Effects of Light With Reduced Short Wavelength Components on Parameters of Circadian Rhythm and Performance in an Experimental Night Shift Model

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Abstract: Shift work is associated with alterations in physiological circadian patterns resulting in chronic diseases, e.g. cardiovascular disorders or major depression. The intensity and spectral composition of light is known to affect the 24 h-rhythm of our body. We investigated the effects of two different lighting environments on parameters of circadian rhythm and performance in healthy volunteers during an experimental night shift. Test light with a low color temperature was compared to normal light with a higher color temperature. Melatonin synthesis, red and white blood count, blood pressure, heart rate and indicators of performance were analyzed. Nocturnal increases in melatonin were more pronounced under low color temperature lighting conditions. This was not associated with limited degrees of arousal or vigilance. Maintenance of a normal nocturnal rhythm of melatonin with adapted illumination may provide a benefit for employees well-being without affecting their productivity.

INTRODUCTION

An increasing number of employees in industrialized nations perform night shift work on a regular basis. Among the risks associated with night shift jobs are increased error and accident rates as well as production of substandard goods [1,2]. While the deliberate disturbance of a physiological circadian rhythm is supposed to be involved in the pathogenesis of cardiovascular disorders or major depression [3,4], the evidence connecting shift work and an increased incidence of certain types of tumors, e.g. colorectal and breast cancer, is discussed controversially [5-7]. Although the exact mechanisms underlying these detrimental processes are unclear at present, nocturnal bright light exposure is regarded to be one decisive factor in the development of these harmful events. Circadian rhythms can be found in every cell of our body. The cellular oscillations are coordinated via superior centres in the suprachiasmatic nucleus (SCN) representing the master clock of the human organism. The activities of the SCN are synchronized by exogenous factors appearing in a periodical fashion with the natural light-dark cycle being the most prominent Zeitgeber. Light signals are most likely received by melanopsin-containing ganglion cells already described in the retina of rodents [8], primates [9], and humans [10]. The mode of action and the sensitivity of these cells were primarily investigated in connection with the light-induced suppression of melatonin synthesis. Melatonin is a hormone produced in the pineal gland. Synthesis and release of melatonin into the blood stream oscillates in a circadian pattern with the highest concentrations found late at night (at ~ 04:00 a.m.) and decreasing values in the course of the day reaching a nadir at ~ 04:00 p.m. [11]. Inhibition of melatonin release triggered by light was first described by Klein and Weller [12] in rodents while Lewy et al. [13] detected a corresponding effect of light on melatonin in humans. A light intensity of 100 lux seems to be efficient to attenuate pineal synthesis of the hormone [14]. Besides its intensity, the spectral composition of light exerts a substantial impact on melatonin generation. The action spectrum of melanopsin-containing ganglion cells exhibit a maximal photosensitivity at ~ 460 nm [15,16]. Therefore, the application of filtered light with reduced short wavelength components (i.e. low color temperature) represents a potential concept for the mitigation or avoidance of light-induced detrimental consequences during night shift. The aim of the present study was to investigate the effects of two differently illuminated office workplaces on melatonin synthesis in an experimental night shift setup. Light with a low color temperature (1700 K at eye level) was compared to light with a higher color temperature (6300 K at eye level). In addition to melatonin, other physiological markers following a circadian pattern (heart rate, blood pressure, white and red blood count) were investigated. Ergonomical interventions aiming at the maintenance of a biological circadian profile can only be successful on the condition that they do not interfere with the employees productivity. Thus, a number of tests were performed to elucidate the effects of the different...
lighting environments on the efficiency of the study participants.

MATERIALS AND METHODOLOGY

Study Participants and Study Design

11 healthy male volunteers (age: 23 (range: 22-29) years; body weight: 74 (range: 62-84) kg; height: 177 (range: 172-181) cm) without any known metabolic, cardiovascular, renal, hepatic, neurological, psychiatric or sleeping disorders requiring acute or chronic medication participated in our study. Test persons had not undertaken shift work or transmeridian travel in the month prior to the study. Subjects were instructed to refrain from excessive physical exercise, alcohol consumption, and over-the-counter medication during the course of the study. At the beginning of every night of the study, a sleeping protocol of the preceding night was recorded. No exceptional sleeping behaviour was reported. The project was approved by the Ethics Committee of the Medical University of Innsbruck, and all participants gave written informed consent. The study was performed at the Bartenbach LichtLabor, Aldrans, Austria following a cross-over design with three consecutive days of experimental office work in two different lighting environments but otherwise constant workplace conditions (area: 20 m²; ceiling height: 2.6 m; temperature: 23°C ± 2°C regulated via air handling unit, environmental luminance: 300 cd/m²). The volunteers were randomly assigned to start with three consecutive nights of simulated shift work either in an unfiltered bright light environment (luminaire: wallwasher, color temperature 6300 K at eye level; color rendering index: 89) or in a lighting environment with reduced short-wavelength components (luminaire: wallwasher, color temperature 1700 K at eye level; color rendering index: 80). Following an 11-day break, the study participants conducted a second run of three consecutive night shifts in the other lighting environment. Each night shift started at 10:00 p.m. at night and ended at 06:00 a.m. in the morning.

Laboratory Measurements

At the beginning (22:00 p.m., labeled B1 throughout the manuscript) and at the end (06:00 a.m., labeled B2 throughout the manuscript) of each night of the study, blood samples were taken from an antecubital vein and processed at the Central Institute for Medical and Chemical Laboratory Diagnostics, Innsbruck, Austria. Erythrocyte count, hemoglobin, hematocrit, and white blood cell count were measured by standard methods.

Serum melatonin was assayed using a commercially available radio-immunoassay kit (Buehlmann Melatonin direct RIA, Buehmann Laboratories AG, Schoenenbuch, Switzerland) with an analytical sensitivity of 0.84 pg melatonin/ml. In brief, samples, assay controls, and melatonin in human serum matrix as calibrators were incubated with 125I-labeled melatonin and a specific rabbit anti-melatonin antibody for 20 h at 4°C in polystyrene tubes. Subsequently, solid phase bound anti-rabbit second antibody was added into each tube and incubated for an additional 15 min at 4°C to precipitate the antibody-bound fraction. Finally, ultrapure water was added and the tubes were centrifuged at 2000 g for 2 min at room temperature. Supernatants were discarded and 125I activity in the precipitates was detected for 2 min in a γ-counter. Unknown samples were calculated using a standard curve generated by the calibrators provided by the manufacturer. All measurements were performed in duplicate. Data are given as pg melatonin/ml. All melatonin analyses were done at the Central Institute for Medical and Chemical Laboratory Diagnostics, Innsbruck, Austria.

Blood Pressure and Heart Rate

Blood pressure and heart rate were recorded using an automatic device (Omron M5, Omron Medizintechnik Handelsgesellschaft mbH, Mannheim, Germany). First data were collected at the beginning of the experimental night shift. Subsequently, a number of six measurements were performed between the ability testings adding up to a total of seven recordings per night labeled M1 to M7. Approximate time points for each measurement were: M1 at 10:30 p.m.; M2 at 11:30 p.m.; M3 at 0:15 a.m.; M4 at 1:45 a.m.; M5 at 2:45 a.m.; M6 at 3:30 a.m.; and M7 at 5:30 a.m., respectively. Mean values of three readings were used for statistical analyses.

Ability Test Systems

Simulated office work consisted of general and special ability tests all being part of the Vienna Test System® (Dr. G. Schuhfried Ltd., Moedling, Austria). The tests were performed twice each night with the first test being labeled A1 and the second being labeled A2 throughout the manuscript.

Flicker/Fusion Frequency

In order to evaluate central nervous activation or arousal, flicker/fusion frequency analyses were performed at 10:15 p.m. and at 5:30 a.m. The test is divided into two parts. At first, frequency of flickering light is increased until a constant light is perceived (“fusion frequency”). In the second part of the experiment, the frequency of constant light is decreased until the subject senses flickering light again (“flicker frequency”). Both transitions are captured by the study participants via pressing a key at the flicker/fusion device. The critical frequency is stored and threshold values are calculated from eight consecutive measurements. Total running time of this test was 10 min. Abbreviations for evaluation scores used in the manuscript are FUF for “fusion frequency” and FLF for “flicker frequency”, respectively.

Continuous Attention

Long-term selective attention and concentration ability were assessed twice per night by the continuous attention test at 11:00 p.m. and at 3:00 a.m. Rows of triangles were presented on a computer monitor with their tips pointing either up or down. A fixed number of tips pointing down had to be responded by the study participants by pushing a reaction button on the keyboard. Duration of the test was 35 min. For analyses, the sum of correct hits (SUMC) was calculated. Auxiliary parameters were the sum of incorrect hits (SUMI), the mean time for correct hits (MEANC), and the mean time for incorrect hits (MEANI).

Reaction Time Analyses

Potential slow-downs of cognitive speed were assessed by monitoring the three stages of activity regulation (discriminative perception, cognitive processing, and motor response organization) at 11:30 p.m. and at 3:00 a.m., respec-
tively. Discrimination of figures (circle, rectangle, square, star, cross, ellipse) on a computer monitor was aggravated by partial covering with a grid. Variation of the cognitive processing stage was realized by the search for one or two of two simultaneously presented figures. Variation of motor response organization was achieved by modifying the complexity of the reaction to be executed (either with one finger or with a sequence of three keys). Subjects were guided through the experiments interactively. 14 series with either 20 individual stimuli, each for choice reaction tasks or 16 individual stimuli for visual search tasks, were processed. A total running time of 25 min was required. Scores are expressed as information processing (IP), perception (PER), and motor response organization (MRO). Additional scoring parameters were choice reaction time (CRT) and reaction time for visual search (VSRT).

**Vigilance Testing**

Subjects were required to maintain continuous attention although the working environment became stale and signals appeared only randomly and did not cause agitation. To detect continuous vigilance, stimuli with a low intensity and a low frequency of critical events were provided. A flashing dot on the computer monitor travelling in a circular pattern in distinct intervals had to be observed. At random, the interval was doubled, which had to be recorded by the study participants *via* pressing a button on the keyboard. Vigilance testing was performed at midnight and at 3:30 a.m. Total running time of the test was 70 min. Main scoring parameter for analyses was the number of correct hits (NUMC), auxiliary values included number of incorrect hits (NUMI), and mean value of reaction time for correct hits (MRTC).

**Statistical Analyses**

To compare between three or more matched groups with repeated measures Friedman tests were performed. To exactly compare two paired groups, we used the Wilcoxon matched pairs test. To test whether two variables varied together, a nonparametric Spearman correlation was calculated. For presentation of descriptive data, box and whiskers plots were used. The box represents the median together with the 25th and 75th quartile, while the whiskers show the highest and lowest values, respectively. P-values < 0.05 were considered to be significant.

**RESULTS**

Both lighting environments were tolerated by the study participants without adverse effects and the experiments were completed by all volunteers. Fig. (1) summarizes the results of melatonin measurements performed in serum samples obtained at the beginning (B1) and at the end (B2) of each night of the study either in the room with filtered test light (room T, Fig. 1A) or in the room with full-spectrum normal light (room N, Fig. 1B). In room T, melatonin concentrations were significantly increased at B2 as compared to B1 on the 2nd and 3rd night of the study, while in room N a significant change could only be observed at night 2. To gain further insight into the potentially distinct effects of the different illuminations on melatonin synthesis, the differences in its serum concentrations between B2 and B1 were calculated and compared between the two lighting conditions. ΔB2B1 values were significantly lower in room N at night 1 and 2 of the study when compared to the respective data obtained in room T. This stabilizing effect of filtered light on melatonin generation was confirmed by direct comparison of serum melatonin concentrations obtained from samples in room T and room N, respectively. While no differences in melatonin levels between both office accommodations could be found in specimen collected at the beginning of each night of the study, melatonin was significantly lower in room N at B2 when compared to the corresponding values in room T at every night of the experiments.

With respect to red blood count, no significant differences between B2 and B1 could be found under both lighting conditions at any night of the study (Table 1). Erythrocyte count as well as hemoglobin concentrations and hematocrit remained unchanged in the course of the study. It should be noted that in all study participants the corresponding values were within the normal range. Regarding white blood count, we observed significant increases in the percentage of lymphocytes and eosinophils when comparing B2 with B1 at every night of the study that were independent of the illumi-
In contrast, no significant changes were observed in the of-
cal comparison of these differences between room T and
room N did not point out a discriminative effect of the light-
vation. In contrast, percentage of segmented neutrophils re-
vealed significant depletions in the course of each night of
the experiments in both filtered and full-spectrum office en-
vironments (Table 1). Again, ΔB2B1 values were calculated
for each parameter and for every night of the study. Statis-
tical comparison of these differences between room T and
room N. Regarding the auxiliary parameters, MRTC was significantly higher in both
rooms at A2 on the first night of the study, and NUMI was
significantly higher at A2 in both rooms on the second
night when compared to the initial values obtained at A1. Cognitive speed (process-
ing, perception, and motor response organization for choice
reaction tasks) were determined via Reaction Time Analyses
at 11:30 p.m. (A1) and at 3:00 a.m. (A2). Taken together, the
main scoring parameters IP, PER, MRO as well as CRT re-
main unchanged throughout the experiments and were
independent of the illumination. Concerning VSRT, we ob-
served a significant decrease at A2 on the third night in the
room with unfiltered bright light when comparing to the re-
spective data at A1. Vigilance tests were scheduled at mid-
night (A1) and at 3:30 a.m. (A2) in order to evaluate atten-
tion of study participants under continuous stress. When
compared to A1 values, the major parameter NUMC turned
out to be significantly reduced at the end of night 1 and 2
in room T, but only at night 1 in room N. With respect to auxili-
ary parameters, MRTC was significantly higher in both
rooms at A2 on the first night of the study, and NUMI was
increased at A2 as compared to A1 at night 2 in room N.

**DISCUSSION**

Melatonin is one of the best studied circadian parameters
in humans. The 24 h-profile of melatonin shows a peak at
approximately 4:00 a.m. Since melatonin production de-
clines with age, we have chosen to enroll volunteers belong-
ing to an age group that still has a high melatonin production
capability as well as a low inter-individual variability in
melatonin synthesis [17]. In our study, serum melatonin
concentrations were highest at the second time point of blood
Fig. (2). Time course of systolic blood pressure on the first, second, and third night either in a lighting environment with reduced short wavelength components (T1, T2, and T3; Figs. A, B, and C, respectively) or in an unfiltered bright light environment (N1, N2, and N3; Figs. D, E, and F, respectively). Time points for each measurement were: M1 at 10:30 p.m.; M2 at 11:30 p.m.; M3 at 0:15 a.m.; M4 at 1:45 a.m.; M5 at 2:45 a.m.; M6 at 3:30 a.m.; and M7 at 5:30 a.m. Data are given as median together with the 25th and the 75th quartile, whiskers represent highest and lowest values. Friedman tests for matched groups with repeated measures were performed to analyze potential time effects, resulting P-values are given in the corresponding panel.

Sample collection, i.e. at 6:00 a.m., which is in congruence with the reported circadian rhythm of the hormone. With respect to the different illuminations, significantly lower melatonin values in unfiltered light as compared to light with reduced short wavelength components found in our study confirms previous observations establishing bright light as a potent suppressor of pinealocyte melatonin synthesis. Thus, altered diurnal melatonin profiles are a regular finding among shift workers [18,19], albeit there seems to be a capacity to adapt to a pattern of shift work with a higher number of consecutive nights and a fixed sleep schedule during the days [20]. Comparable to the preserving effect of filtered light on melatonin generation observed in our experiments, Kayumov et al. [21] reported a normal night time profile of melatonin in a simulated shift work model in participants wearing light-filtering goggles that blocked wavelengths of less than 530 nm. With respect to the key parameter melatonin, the present study provides evidence that the benefits of
Fig. (3). Time course of diastolic blood pressure on the first, second, and third night either in a lighting environment with reduced short wavelength components (T1, T2, and T3; Figs. A, B, and C, respectively) or in an unfiltered bright light environment (N1, N2, and N3; Figs. D, E, and F, respectively). Time points for each measurement were: M1 at 10:30 p.m.; M2 at 11:30 p.m.; M3 at 0:15 a.m.; M4 at 1:45 a.m.; M5 at 2:45 a.m.; M6 at 3:30 a.m.; and M7 at 5:30 a.m. Data are given as median together with the 25th and the 75th quartile, whiskers represent highest and lowest values. Friedman tests for matched groups with repeated measures were performed to analyze potential time effects, resulting P-values are given in the corresponding panel.
reducing short-wavelength light can be achieved in an almost natural office accommodation, too.

Erythrocyte count, hematocrit, and hemoglobin concentration remained unchanged throughout the experiments and were not influenced by the different lighting environments. An increase in hematocrit and hemoglobin concentrations with average morning values being significantly higher than the respective data of the evening before have been reported in the literature [22,23]. Since these changes were due to nocturnal reductions in plasma volume, we conclude that our study participants kept fluid loss and fluid replacement in balance in the course of the study. Nocturnal increases in eosinophils and lymphocytes as well as decreases in segmented neutrophils observed in our study might be due to circadian variations in glucocorticosteroid synthesis, predominantly cortisol. The physiological 24 h-profile of cortisol establishes minimal serum values in the evening and a surge in the early morning hours. Glucocorticosteroids have been considered to regulate immune cell systems through modulation of apoptotic cell death. Thus, experimentally
Table 2. Results of Ability Test Systems (Vienna Test System®, Dr. G. Schuhfried Ltd., Moedling, Austria). Data are Given as Mean ± SEM

<table>
<thead>
<tr>
<th></th>
<th>Night 1 A1</th>
<th>Night 1 A2</th>
<th>Night 2 A1</th>
<th>Night 2 A2</th>
<th>Night 3 A1</th>
<th>Night 3 A2</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Filtered Light</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Flicker Frequency</td>
<td>43.3 ± 1.9</td>
<td>38.9 ± 0.8*</td>
<td>41.2 ± 0.7</td>
<td>37.8 ± 0.9*</td>
<td>39.9 ± 0.9</td>
<td>38.4 ± 0.9*</td>
</tr>
<tr>
<td>Unfiltered Light</td>
<td>40.6 ± 0.8</td>
<td>38.2 ± 1.0*</td>
<td>40.6 ± 0.9</td>
<td>38.8 ± 0.9*</td>
<td>40.4 ± 0.9</td>
<td>39.1 ± 0.9*</td>
</tr>
<tr>
<td>Fusion Frequency</td>
<td>39.0 ± 0.8</td>
<td>38.5 ± 0.9</td>
<td>40.0 ± 0.9</td>
<td>39.0 ± 0.9</td>
<td>40.5 ± 0.8</td>
<td>39.5 ± 0.7</td>
</tr>
<tr>
<td>Unfiltered Light</td>
<td>39.3 ± 0.7</td>
<td>37.6 ± 0.8</td>
<td>39.7 ± 0.8</td>
<td>38.7 ± 0.8</td>
<td>39.7 ± 0.8</td>
<td>38.7 ± 0.8</td>
</tr>
<tr>
<td>Continuous Attention SUMC</td>
<td>268 ± 4</td>
<td>258 ± 5</td>
<td>271 ± 2</td>
<td>250 ± 15*</td>
<td>268 ± 3</td>
<td>260 ± 10</td>
</tr>
<tr>
<td>Unfiltered Light</td>
<td>266 ± 5</td>
<td>252 ± 6*</td>
<td>270 ± 4</td>
<td>264 ± 4*</td>
<td>269 ± 2</td>
<td>259 ± 8</td>
</tr>
<tr>
<td>Continuous Attention MEANC</td>
<td>0.73 ± 0.02</td>
<td>0.74 ± 0.02</td>
<td>0.70 ± 0.02</td>
<td>0.73 ± 0.02</td>
<td>0.71 ± 0.02</td>
<td>0.70 ± 0.02</td>
</tr>
<tr>
<td>Unfiltered Light</td>
<td>0.72 ± 0.02</td>
<td>0.75 ± 0.02*</td>
<td>0.70 ± 0.02</td>
<td>0.72 ± 0.02</td>
<td>0.75 ± 0.02</td>
<td>0.72 ± 0.02*</td>
</tr>
<tr>
<td>Continuous Attention SUMI</td>
<td>17.5 ± 4.8</td>
<td>19.0 ± 3.7</td>
<td>10.5 ± 1.7</td>
<td>13.1 ± 2.0*</td>
<td>10.2 ± 1.6</td>
<td>14.1 ± 1.6</td>
</tr>
<tr>
<td>Unfiltered Light</td>
<td>11.4 ± 2.1</td>
<td>16.4 ± 3.1</td>
<td>10.2 ± 1.8</td>
<td>14.7 ± 2.7*</td>
<td>9.7 ± 2.0</td>
<td>14.6 ± 2.6*</td>
</tr>
<tr>
<td>Continuous Attention MEANI</td>
<td>0.76 ± 0.03</td>
<td>0.76 ± 0.03</td>
<td>0.70 ± 0.03</td>
<td>0.76 ± 0.03*</td>
<td>0.67 ± 0.04</td>
<td>0.72 ± 0.03</td>
</tr>
<tr>
<td>Unfiltered Light</td>
<td>0.74 ± 0.03</td>
<td>0.78 ± 0.02</td>
<td>0.73 ± 0.03</td>
<td>0.74 ± 0.03*</td>
<td>0.68 ± 0.03</td>
<td>0.71 ± 0.04</td>
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<tr>
<td>Reaction Time Analyses IP</td>
<td>302 ± 31</td>
<td>263 ± 34</td>
<td>252 ± 17</td>
<td>277 ± 26</td>
<td>252 ± 25</td>
<td>253 ± 38</td>
</tr>
<tr>
<td>Unfiltered Light</td>
<td>273 ± 39</td>
<td>322 ± 38</td>
<td>256 ± 32</td>
<td>249 ± 40</td>
<td>257 ± 33</td>
<td>231 ± 26</td>
</tr>
<tr>
<td>Reaction Time Analyses PER</td>
<td>27.3 ± 9.8</td>
<td>6.8 ± 16.0</td>
<td>9.4 ± 11.8</td>
<td>-10.3 ± 15.3</td>
<td>29.0 ± 16.0</td>
<td>22.3 ± 9.9</td>
</tr>
<tr>
<td>Unfiltered Light</td>
<td>4.8 ± 11.7</td>
<td>16.8 ± 16.8</td>
<td>29.0 ± 16.7</td>
<td>16.5 ± 16.5</td>
<td>13.1 ± 13.1</td>
<td>4.7 ± 4.7</td>
</tr>
<tr>
<td>Reaction Time Analyses MRO</td>
<td>22.9 ± 42.7</td>
<td>51.0 ± 32.9</td>
<td>36.1 ± 51.8</td>
<td>15.3 ± 32.5</td>
<td>-39.8 ± 34.5</td>
<td>40.1 ± 31.2</td>
</tr>
<tr>
<td>Unfiltered Light</td>
<td>11.9 ± 33.0</td>
<td>-6.9 ± 31.7</td>
<td>37.8 ± 30.3</td>
<td>49.0 ± 21.7</td>
<td>-27.3 ± 24.7</td>
<td>7.4 ± 32.4</td>
</tr>
<tr>
<td>Reaction Time Analyses CRT</td>
<td>582 ± 55</td>
<td>568 ± 58</td>
<td>568 ± 57</td>
<td>569 ± 50</td>
<td>543 ± 38</td>
<td>522 ± 40</td>
</tr>
<tr>
<td>Unfiltered Light</td>
<td>616 ± 57</td>
<td>604 ± 47</td>
<td>563 ± 54</td>
<td>566 ± 59</td>
<td>545 ± 49</td>
<td>531 ± 44</td>
</tr>
<tr>
<td>Reaction Time Analyses VSRT</td>
<td>772 ± 61</td>
<td>796 ± 65</td>
<td>714 ± 58</td>
<td>758 ± 43</td>
<td>708 ± 51</td>
<td>693 ± 47</td>
</tr>
<tr>
<td>Unfiltered Light</td>
<td>865 ± 71</td>
<td>841 ± 54</td>
<td>789 ± 74</td>
<td>767 ± 67</td>
<td>769 ± 72</td>
<td>708 ± 56</td>
</tr>
<tr>
<td>Vigilance NUMC</td>
<td>48.6 ± 3.6</td>
<td>35.5 ± 3.7*</td>
<td>45.3 ± 4.4</td>
<td>38.2 ± 4.3*</td>
<td>44.3 ± 3.6</td>
<td>40.9 ± 4.7</td>
</tr>
<tr>
<td>Unfiltered Light</td>
<td>47.6 ± 3.3</td>
<td>34.3 ± 4.4*</td>
<td>41.5 ± 2.5</td>
<td>37.7 ± 3.7*</td>
<td>40.0 ± 4.1</td>
<td>40.6 ± 3.8</td>
</tr>
<tr>
<td>Vigilance MRTC</td>
<td>0.73 ± 0.04</td>
<td>0.88 ± 0.07*</td>
<td>0.83 ± 0.07</td>
<td>0.85 ± 0.06</td>
<td>0.82 ± 0.07</td>
<td>0.82 ± 0.07</td>
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<tr>
<td>Unfiltered Light</td>
<td>0.77 ± 0.04</td>
<td>0.89 ± 0.06*</td>
<td>0.82 ± 0.07</td>
<td>0.83 ± 0.06</td>
<td>0.83 ± 0.07</td>
<td>0.83 ± 0.06</td>
</tr>
<tr>
<td>Vigilance NUMI</td>
<td>16.5 ± 6.6</td>
<td>11.3 ± 3.1</td>
<td>8.9 ± 2.0</td>
<td>9.7 ± 1.9</td>
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<td>9.0 ± 2.5</td>
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<tr>
<td>Unfiltered Light</td>
<td>9.9 ± 3.8</td>
<td>13.0 ± 3.2</td>
<td>7.4 ± 2.8</td>
<td>12.4 ± 3.4*</td>
<td>5.6 ± 1.5</td>
<td>9.0 ± 2.0</td>
</tr>
</tbody>
</table>

A (1,2): the first or second ability test during the experimental night shift; specific time points for the start of the respective tests were: Flicker/Fusion Frequency A1 = 10:15 p.m., Flicker/Fusion Frequency A2 = 5:30 a.m.; Continuous Attention A1 = 11:00 p.m., Continuous Attention A2 = 2:15 a.m.; Reaction Time Analysis A1 = 11:35 p.m., Reaction Time Analysis A2 = 3:00 a.m.; Vigilance A1 = midnight, Vigilance A2: 3:30 a.m.

*P<0.05 as compared to the first measurement (A1) on the respective night of the study protocol (Wilcoxon matched pairs test).

Abbreviations: CRT = choice reaction time; IP = information processing; MEANC = mean time for correct hits; MEANI = mean time for incorrect hits; MRO = motor response organization; MRTC = mean value of reaction time for correct hits; NUMC = number of correct hits; NUMI = number of incorrect hits; PER = perception; SUMC = sum of correct hits; SUMI = sum of incorrect hits; VSRT = reaction time for visual search.

applied dexamethasone, prednisolone, or hydrocortisone induced apoptosis in eosinophils and lymphocytes, while programmed cell death in neutrophils was averted [24,25]. Since we did not measure serum cortisol levels, we can only speculate that a nocturnal decrease in glucocorticosteroid synthesis and release is responsible for the alterations in white blood count. Interestingly, variations in white blood count have recently been documented by Nishitani and Sakakibara [26] to be more pronounced in shift workers as compared to daytime workers. The authors suggested that quality of sleep might affect white blood status. It should be noted that the changes did not differ between the room with...
filtered light and the room with full-spectrum light, thus being independent of the chosen illumination.

Blood pressure follows a distinct circadian profile with a characteristic decline during sleep (nocturnal values being 10-20% lower than the mean daytime measurements) followed by a surge in the early morning hours [27]. Since circadian rhythm in blood pressure is largely dependent on regular sleep-wake patterns, night shift is usually associated with rapid alterations in the course of 24 h systolic and diastolic blood pressure behaviour [28]. In our study, systolic and diastolic blood pressure remained within a normal circadian design in the room with filtered light at night 1 and 3 of the study, while there were no significant overall time effects in the room with full-spectrum light, implying that the circadian profile was deranged. However, the preserving effect of light with reduced short wavelength components on nocturnal variations in blood pressure values was not consistent throughout the experiments since it was not present on the second night. Circadian oscillations of the autonomic nervous system with a prevalence of sympathetic tone at daytime and a switch towards parasympathetic predominance at night results in a nocturnal decrease in heart rate [29,30]. Although the effects of an irregular sleep-wake cycle on heart rate associated with shift work are discussed controversially [31,32], light with intensities sufficient to affect circadian patterns of melatonin has been shown to influence heart rate in humans [33]. Friedman analyses of our data yielded a significant change in nocturnal heart rate with decreases in the course of all nights in filtered light and at both nights 2 and 3 in full-spectrum light. These data do not support the concept of a differentiating effect of the chosen lighting environments on heart rate values.

Maintenance of a physiological circadian rhythm represents only one ergonomical aspect for the design of workplace illuminations. Another important topic is the employees’ efficiency and well-being. In this regard, pulses of bright light have been reported to exert beneficial effects on alertness as well as performance in night shift workers [34,35]. In a study by Eastman and co-workers [36], high intensities of bright light (5000 lux) modulated phase shifting of temperature which was associated with better daytime recovery, fatigue, and overall mood as compared to 500 lux dim light controls in a simulated night shift model. These data demonstrate a favourable effect of intermittent nocturnal exposure to bright light. Therefore, we had to take into account a potentially detrimental effect of light with reduced short wavelength components on productivity and performance of our study participants. In order to assess this question, various ability tests were performed throughout the course of the study. Flicker/Fusion Frequency analysis is regarded as an indicator for central-nervous activation. In our study, flicker frequency was significantly decreased at the end of each night shift, which can be interpreted as a reduction in volunteer’s arousal. However, these changes were observed under both lighting conditions and statistical comparison of ΔA2A1 values did not result in a significantly different impact of workplace illumination on this parameter. Continuous Attention testing is designed to evaluate long-term attention and concentration ability, and thereby allowing an indirect assessment of general performance. Although some of the parameters included in the respective analyses were significantly altered when comparing the second with the first time point of data collection, we could not detect an unambiguous deleterious effect of filtered light on the level of attentiveness of our study participants. As with Flicker/Fusion Frequency, ΔA2A1 values could not discriminate between filtered and full-spectrum light. According to the different stages of activity regulation, we could not observe significant alterations in information processing, perception, and motor response organization measured by Reaction Time Analyses on the three nights of the study. These results do not support the hypothesis that filtered light deteriorates cognitive speed in our experimental night shift model. However, there was a significant drop in the number of correct hits assessed during Vigilance testing at both night 1 and 2 under filtered light that was present only at night 1 under full-spectrum light. Although ΔA2A1 values for this parameter did not differ between the two office accommodations at any night of the study, our findings indicate a possible impairment of the performance of monotonous monitoring tasks in our test light system with reduced short wavelength components. Apart from the Vigilance test data, filtered light did not affect alertness and performance in our experimental setup. Thus, our data do not support the concept of a detrimental effect of maintaining circadian melatonin rhythm on fatigue, precision, or on-the-job attentiveness. This view is confirmed by Paul et al. [37], who did not find impaired psychomotor performance (attentiveness, reaction time, motor response organization) in volunteers treated with exogenously applied melatonin. Moreover, in the above-mentioned study by Kayumov and co-workers [21], preservation of nocturnal melatonin levels did not exert an influence on performance data or subjective sleepiness as well.

CONCLUSION

The aim of this study was to investigate the effects of a lighting environment with reduced short-wavelength components on physiological parameters following a circadian pattern as well as on parameters of performance and efficiency in a simulated night shift setup. The major finding of our study is the maintenance of the nocturnal rhythm of melatonin under test light conditions as compared to unfiltered bright light which was not accompanied by losses in the volunteer’s alertness or general performance. With respect to the potentially harmful effects of disturbing a regular circadian rhythm, installation of workplaces with adapted illumination may provide a benefit for employees working night shifts.

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REFERENCES