The dynamics of a model of a free-living nitrogen-fixing population, grown in a chemostat alone or in competition with a non-nitrogen-fixing population, was studied through bifurcation analysis. It is demonstrated that the ability of the nitrogen-fixing population to survive depends on the ammonia and carbon source concentrations and the kinetics parameters of the system. Under competitive conditions the nitrogen-fixing population, characterized by low growth rates, can survive alone at stable steady state only at low dilution rate values. Especially, there is a threshold of the dilution rate above which only the competitive population can survive.

When amensalistic interactions are established, coexistence of both populations is favored. At low dilution rate values the survival of the nitrogen-fixing population alone is restricted while this population can survive at higher ammonia concentrations either in coexistence state or in multistability state together with the competitive population. When nitrogen-fixing population successfully inhibits its competitor, then its survival is enhanced towards the coexistence state.

Keywords: Population dynamics, nitrogen fixation, competition, coexistence.

1. INTRODUCTION

The biogeochemical cycle of nitrogen is essential for agriculture, as well as for the productivity of natural ecosystems. Nitrogen being of major importance for all living organisms is often a limiting growth factor in soil ecosystem. Soil reservoirs are fed by biologically and/or by chemically fixed nitrogen (in the form of fertilizers), but the balance is frequently negative due to significant losses through denitrification, erosion, leaching and volatilization.

Biological nitrogen fixation is considered as the limiting step of the nitrogen cycle, as this process is functioned by a restricted number of prokaryotes, including bacteria of the genus *Azospirillum* and specific symbiotic associations [1-3]. Other organisms (animals, plants, microorganisms) require fixed forms of nitrogen.

Abiotic factors and biotic interactions affect the survival of free-living nitrogen-fixing populations. Within the chemical factors affecting nitrogen-fixing bacteria are included the concentration and nature of organic components excreted by plant roots [4-13], while within biotic interactions are included predation and parasitism, competition for resources and interspecific microbial interactions having either

negative (i.e. inhibitory) or positive (i.e., through the increase of nutrient availability) effects on the microbial community [14-19].

In the natural ecosystems the low concentration of the carbon source is critical for microbial growth and survival, especially for nitrogen-fixing populations, since the fixation of nitrogen is a highly energy consuming process [20]. Furthermore, the nitrogenase complex (responsible for the conversion of molecular nitrogen into ammonium), is inhibited by ammonium nitrogen, in both transcriptional and enzymatic activity levels [21]. Ammonium nitrogen under aerobic conditions enhances the growth of *Azospirillum* [22] but under high ammonium and low dissolved oxygen concentrations the growth is inhibited [10, 23-25]. Under microaerobic conditions *Azospirillum* can grow in the presence of low ammonium nitrogen concentration, while after the depletion of ammonium nitrogen, they are able to grow by fixing molecular nitrogen [25-28].

Numerous mathematical models have been used in the study of interactions between species that share common resources and of the effect of predators on the dynamics of biological systems [29-39]. In the traditional models (i.e. in the Lotka-Volterra competition model and similar) the interaction of one species to the other is always negative, while coexistence occurs only if between-species competition is weaker than within species. Kooi and Kooiman [32] proved that a competitor can stabilize a nutrient-prey-predator system, while Zhang [40], introducing mutualism in competitive

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systems, showed that coexistence of populations is promoted while the carrying capacity of the system is enhanced.

Recently, the dynamics of free-living nitrogen-fixing populations in antagonistic and predation conditions was studied using several mathematical models [41, 42]. By means of bifurcation analysis it was shown that, under a variety of conditions, the presence of competitors and predators is beneficial for the system and enhances coexistence.

The aim of this work is to study the dynamics of a free-living nitrogen-fixing population grown in a chemostat alone or together with a competitive non-nitrogen-fixing population. Specifically, it is reported the ability of the nitrogen-fixing population to survive, although it is initially disadvantaged by a lower growth rate comparing to that of its competitor.

2. MODELING

Dynamics of nitrogen-fixing populations was studied using the following models published in Ecological Modeling [41], except for Model I which appears in this paper for the first time.

Model I

In the first model it is assumed that a nitrogen-fixing microbial population (N_1) grows alone in a chemostat under both carbon and nitrogen limitation.

Equations of this system are:

Balance of N_1 populations:

$$\frac{dN_1}{dt} = (r_1 - D) \cdot N_1 \quad (1)$$

where, r_1 is the specific growth rates of the N_1 population and D is the chemostat dilution rate.

Balance of carbon and energy source:

$$\frac{dS}{dt} = D \cdot (S_f - S) - r_1 \cdot N_1 \cdot \frac{1}{Y_{N_1/S}} - r_E \cdot E \cdot N_1 \cdot \frac{1}{Y_{NH_3/S}} \quad (2)$$

where, S_f and S are the carbon source concentrations in the inlet and outlet medium, respectively; $Y_{N_1/S}$ is the yield coefficient of N_1 on S ; E is the nitrogenase concentration (units per volume of N_1); r_E is the specific rate of nitrogen fixation; $Y_{NH_3/S}$ is the ammonium nitrogen yield coefficient on S .

Balance of ammonium nitrogen:

$$\frac{dNH_3}{dt} = D \cdot [(NH_3)_f - (NH_3)] + r_E \cdot E \cdot N_1 - r_1 \cdot N_1 \cdot \frac{1}{Y_{N_1/NH_3}} \quad (3)$$

where, $(NH_3)_f$ and (NH_3) are the ammonium nitrogen concentrations in the inlet and outlet medium, respectively; Y_{N_1/NH_3} is the yield coefficient on ammonium nitrogen of N_1 .

Balance of nitrogenase (units per volume of N_1):

$$\frac{dE}{dt} = q_E - r_D \cdot E - r_1 \cdot E \quad (4)$$

where, q_E is the specific rate of nitrogenase synthesis; r_D is the specific rate of nitrogenase destruction. In Eq. (4) the term $r_1 \cdot E$ represents the decrease of active protein concentration observed due to the increase of the cell volume [43].

The specific growth rate of the N_1 population is modeled by Monod's model [44] as modified by Graef and Andrews [45]:

$$r_1(S, NH_3) = r_{1\max} \cdot \frac{S}{K_{1S} + S} \cdot \frac{NH_3}{K_{1NH_3} + NH_3 + \left(K_{iNH_3} \cdot (NH_3)^2 \right)} \quad (5)$$

where, $r_{1\max}$ is the maximum specific growth rate; K_{1S} and K_{1NH_3} are the saturation constants for S and NH_3 , respectively; K_{iNH_3} is the inhibition constant of ammonium nitrogen on N_1 .

Nitrogen fixing activity is regulated in both nitrogenase synthesis (transcriptional regulation) and nitrogenase activity levels, as following (Eqs. 6-9).

The specific rate of nitrogenase synthesis is given as:

$$q_E(Q, r_1) = a \cdot r_1 \cdot Q \quad (6)$$

where, Q is described by

$$Q(NH_3) = \frac{1}{1 + C \cdot NH_3} \quad (7)$$

where C is a parameter related to the *nif* operators found under catabolic repression. Q is a function of the repressor compound (NH_3) [41] and accounts for modification of the rate of nitrogenase synthesis via a catabolic repression control mode.

The specific rate of nitrogen fixation is given by the equation,

$$r_E(S, NH_3) = r_{E\max} \cdot \frac{K_{iE}}{K_{iE} + NH_3} \cdot \frac{S}{K_{SE} + S} \quad (8)$$

where, $r_{E\max}$ is the maximum specific rate of nitrogen fixation; K_{SE} is the saturation constant; K_{iE} is the inhibition constant.

A conceptual diagram illustrating the system described by Model I is given in Fig. (1).

Model II

In the second model (described in Kavadia *et al.* [41]) it is assumed that a nitrogen-fixing microbial population (N_1) grows in a chemostat, together with a competitive microbial population (N_2), while no other interactions (except for competition for the common resources) between the two populations occur. The equations (1) and (4)-(8) concerning the nitrogen-fixing population are already presented in model I. Additional equations of this system are:

Balance of N_2 population:

$$\frac{dN_2}{dt} = (r_2 - D) \cdot N_2 \quad (9)$$

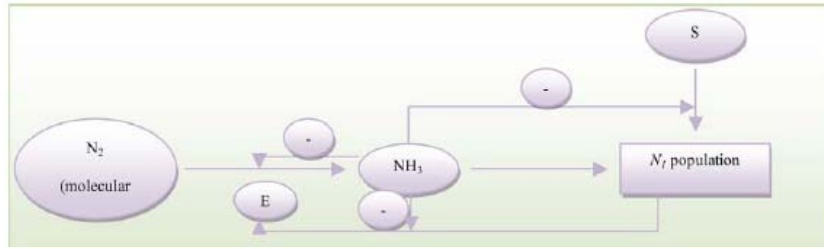


Fig. (1). Conceptual diagram which illustrates the model used for the study of the dynamics of a nitrogen-fixing population (N_1) grown alone in a chemostat in the presence of ammonium nitrogen (NH_3). The limiting growth factors are the carbon and energy source (S) and the NH_3 . When the concentration of NH_3 is low in the growth environment, S is partitioned in order to sustain both, synthesis of cellular mass of N_1 and nitrogenase activity (E). When NH_3 is accumulated in the growth environment, then, besides N_1 population, E synthesis and activity are inhibited.

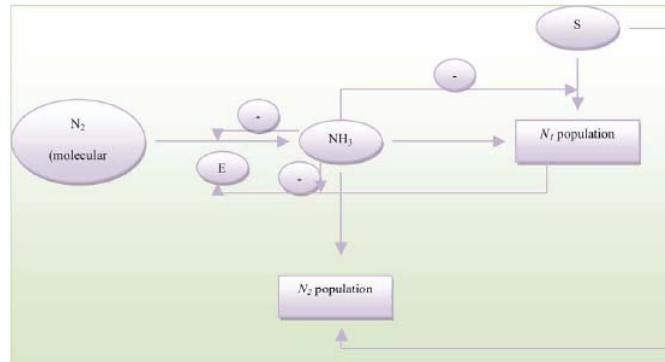


Fig. (2). Conceptual diagram which illustrates the models used for the study of the dynamics of a nitrogen-fixing population grown in antagonistic conditions. A nitrogen-fixing population (N_1) grows together with a competitive microbial population (N_2) in the presence of ammonium nitrogen (NH_3). The limiting growth factors are, for both populations, the carbon and energy source (S) and the NH_3 . When the concentration of NH_3 is low in the growth environment, S is partitioned in order to sustain both, synthesis of cellular mass of N_1 and nitrogenase activity (E). When NH_3 is accumulated in the growth environment, then, besides N_1 population, E synthesis and activity are inhibited. The presence of N_2 population, however, could discharge the system from NH_3 , and therefore could promote survival of N_1 population.

where, r_2 is the specific growth rate of the N_2 population; D is the chemostat dilution rate.

Balance of carbon and energy source:

$$\frac{dS}{dt} = D \cdot (S_f - S) - r_1 \cdot N_1 \cdot \frac{1}{Y_{N_1/S}} - r_2 \cdot N_2 \cdot \frac{1}{Y_{N_2/S}} - r_E \cdot E \cdot \frac{1}{Y_{NH_3/S}} \cdot N_1 \quad (10)$$

where, S_f and S are the carbon source concentrations in the inlet and outlet medium, respectively; $Y_{N_1/S}$, $Y_{N_2/S}$ are the yield coefficients of N_1 and N_2 on S , respectively; E is the nitrogenase concentration (units per volume of N_1); r_E is the specific rate of nitrogen fixation; $Y_{NH_3/S}$ is the ammonium nitrogen yield coefficient on S .

Balance of ammonium nitrogen:

$$\frac{dNH_3}{dt} = D \cdot [(NH_3)_f - (NH_3)] + r_E \cdot E \cdot N_1 - r_1 \cdot N_1 \cdot \frac{1}{Y_{N_1/NH_3}} - r_2 \cdot N_2 \cdot \frac{1}{Y_{N_2/NH_3}} \quad (11)$$

where, $(NH_3)_f$ and (NH_3) are the ammonium nitrogen concentrations in the inlet and outlet medium, respectively; Y_{N_1/NH_3} and Y_{N_2/NH_3} are the yield coefficients on ammonium nitrogen of N_1 and N_2 , respectively.

The specific rate for microbial growth of the non-nitrogen-fixing population (N_2) is given by the following equation:

$$r_2(S, NH_3) = r_{2max} \cdot \frac{S}{K_{2S} + S} \cdot \frac{NH_3}{K_{2NH_3} + NH_3} \quad (12)$$

where, r_{2max} is the maximum specific growth rate of the N_2 population; K_{2S} and K_{2NH_3} are the saturation constants for S and NH_3 , respectively.

A respective diagram illustrating the system described by Model II is given in Fig. (2).

Model III

The third model involves the ability of the nitrogen-fixing population to establish an amensalistic interaction through the synthesis of an inhibitory substance released in the growth environment [41].

The balance equation for the inhibitor in the chemostat is used in addition to balance equations (1), (4) and (9)-(11) which describe model II:

$$\frac{dI}{dt} = (a_I \cdot r_1 + b_I) \cdot N_1 - D \cdot I \quad (13)$$

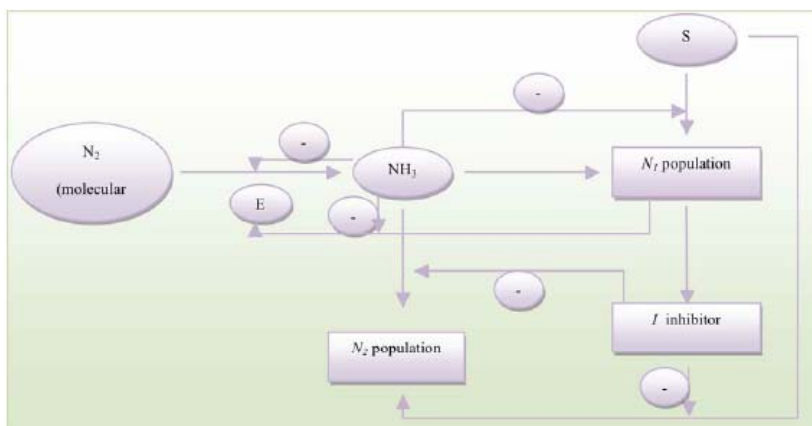


Fig. (3). Conceptual diagram which illustrates the model used for the study of the dynamics of a nitrogen-fixing population grown in amensalistic conditions. A nitrogen-fixing population (N_1) grows together with a competitive microbial population (N_2) in the presence of ammonium nitrogen (NH_3). The limiting growth factors are, for both populations, the carbon and energy source (S) and the NH_3 . When the concentration of NH_3 is low in the growth environment, S is partitioned in order to sustain both, synthesis of cellular mass of N_1 and nitrogenase activity (E). When NH_3 is accumulated in the growth environment, then, besides N_1 population, E synthesis and activity are inhibited. The presence of N_2 population, however, could discharge the system from NH_3 , and therefore could promote survival of N_1 population. Additionally, the N_1 population is establishing an amensalistic interaction through the synthesis of an inhibitory substance (I) released in the growth environment.

where, a_I is a stoichiometric constant for production of the inhibitor (I) during primary metabolic growth; b_I is a stoichiometric constant for production of the inhibitor (I) during secondary metabolic growth.

Accordingly, equation (12) that describes specific growth rate of the N_2 population should be replaced by the following equation:

$$r_2(S, NH_3) = r_{2max} \cdot \frac{S}{K_{2S} + S} \cdot \frac{NH_3}{K_{2NH_3} + NH_3} \cdot \frac{K_i}{K_i + I} \quad (14)$$

where, K_i is the inhibition constant.

The diagram illustrating this system is given in Fig. (3).

3. METHODOLOGY

The parameter values have major effect on the long-term behavior of the system. Of particular importance is the effect of the operating parameters, i.e. of the chemostat dilution rate D controlled by the medium flowrate through the reactor and the ammonium nitrogen concentration in the feed.

The effect of the operating parameters on the system is illustrated through two-parameter bifurcation diagrams. The construction of these diagrams is accomplished by developing numerical algorithms based on the continuation algorithm AUTO [46]. For the three above-mentioned systems there are four possible steady states and the possibility of stable periodic states of coexistence of N_1 and N_2 in which the population densities in the chemostat vary periodically with time:

- (1) Extinction of all populations: $N_1 = N_2 = 0$ (washout state).
- (2) Survival of N_1 only: $N_1 > 0, N_2 = 0$ (N_1 state).
- (3) Survival of N_2 only: $N_1 = 0, N_2 > 0$ (N_2 state).
- (4) Survival of N_1 and N_2 only: $N_1 > 0, N_2 > 0$ (N_1N_2 state)

Other important parameters are the inhibition constant K_{iNH_3} of ammonium nitrogen on the nitrogen-fixing population (representing the susceptibility of the nitrogen-fixing population to the ammonium containing media and affecting the specific growth rate of the N_1 population) and r_{max} and K_S of the N_1 and N_2 populations that determine which population have the competitive advantage. The value of operating parameter S_f for all bifurcation diagrams was kept constant at $S_f = 5$ g/L. The common parameter values used in all models for the cases studied in this paper were: $r_{1max} = 0.15$ h⁻¹, $r_{2max} = 0.2$ h⁻¹, $r_{Emax} = 0.2$ h⁻¹, $K_{iNH_3} = 0.5$ g/L, $K_i = 0.01$ or 0.2 g/L, $K_{1NH_3} = 0.1$ g/L, $K_{iE} = 0.02$ g/L, $K_{SE} = 0.2$ g/L, $Y_{N1/S} = 0.3$, $Y_{N1/NH_3} = 0.3$, $Y_{NH_3/S} = 0.1$, $C = 50$, $a = 2$, $r_{DE} = 0.001$ h⁻¹, $K_{2NH_3} = 0.5$ g/L, $Y_{N2/S} = 0.3$, $Y_{N2/NH_3} = 0.5$, $a_i = 0.1$, $b_i = 0.01$.

4. RESULTS

4.1. Model I: Dynamics of a Nitrogen-Fixing Population Grown Alone in a Chemostat

This system was studied for the case where the nitrogen-fixing population grows alone in a chemostat. This simple model represents the base on which more complicated models of nitrogen-fixing dynamic systems can be developed.

Comparing this system to the conventional one (in which no nitrogen-fixing populations are involved), we observe that ammonium has a double effect on population growth and nitrogen fixation. In particular, ammonium in low concentrations has a positive effect on growth (as nitrogen constitutes a major nutrient), while in high concentrations inhibits both, the bacterial growth and nitrogenase synthesis. The ammonium concentrations in which the growth of the bacterial population and nitrogenase synthesis and activity are inhibited depends on the values of the parameters K_{iNH_3} , C and K_{iE} (see equations 5, 7, 8). In Fig. (4) is shown a representative example of the above mentioned system, plotted as bifurcation diagram of D vs $(NH_3)_f$ for $S_f = 5$ g/L. This diagram consists of a transcritical bifurcation curve (marked T_1), on

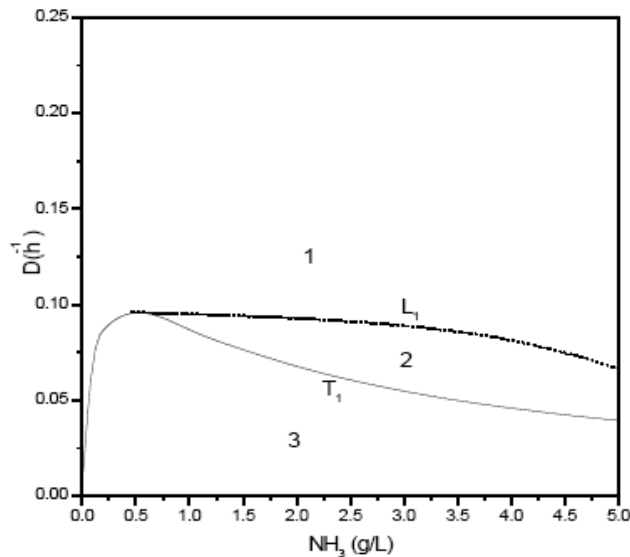


Fig. (4). Bifurcation diagram (D vs $(NH_3)_f$) for a chemostat system in which a nitrogen-fixing microbial population (N_1) grows alone.

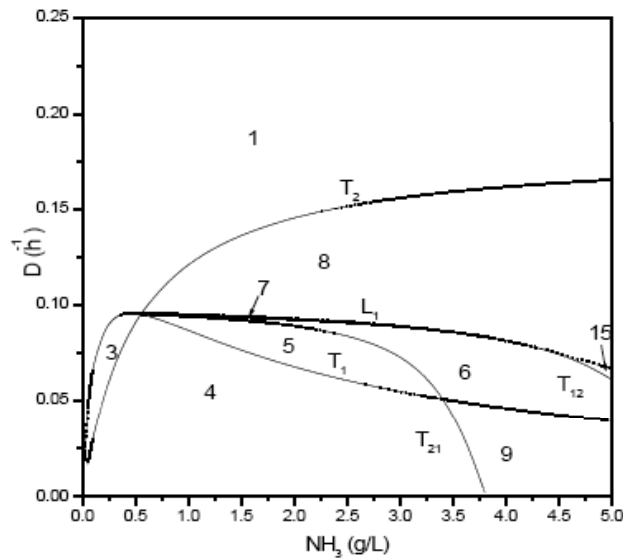


Fig. (5). Bifurcation diagram (D vs $(NH_3)_f$) for a chemostat system in which a nitrogen-fixing microbial population (N_1) grows together with its competitor (N_2).

which two steady states exchange stability characteristics and one limit-point bifurcation curve (L_1) on which two steady states are born. The character of each steady state in the regions of the bifurcation diagrams is symbolized by S and U1. S denotes stable steady state, whereas U1 indicates an unstable steady state with one positive eigenvalue.

It is considered that, for the given set of parameter values, the population is strongly inhibited by ammonium nitrogen. In region 1 of the diagram only extinction of the populations is possible, since the washout state is the only stable (S-state). In region 2, multistability is obtained since two steady states are stable, the washout state and the state of survival of the N_1 population (S and U1, S states, respectively). It depends on the initial state of the chemostat which of the two states will be eventually reached. In region 3 N_1 population survives at S-state.

4.2. Model II: Dynamics of a Nitrogen-Fixing Population Lacking of the Competitive Advantage Towards an antagonistic for the Common Resources Population

The dynamic behaviour of the system, where population N_1 is strongly inhibited by ammonium nitrogen and at the same time has not competitive advantage towards population N_2 , is presented in Fig. (5). In these conditions we expect that population N_1 can survive with difficulty.

The diagram D vs $(NH_3)_f$ for $S_f=5$ g/L consists of four transcritical bifurcation curves (marked T_1, T_2, T_{12}, T_{21}), on which two steady states exchange stability characteristics and one limit-point bifurcation curve (L_1) on which two steady states are born. The character of each steady state in the various regions of the diagram is symbolized by S, U1 and U2. S denotes stable steady state, whereas U1 indicates an unstable steady state with one positive eigenvalue and U2 an unstable steady state with two eigenvalues with positive real parts. In region 1 of the diagram only extinction of the populations is possible, since the washout state is the only stable. In region 2 two steady states are stable, the washout state and the state of survival of the N_1 population (multistability). It depends on the initial state of the chemostat which of the two states will be eventually reached. In regions 3-5 N_1 population survives alone in stable steady state (either S or U2, S states). Multistability is also observed in regions 6, 7, 9 since both the N_1 state (U2,S/U1,S/S states, respectively) and the N_2 state (S state in all regions) are stable. In regions 8 and 15 the population N_2 dominates over N_1 , as the N_2 state is the only stable. Coexistence exists only in regions 6 and 9, but it is at unstable state (U1 state).

4.3. Model III: Dynamics of a Nitrogen-Fixing Population Lacking of the Competitive Advantage Towards an Antagonistic for the Common Resources Population but Able to Establish Amensalistic Interactions

In this model the population N_2 is inhibited by the compound I , produced during both primary and secondary metabolic growth of N_1 . The population N_1 has the competitive advantage over N_2 at low ammonium concentrations (e.g. $(NH_3)_f < 0.7$ g/L). However, due to the high K_{iNH_3} value, the population N_1 is strongly inhibited at high ammonium nitrogen concentrations, and therefore, loses its competitive advantage.

The bifurcation diagram of D vs $(NH_3)_f$ (for $S_f=5$ g/L) is shown in Fig. (6) where the numbering of the regions is consistent with the one in the diagram shown in Fig. (5). This diagram contains four transcritical bifurcation curves (T_1, T_2, T_{12}, T_{21}), two limit point bifurcation curves (L and L_1) and one Hopf bifurcation curve (HB) on which a stable periodic state of coexistence is born. Comparing this diagram with the one shown in Fig. (5) we observe that four new regions appear (namely 10-13) due to the additional interaction between the two populations, caused by the inhibitory compound. Furthermore, the region of multistability 15 (in which both states of survival of N_1 and coexistence are stable) is not present, whereas region 5 (in which the state of survival of N_1 is the only stable) is significantly reduced.

Multistability is observed in region 11 since the states of survival of each population alone (N_1 and N_2 states are char-

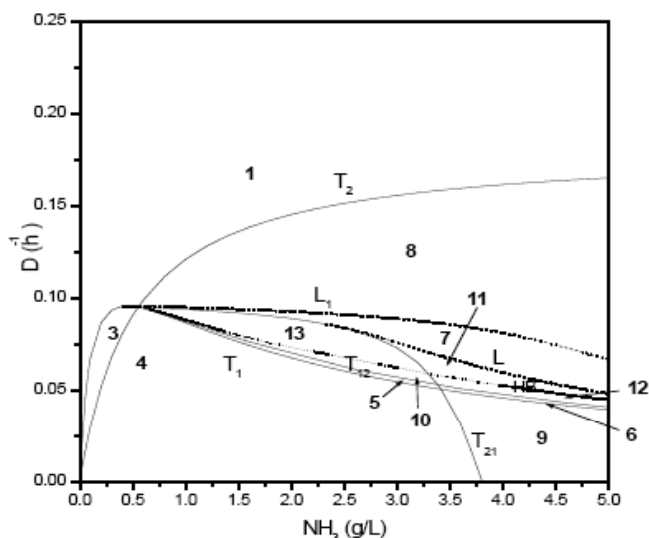


Fig. (6). Bifurcation diagram (D vs $(NH_3)_f$) for a chemostat system in which a nitrogen-fixing microbial population (N_1), which is able to establish an amensalistic interaction in the growth environment, grows together with its competitor (N_2).

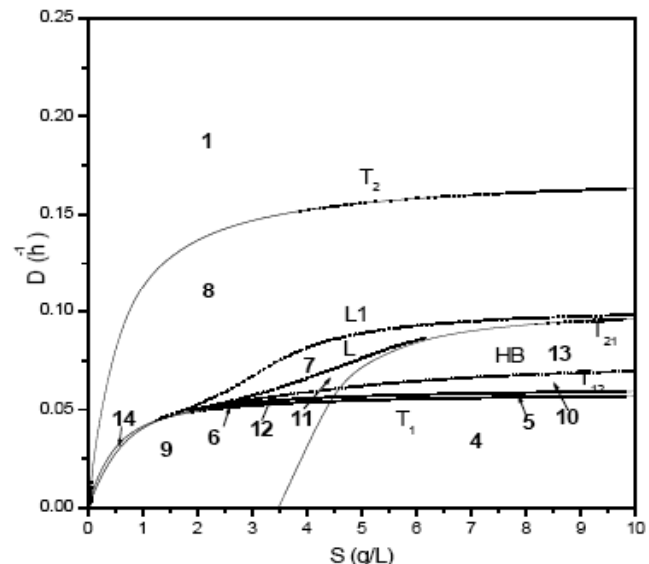


Fig. (7). Bifurcation diagram (D vs S_f) for a chemostat system in which a nitrogen-fixing microbial population (N_1), which is able to establish an amensalistic interaction in the growth environment, grows together with its competitor (N_2).

acterized as U1, S and S states respectively), as well as the coexistence state (U1, S states), are stable. It depends on the initial conditions which state will be reached by the system. Multistability is also observed in region 13 where both the state of survival of N_1 (U1, S states) and state of survival of coexistence (S state) are stable. In region 10 there is also multistability, since survival of the N_1 population is observed in a stable steady state (U1,S states) together with a stable coexistence periodic state (U2,(P) states). In region 12 survival of each population alone is observed in a stable steady state and coexistence in a stable periodic state (U1,S /S/U2,U1(P) states respectively for N_1/N_2 and N_1N_2 states). Finally, in region 14 only population N_2 can survive at stable steady state (S state).

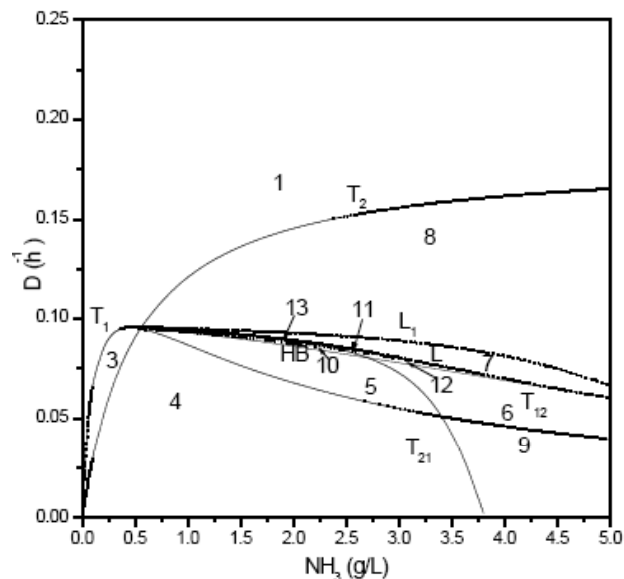


Fig. (8). Bifurcation diagram (D vs $(NH_3)_f$) for a chemostat system in which a nitrogen-fixing microbial population (N_1), which is able to establish an amensalistic interaction in the growth environment, grows together with its competitor (N_2).

The bifurcation diagram of D vs S_f (for $(NH_3)_f = 3$ g/L) is given in Fig. (7). At this NH_3 concentration, population N_1 is strongly inhibited. It is observed that population N_1 survives alone at $S > 3.5$ g/L and $D < 0.05$ h⁻¹ (regions 4, 5). At low carbon concentration population N_1 can survive at low dilution rate values ($D < 0.1$ h⁻¹) depending on initial conditions (regions 6, 7, 9, 11 and 12). Multistability exists in regions 10 and 13 as population N_1 survives alone at stable steady state and in coexistence with N_2 at stable periodic state (region 10) or in coexistence with N_2 at stable steady state (region 13). Finally in region 8 population N_2 survives alone in stable state for a wide range of parameter values.

Fig. (8) shows that the population N_1 is strongly inhibited by ammonium nitrogen (due to the high K_{iNH_3} value) and thus loses its competitive advantage even at low NH_3 concentration. However, N_1 could survive by creating an unsuitable environment for its competitor. Comparing to Fig. (6) it is observed a reduction of regions 11 (N_1 and N_2 and N_1N_2 at stable steady state) and 13 (N_1 and N_1N_2 at stable steady state) and a benefit of region 5 where N_1 dominates. Also regions 7 (N_1 and N_2 at stable steady state) and 12 (N_1 and N_2 at stable steady state and N_1N_2 at stable periodic state) are reduced, while region 6 (N_1 and N_2 at stable steady state) is increased.

DISCUSSION

The role of the free-living nitrogen-fixing populations is of great significance for the biogeochemical cycle of nitrogen [1], and these bacteria have been successfully used in environmental applications [24, 28, 47] and sustainable agriculture [10, 48]. Their ability to survive and fix nitrogen is highly influenced by ammonium nitrogen concentration in the soil. Specifically, under high ammonia concentration the nitrogenase complex becomes inactivate and the bacteria stop fixing nitrogen, while under low dissolved oxygen concentration ammonium inhibits microbial growth. Further-

more, nitrogen fixation is a highly energy consuming process, while the carbon and energy sources are frequently present at low concentration that are actually limiting for the free-living nitrogen-fixing populations growing competitively in the soil microbial community.

The theoretical population picked in this study was sensitive to high ammonia concentrations and by default it was considered to grow with low growth rate. It was shown that the parameter set used (i.e. the low growth rate of N_1 population and the high inhibition by ammonia) constrains its ability to survive. For instance at high dilution rate values only N_2 population survives at stable steady state.

Although a number of models have been proposed for the study of the biogeochemical cycle of nitrogen [49-55], studies of the dynamics of nitrogen-fixing populations have not been published. Only some interactions between *Azospirillum* and other microorganisms have been previously studied using specific experimental systems [4, 56, 57]. Experimental studies regarding prey-predator interactions showed that nitrogen fixing bacteria (as well as other Gram negative species) are prey for several important soil predators such as *Bdellovibrio* and related organisms and protozoa [14-18].

Competitive interactions are often established between microbial populations, since many nutrients are found in limited concentrations in common habitats [4]. However, under certain conditions the presence of antagonistic populations could relieve the growth environment from certain inhibitors, such as ammonium nitrogen. Specifically the nitrogen-fixing populations can be affected by their competitors in two opposite ways [6, 7, 40]: negatively due to the competition occurring for the common resources, and positively due to the decrease of the ammonium nitrogen concentration related to the growth of the competitive population.

In this work we show that at high concentrations of the carbon source (i.e., $S_f=5$ g/L), the N_1 population can survive at stable steady state for a wide range of parameter values and operating conditions even without competitive advantage towards the second population. This fact induces major changes in regions configuration. Comparing to Kavadia *et al.* [41] it is observed that the growth rate of the population has a significant impact on its survival. Specifically, it is shown that N_1 population cannot survive above a threshold of the dilution rate ($D>0.1$ h⁻¹). Also, the threshold of ammonia at which N_1 population is able to survive in multistability conditions (N_1 and N_2 state) is getting lower. Therefore the lower growth rate of N_1 population joint with the high inhibition by ammonia concentration constrains its survival ability. At high dilution rate values N_2 is the only survivor at stable steady state regardless of ammonia and carbon concentrations.

The balance of positive – negative effects of population N_2 on the survival of N_1 could be modified by population N_1 itself, i.e., by establishing an amensalistic interaction through the production of an inhibitory substance that has a negative effect on the specific growth rate of population N_2 or by altering the operating conditions i.e. by shifting the C/N ratio in the feed. In the present work the result of amensalism is the occurrence of coexistence state against the ability of N_1

population to survive alone at stable steady state. The threshold of dilution rate, above which only N_2 population survives, exists also in this case. When the inhibition on N_2 population is strong then multistability state is favored towards coexistence state. The N_1 population can survive at stable steady state, depending on the initial conditions. Comparing to Kavadia *et al.* [41], the major difference observed is the narrow range of the regions in which N_1 population can survive either alone or in coexistence with N_2 . Amensalism gives the ability to N_1 to survive under high NH_3 concentration, but the low growth rate, in regard to previous work, constrains the population at low dilution rate values. In the case of *DvsS*, N_1 population is at stable steady state at very low dilution rate values, while it can survive in coexistence with N_2 at low D values and high S concentrations.

In the case of amensalism N_1 population is able to survive in stable steady state at low carbon substrate concentration. Specifically, multistability is observed (N_1 state and N_2 state), while at very low carbon concentration values only N_2 population survives at stable steady state. At high concentration of the carbon source, N_1 population will survive either alone or in coexistence with its competitor in stable steady state (multistability). Also at conditions of strong inhibition on N_2 population the coexistence state is diminished for the benefit of N_1 survival at steady state.

The nitrogen fixation rate is also affected by the lower growth rate of N_1 population. Although N_1 can survive in a stable steady state at high NH_3 concentration, with the help of N_2 , it fixes nitrogen only at low NH_3 concentrations while the nitrogen fixation rate is conversely proportional to NH_3 concentration and to dilution rate values. A possible effect of the lower growth rate of N_1 population on nitrogen fixation rate would be presented similar to the results of Kavadia *et al.* [42], but at lower dilution rate values.

CONCLUSIONS

A nitrogen-fixing population growing alone in a chemostat can survive at stable steady state only at low ammonia concentrations and dilution rate values. The presence of a second population, which has the competitive advantage over N_1 population, induces major changes in the system. The combination of ammonia inhibition and competition lead to the constraint of N_1 population at low dilution rate values. When the nitrogen-fixing population is able to establish an amensalistic interaction in the chemostat, the regions where the N_1 population survives alone at stable steady state are reduced and the coexistence region is favoured. When the inhibition on the competitive population is strong then the coexistence state is disadvantaged and multistability state is favoured.

DISCLOSURE

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REFERENCES

- [1] J.R. Postgate, "The Fundamentals of Nitrogen Fixation", Cambridge University Press, London, 1982.

- [2] J.E. Bailey, and D.F. Ollis, "Biochemical Engineering Fundamentals", 2nd ed. McGraw- Hill Int., Singapore, 1986.
- [3] R.M. Atlas, and R. Bartha, "Microbial Ecology: Fundamentals and Applications", 3rd ed. Addison-Wesley, Reading, MA, 1993.
- [4] I. Cacciari, M. Del Gallo, S. Ippoliti, D. Lippi, T. Pietrosanti, and W. Pietrosanti, "Growth and survival of azospirillum brasilense and arthrobacter giacomelloi in binary continuous culture", *Plant Soil*, vol. 90, pp. 107-116, 1986.
- [5] R.V. Solé, J. Bascompte, and J. Valls, "Stability and complexity of spatially extended two-species competition", *J. Theor. Biol.*, vol. 159, pp. 469-480, 1992.
- [6] A. Roberts, and L. Stone, "Advantageous indirect interactions in systems of competition", *J. Theor. Biol.*, vol. 228, pp. 367-375, 2004.
- [7] U. Bastolla, M. Lassig, S.C. Manrubia, and A. Valleriani, "Biodiversity in model ecosystems, I: coexistence conditions for competing species", *J. Theor. Biol.*, vol. 235, pp. 521-530, 2005.
- [8] Y. Bashan, and G. Holguin, "Azospirillum plant relationships : environmental and physiological advances (1990-1996)", *Can. J. Microbiol.*, vol. 43, pp. 103-121, 1997.
- [9] D.A. Zuberer, "Biological Dinitrogen Fixation: Introduction and Nonsymbiotic" In Principles and Applications of Soil Microbiology ed. Sylvia, D.M., Fuhrmann, J.J., Hartel, P.J. and Zuberer, D.A, New Jersey: Prentice Hall, 1998, pp. 295-321
- [10] Y. Bashan, G. Holguin, and L.E. de-Bashan, "Azospirillum-plant relationships : physiological, molecular, agricultural, and environmental advances (1997-2003)", *Can. J. Microbiol.*, vol. 50, pp. 521-577, 2004.
- [11] M.G. Hassouna, M.A.M. El-Saedy, and H.M.A. Saleh, "Biocontrol of soil-borne plant pathogens attacking cucumber (*Cucumis sativus*) by rhizobacteria in a semiarid environment", *Arid Soil Res. Rehabil.*, vol. 12, pp. 345-357, 1998.
- [12] K. Kavitha, K.S. Meenakumari, and P. Sivaprasad, "Effect of dual inoculation of native arbuscular mycorrhizal fungi and Azospirillum on suppression of damping off in chilli", *Ind. Phytopathol.*, vol. 56, pp. 112-113, 2003.
- [13] A.M. Romero, O.S. Correa, S. Moccia, and J.G. Rivas, "Effect of Azospirillum – mediated plant growth promotion on the development of bacterial diseases on fresh-market and cherry tomato", *J. Appl. Microbiol.*, vol. 95, pp. 832-838, 2003.
- [14] L.M. Mallory, C.-S. Yuk, L.-N. Liang, and M. Alexander, "Alternative prey: a mechanism for elimination of bacterial species by protozoa", *Appl. Environ. Microbiol.*, vol. 46, pp. 1073-1079, 1983.
- [15] J.J. Germida, "Isolation of Bdellovibrio spp. that prey on Azospirillum brasilense in soil", *Can. J. Microbiol.*, vol. 33(5), pp. 459-461, 1987.
- [16] R. Rønn, J. Grunert, and F. Ekelund, "Protozoan response to addition of the bacteria *Mycobacterium chlorophenolicum* and *Pseudomonas chlororaphis* to soil microcosms", *Biol. Fertil. Soils.*, vol. 33, pp. 126-131, 2001.
- [17] Y. Davidov, D. Huchon, S.F. Koval, and E. Jurkevitch, "A new apoteobacterial clade of Bdellovibrio-like predators : implications for the mitochondrial endosymbiotic theory", *Environ. Microbiol.*, vol. 8, no. 12, pp. 2179-2188, 2006.
- [18] A.M. Rogosky, P.L. Moak, and E.A.B. Emmert, "Differential Predation by Bdellovibrio bacteriovorus 109J", *Curr. Microbiol.*, vol. 52(2), pp. 81-85, 2006.
- [19] N.S. Strigul, and L.V. Kravchenko, "Mathematical modeling of PGPR inoculation in the rizosphere", *Environ. Modell. Softw.*, vol. 21, pp. 1158-1171, 2006.
- [20] R.V. Klucas, "Associative Nitrogen Fixation in Plants", In Biology and biochemistry of nitrogen fixation, M.J. Dilworth, A.R. Glenn, Eds. Elsevier: Amsterdam, 1991.
- [21] O. Steenhoudt, and J. Vanderleyden, "Azospirillum, a free-living nitrogen-fixing bacterium closely associated with grasses: genetic, biochemical and ecological aspects", *FEMS Microbiol. Rev.*, vol. 24, pp. 487-506, 2000.
- [22] Y. Okon, S.L. Albrecht, and R.H. Burris, "Factors affecting growth and nitrogen fixation of Spirillum lipoferum", *J. Bacteriol.*, vol. 127, pp. 1248-1254, 1976.
- [23] T. Hurek, B. Reinhold, and E.G. Niemann, "Effect of oxygen on NH₄⁺-grown continuous cultures of Azospirillum spp. and diazotrophic rods closely associated with Kallar grass". *Can. J. Microbiol.*, vol. 33, pp. 919-922, 1987.
- [24] V. Tsagou, I. Kefalogianni, K. Sini, and G. Aggelis, "Metabolic activities in Azospirillum lipoferum grown in the presence of NH₄⁺", *Appl. Microbiol. Biotechnol.*, vol. 62, pp. 574-578, 2003.
- [25] V. Tsagou, and G. Aggelis, "Growth dynamics of Azospirillum lipoferum at steady and transitory states in the presence of NH₄⁺", *J. Appl. Microbiol.*, vol. 100, pp. 286-295, 2006.
- [26] C. Fritzsche, J. Ueckert, and E.-J. Niemann, "Growth parameters of microaerobic diazotrophic rhizobacteria determined in continuous culture". In Nitrogen Fixation M. Polsinelli, R. Materassi, and M. Vincenzini, Eds. Dordrecht, Boston, London: Kluwer Academic Publishers, 1990, pp. 232-234.
- [27] C. Fritzsche, and E.G. Niemann, "Nitrogen fixation in continuous culture with NH₄Cl-containing media", *Appl. Environ. Microbiol.*, vol. 56, pp. 1160-1161, 1990.
- [28] I. Kefalogianni, and G. Aggelis, "Modelling growth and biochemical activities of Azospirillum spp.", *Appl. Microbiol. Biotechnol.*, vol. 58, pp. 352-357, 2002.
- [29] H.L. Smith, and P. Waltman, "The Theory of the Chemostat: Dynamics of Microbial Competition", Cambridge studies in mathematical biology, Cambridge University Press, Cambridge, 1995, Vol. 13.
- [30] M.A. Leibold, "A graphical model of keystone predation: effects of productivity on abundance, incidence and ecological diversity in communities", *Am. Nat.*, vol. 147, pp. 784-812, 1996.
- [31] V. Krivan, and A. Sikder, "Optimal foraging and predator-prey dynamics II", *Theor. Popul. Biol.*, vol. 55, pp. 111-126, 1999.
- [32] B.W. Kooi, and S.A.L.M. Kooijman, "Invading species can stabilize simple trophic systems", *Ecol. Model.*, vol. 133, pp. 57-72, 2000.
- [33] J.D. Pelletier, "Are large complex ecosystems more unstable? A theoretical reassessment with predator switching", *Math. Biosci.*, vol. 163, pp. 91-96, 2000.
- [34] S.A Richards, R.G. Nisbet, W.G. Wilson, and H.P. Possingham, "Grazers and diggers: exploitation competition and coexistence among foragers with different feeding strategies on a single resource", *Am. Nat.*, vol. 155, pp. 266-279, 2000.
- [35] V. Krivan, and J. Eisner, "Optimal foraging and predator-prey dynamics III", *Theor. Popul. Biol.*, vol. 63, pp. 269-279, 2003.
- [36] V. Krivan, "Competitive co-existence caused by adaptive predators", *Evol. Ecol. Res.*, Vol. 5, pp. 1163-1182, 2003.
- [37] G. Aggelis, D.V. Vayenas, V. Tsagou, and S. Pavlou, "Prey-predator dynamics with predator switching regulated by a catabolic repression control mode", *Ecol. Mod.*, vol. 183, pp. 453-464, 2005.
- [38] D.V. Vayenas, G. Aggelis, V. Tsagou, and S. Pavlou, "Dynamics of a two-prey-one-predator system with predator switching regulated by a catabolic repression control-like mode", *Ecol. Mod.*, vol. 186, pp. 345-357, 2005.
- [39] Y. Pei, L. Chen, Q. Zhang, and C. Li, "Extinction and permanence of one-prey multi-predators of Holling type II function response system with impulsive biological control", *J. Theor. Biol.*, vol. 235, pp. 495-503, 2005.
- [40] Z. Zhang, "Mutualism or cooperation among competitors promotes coexistence and competitive ability", *Ecol. Mod.*, vol. 164, pp. 271-282, 2003.
- [41] A. Kavadia, D.V. Vayenas, S. Pavlou, and G. Aggelis, "Dynamics of free-living nitrogen-fixing bacterial populations in antagonistic conditions", *Ecol. Model.*, vol. 200, pp. 243-253, 2007.
- [42] Kavadia, D.V. Vayenas, S. Pavlou, and G. Aggelis, "Dynamics of free-living nitrogen-fixing bacterial populations and nitrogen fixation in a two-prey-one-predator system", *Ecol. Model.*, vol. 218, pp. 323-338, 2008.
- [43] A.G. Fredrickson, "Formulation of structured growth models. (Communication to the editor)", *Biotechnol. Bioeng.*, vol. 18, pp. 720-725, 1976.
- [44] J. Monod, "Research on the growth of bacterial cultures", *Act. Sci. Ind.*, vol. 911, pp. 1-215, 1942 (in French).
- [45] S.P. Graef, and J.F. Andrews, "Mathematical modelling and control of anaerobic digestion", *CEP Symposium Series*, vol. 70, pp. 101-127, 1974.
- [46] E.J. Doedel, A.R. Champneys, T.F. Fairgrieve, Y.A. Kuznetsov, B. Sandstede, and X. Wang, "AUTO 97: Continuation and Bifurcation Software for Ordinary Differential Equations, User's manual", Centre for Research on Parallel Computing, California Institute of Technology, Pasadena, 1997.

- [47] J. Ueckert, and I. Fendrik, "Continuous Culture Applications in Physiological Investigations on Diazotrophic Bacteria", In I. Fendrik, M. Del Gallo, J. Vanderleyden, M. De Zamaroczy, Eds. *Azospirillum VI and related microorganisms*, vol. G37, Springer-Verlag, Heidelberg, 1995.
- [48] E. Bahat-Samet, S. Castro-Sowinski, and Y. Okon, "Arabinose content of extracellular polysaccharide plays a role in cell aggregation of *Azospirillum brasilense*", *FEMS Microbiol. Lett.*, vol. 237, pp. 195-203, 2004.
- [49] D. van Dam, and N. van Breemen, "NICCE: a model for cycling of nitrogen and carbon isotopes in coniferous forest ecosystems", *Ecol. Model.*, vol. 79, pp. 255-275, 1995.
- [50] R.R. Christian, E. Forés, F. Comin, P. Viaroli, M. Naldi, and I. Ferrari, "Nitrogen cycling networks of coastal ecosystems: influence of trophic status and primary producer form". *Ecol. Model.*, vol. 87, pp. 111-129, 1996.
- [51] A. Chapelle, A. Ménesguen, J.-M. Deslous-Paoli, P. Souchu, N. Mazouni, A. Vaquer, and B. Millet, "Modelling nitrogen, primary production and oxygen in Mediterranean lagoon. Impact of oysters farming and inputs from the watershed", *Ecol. Model.*, Vol. 127, pp. 161-181, 2000.
- [52] J.R. Kercher, and J.Q. Chambers, "Parameter estimation for a global model of terrestrial biogeochemical cycling by an iterative method", *Ecol. Model.*, vol. 139, pp. 137-175, 2001.
- [53] M. Corbeels, R.E. McMurtrie, D.A. Pepper, and A.M. O'Connell, "A process-based model of nitrogen cycling in forest plantations. Part II. Simulating growth and nitrogen mineralization of *Eucalyptus globulus* plantations in south-western Australia", *Ecol. Model.*, vol. 187, pp. 449-474, 2005.
- [54] L.C. Bruce, D. Hamilton, J. Imberger, G. Gal, M. Gophen, T. Zohary, and K.D. Hambright, "A numerical simulation of the role of zooplankton in C, N and P cycling in Lake Kinneret, Israel", *Ecol. Model.*, vol. 193, pp. 412-436, 2006.
- [55] B.E. McNeil, R.E. Martell, and J.M. Read, "GIS and biogeochemical models for examining the legacy of forest disturbance in the Adirondack Park, NY, USA", *Ecol. Model.*, vol. 195, pp. 281-295, 2006.
- [56] M. Kloss, K.H. Iwannek, I. Fendrik, and E.G. Niemman, "Enrichment of diazotrophic bacteria from rice soil in continuous culture", *Plant Soil*, vol. 90, pp. 151-164, 1986.
- [57] M. Mar Vázquez, S. César, R. Azcón, and J.M. Barea, "Interactions between arbuscular mycorrhizal fungi and other microbial inoculants (*Azospirillum*, *Pseudomonas*, *Trichoderma*) and their effects on microbial population and enzyme activities in the rhizosphere of maize plants", *Appl. Soil Ecol.*, vol. 15, pp. 261-272, 2000.

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