Elicitation of Trigonelline and 4-Hydroxyisoleucine with Hypoglycemic Activity in Cell Suspension Cultures of *Trigonella foenum graecum* L.

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**Abstract:** Cell suspension cultures of *Trigonella foenum graecum* L. (fenugreek) were initiated from the cotyledon portions of sterile germinated seeds and maintained on MS medium supplemented with 2, 4-dichlorophenoxy acetic acid (2, 4-D) (1 mg/l), kinetin (0.1 mg/l) and sucrose (5%). The changes in cell mass and both trigonelline and 4-hydroxyisoleucine content were followed between days zero and 12. The linear increase in fresh weight was found to be parallel to both trigonelline and 4-hydroxyisoleucine production. Cell suspension cultures treated with 100 μM methyl jasmonate (MJ) for 24 hours showed a noticeable increase in the level of trigonelline and 4-hydroxyisoleucine. The marked improvement in the histological and electron microscopically pictures of pancreas of STZ-diabetic rats fed with extract of cells treated with MJ is coincided with more effective and significant hypoglycemic activity than that for seeds extract. The extract of cultured cells treated with MJ lowered blood glucose from 284 ± 7.4 to 123 ± 8.1 units and increased the insulin level from 4.42 ± 0.23 μU/ml to a high level 8.33 ± 0.41 μU/ml.

**Keywords:** *Trigonella foenum graecum*, MS medium, trigonelline, 4-hydroxyisoleucine, methyl jasmonate, HPLC.

**INTRODUCTION**

Fenugreek (*Trigonella foenum-graecum* L. *Leguminosae*) is one of the oldest medicinal plants. This plant is thought to originate from India or the Middle East. It is now primarily grown in India and in the Mediterranean countries.

It is an annual herbaceous plant belonging to the Fabaceae family, 40-60 cm high, with alternate trifoliate leaves and pale yellow flowers. The leaves and seeds, which mature in long pods, are used to prepare extracts or powders for medicinal use.

In ancient Egypt, fenugreek was used in incense and to embalm mummies. In modern Egypt, fenugreek is still used as a supplement in wheat and maize flour for bread-making [1]. In ancient Rome, fenugreek was purportedly used to aid labor and delivery. In traditional Chinese medicine, fenugreek seeds are used as a tonic, as well as a treatment for weakness and edema of the legs [2]. In India, fenugreek is commonly consumed as a condiment [2] and used medicinally as a lactation stimulant [3]. It is widely considered as an antidiabetic and anticholesterol herb. Various clinical trials of fenugreek showed improvement in glucose tolerance in healthy volunteers as well as in type 2 and type 1 diabetic patients [4-7].

Trigonelline is known to have some hypoglycemic effect. The effect of an alkaloid extract of fenugreek dried seeds (*Trigonella foenum graecum* L.) on blood glucose, serum insulin, serum lipid profile and lipid peroxidation in addition to histological and histochemical study of liver and kidney in streptozotocin induced diabetic albino rats have been studied [8]. Further investigations concerning the protective effect of *Trigonella foenum graecum* L. on the histological structures and function of liver, kidney and pancreas of induced diabetes were studied [9-12].

However, most attention has been given to 4-hydroxyisoleucine. From the seeds of *T. foenum-graecum* an unusual amino acid, 4-hydroxyisoleucine, has been isolated, which significantly decreased the plasma triglyceride levels by 33% (P<0.002), total cholesterol (TC) by 22% (P<0.02), and free fatty acids by 14%, accompanied by an increase in HDL-C/TC ratio by 39% in the dyslipidemic hamster model [13]. In *vitro*, it was shown to increase glucose-induced insulin release in human and rat pancreatic islet cells [14, 15].

Plant tissue culture technique is being used for producing primary and secondary metabolites as early as 1950’s. Since then this technique has been developed for the production of various secondary metabolites [16-19]. As a part of our continuing interest concerning the production of biologically active compounds by tissue culture [20-22], the present work describes optimum conditions to induce accumulation of hypoglycemic trigonelline and 4-hydroxyisoleucine in cell suspension cultures of *Trigonella foenum graecum* L.

**MATERIALS AND METHODS**

**General Procedures**

TLC was performed on silica gel 60 F254-coated aluminum sheets (Merck, Darmstadt, Germany). Plates were visualized by spraying with Dragendorff reagent.
Elicitation of Trigonelline and 4-Hydroxyisoleucine with Hypoglycemic

Effects of Methyl Jasmonate

The quantities of Trigonelline and 4-hydroxyisoleucine in both the Trigonella foenum-graecum seeds and cultured cells (control & treated) were determined by HPLC. 30 μl was injected into a reversed phase (hydroxyisoleucine (A) and acetonitrile (B) were used as solvents. The employed gradient was: 10-30% (B) in 30 min. The flow rate was 1 ml/min and the detection wavelength set to 265 nm. Trigonelline and 4-hydroxyisoleucine in a concentration of 1 μg/100 μl were used separately as reference compounds. The quantities of both were determined by HPLC. Moreover, HPLC analysis was performed as previously published [26]. The STZ was dissolved in 0.1 ml of citrate buffer (pH 4.5). A single dose of STZ (60 mg/kg) was injected intraperitoneally in animals for induction of diabetes.

Animals and Induction of Diabetes

Male Wister rats weighing 180–200 g were obtained from the experimental animal care centre, King Abdelaziz University, Jeddah, Saudi Arabia. Induction of diabetes was performed as previously published [26]. The STZ was dissolved in 0.1 ml of citrate buffer (pH 4.5). A single dose of STZ (60 mg/kg) was injected intraperitoneally in animals for induction of diabetes.

HPLC Analysis

The quantities of Trigonelline and 4-hydroxyisoleucine in both the Trigonella foenum-graecum seeds and cultured cells (control & treated) were determined by HPLC. 30 μl was injected into a reversed phase (hydroxyisoleucine (A) and acetonitrile (B) were used as solvents. The employed gradient was: 10-30% (B) in 30 min. The flow rate was 1 ml/min and the detection wavelength set to 265 nm. Trigonelline and 4-hydroxyisoleucine in a concentration of 1 μg/100 μl were used separately as reference compounds. The quantities of both were determined by HPLC. Moreover, HPLC analysis was performed as previously published [26]. The STZ was dissolved in 0.1 ml of citrate buffer (pH 4.5). A single dose of STZ (60 mg/kg) was injected intraperitoneally in animals for induction of diabetes.
STZ-treated rats were given 5% glucose in their drinking water for the first 24 h to counter any initial hypoglycemia. Normal animals were similarly injected with vehicle only. On the third day, the animals were checked for the presence of glucose in the urine using enzymatic test strips. The animals were maintained under standard conditions of temperature 25 ± 5 °C and 55 ± 5% relative humidity with a regular 12 h light: 12 h dark cycle and allowed free access to standard laboratory food (Purina Chow) and water. All animals were treated humanely in accordance with the guideline for care of animals as set by WHO.

Biological Study

As mentioned previously in literature [26]; the rats were randomly divided into four groups of 10 animals each. The first group was received citrate buffer alone (normal), the second was STZ-diabetic rats (control), the third was the STZ-diabetic rats fed orally with 50 mg/kg/day seeds extract of *Trigonella foenum graecum*, and while the fourth was the STZ-diabetic rats fed orally with 50 mg/kg/day extract of cultured cells treated with MJ for 10 successive days.

On the eleventh day, the rats were subjected to light ether anesthesia and killed by cervical dislocation. Trunk blood was collected into heparinised chilled tubes containing sodium fluoride (to inhibit glycolysis). Serum was separated by centrifugation at 4 °C for 15 min at 3000 rpm and stored at -20 °C until determination of serum glucose and serum insulin concentrations.

Haematoxylin and Eosin (Hx&E) Staining

The staining was done to demonstrate the morphologic appearance and the integrity of the specimens. Equal samples from the pancreatic tails (5mm x 5mm) were taken and processed immediately. Paraffin sections were cut at 6um, prepared and stained with Ehrlich hematoxyline and eosin [27].

Ultrastructural Technique

Adjacent sections of pancreas were removed and placed in 0.1M cacodylate buffer containing 3% glutaraldehyde for electron microscopy. After fixation, sections were dehydrated in a series of ethanol rinses, cleared with propylene oxide, and embedded in epon. Ultrathin sections (70-80 nm) were cut using a diamond knife and double stained with uranyl acetate and lead citrate and examined under a transmission electron microscope [27].

Biochemical Analysis

Serum blood glucose levels were estimated by glucose oxidase method [28]. Serum insulin levels were determined by GenWay INS-EASIA [29].

Statistical Analysis

Data are expressed as means ± S.E.M. Statistical comparison between different groups were done using one-way analysis of variance (ANOVA) followed by the Tukey–Kramer multiple comparison test, to judge the difference between various groups. Significance was accepted at $P < 0.05$.

RESULTS AND DISCUSSION

Trigonelline and 4-Hydroxyisoleucine Accumulation and Growth Curve

A linear increase in fresh weight was observed between day 2 and day 8 after inoculation of cells into fresh MS medium. HPLC analysis of trigonelline and 4-hydroxyisoleucine of untreated cell suspension cultures of *Trigonella foenum graecum* marked a linear increase in their content between day 4 and day 8 paralleled cell growth (Figs. 2 and 3).

Effect of MJ on Trigonelline and 4-Hydroxyisoleucine Content

The composition of fenugreek seeds was extensively studied and their composition reveals a few specific features [14, 15, 30] (Table 1).

MJ is a substance used in plant defence and many diverse developmental pathways such as seed germination, root growth, flowering, fruit ripening and senescence. In addition,
MJ is involved in signal transduction and induces the transcription of biosynthetic enzymes involved in the formation of defence compounds in plants [31, 32]. Since MJ can induce the formation of secondary metabolites in other systems [21, 32-34], we have tried to stimulate the accumulation of trigonelline and 4-hydroxyisoleucine in cell suspension cultures of *Trigonella foenum-graecum* by adding MJ.

The cell suspension cultures of *Trigonella foenum-graecum* (4 days old) were treated with 100 μM methyl jasmonate (MJ) and incubated for 24 h.

The trigonelline and 4-hydroxyisoleucine were extracted and determined quantitatively by HPLC and their estimation was done by peak area calculations. MJ at a concentration of 100 μM induced the accumulation of trigonelline about 2-fold and 4-hydroxyisoleucine about 3-fold as compared to the seeds extract (Table 1).

**Antidiabetic Activity**

STZ-diabetic rats fed with seeds extract showed lowered blood glucose from 284 ± 7.4 to 144 ± 7.81 and an increase in the insulin level from 4.42 ± 0.23 μU/ml to a high level 6.9 ± 0.47 μU/ml. STZ-diabetic rats fed with extract of cells treated with MJ showed lowered blood glucose from 284 ± 7.4 to 123 ± 8.1 and an increase in the insulin level from 4.42 ± 0.23 μU/ml to a high level 8.33 ± 0.41 μU/ml (Table 2).

The considerable increase in level of both trigonelline and 4-hydroxyisoleucine in the cultured cells treated with MJ as mentioned above (Table 1) compared to the level in the seeds extract is coincided with more effective and significant hypoglycemic activity of extract of the cells treated with MJ. Furthermore, the results of this study agree with published data [8, 13, 14] which reported the hypoglycemic effect of both trigonelline and 4-hydroxyisoleucine. Trigonelline is known to have some hypoglycemic effect. However, most attention was given to 4-hydroxyisoleucine [15]. 4-Hydroxyisoleucine has been shown to increase glucose-induced insulin release without interacting with other agonists of insulin secretion such as tolbutamide and glyceraldehydes, thus demonstrating a novel *in vitro* insulinotropic activity [14].

**Histological Picture**

All pancreatic cells of the normal rats were present in their normal proportions. The acinar cells which stained strongly are arranged in lobules with prominent nuclei.

**Table 1.** Trigonelline and 4-Hydroxyisoleucine Concentrations in Seeds and Cell Suspension Cultures Extracts of *Trigonella foenum-graecum*

<table>
<thead>
<tr>
<th>Extract</th>
<th>% of trigonelline and 4-hydroxyisoleucine / seed weight</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>Trigonelline</td>
</tr>
<tr>
<td></td>
<td>Found*, ± SD</td>
</tr>
<tr>
<td>Seeds</td>
<td>0.13 ± 0.03</td>
</tr>
<tr>
<td>Control (untreated cells)</td>
<td>0.16 ± 0.02</td>
</tr>
<tr>
<td>Cells treated with 100 μM MJ</td>
<td>0.27 ± 0.04</td>
</tr>
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*The data are mean values of two independent experiments.*
The islet cells are seen embedded within the acinar cells and surrounded by a fine capsule (Fig. 4).

Pancreatic cells of the STZ-diabetic rats (control) showing a progressive distortion in the histoarchitecture of the pancreatic parenchyma and stroma compared to normal group.

A marked increase in the adipose tissue was found at the expense of the functional pancreatic mass, many fragmented islets and areas of progressive interstitial fibrosis (Fig. 5) with a heavy lymphocytic infiltration in and around it with an ill-defined capsule (Fig. 6).

The pancreas of STZ-diabetic rats fed with seeds extract showed the acinar cells to be normal. The islets were mildly increased in number with scanty lymphocytic infiltration within the well defined capsule (Fig. 7).

The pancreas of STZ-diabetic rats fed with extract of cells treated with MJ showed normal appearance of acinar cells. The islets are present with a large proportion of islet cells as compared with control. There are few inflammatory cells within the well defined capsule and an increase in the size of blood vessels (Fig. 8).

**Ultrastructural Picture**

Transmission electron photomicrograph of pancreas of normal rats showed alpha cells with normal sized cytoplasmic secretory granules. The nuclei are regularly outlined and have two nucleoli (Fig. 9).

Transmission electron photomicrograph of pancreas of STZ-diabetic rats (control) showed the damaged beta cell contains a large vacuole (V) & amorphous material (M) filling a space formerly occupied by beta granules and the

<table>
<thead>
<tr>
<th>Groups</th>
<th>Fasting blood glucose (mg/dl)</th>
<th>Serum insulin (μU/ml)</th>
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<tbody>
<tr>
<td>Normal rats</td>
<td>77 ± 3.1</td>
<td>10.7 ± 0.53</td>
</tr>
<tr>
<td>STZ-diabetic rats (control)</td>
<td>284 ± 7.4</td>
<td>4.42 ± 0.23</td>
</tr>
<tr>
<td>STZ-diabetic rats fed with seeds extract</td>
<td>144 ± 7.81</td>
<td>6.9 ± 0.47</td>
</tr>
<tr>
<td>STZ-diabetic rats fed with extract of cells treated with MJ</td>
<td>123 ± 8.11</td>
<td>8.33 ± 0.51</td>
</tr>
</tbody>
</table>

*Data are expressed as means ± S.E.M. *P < 0.05 between normal and diabetic control and between diabetic control and diabetic fed on treated cells extract.
nucleus (N) is fairly regular in outline with two well defined nucleoli (Fig. 10).

Transmission electron photomicrograph of pancreas of STZ-diabetic rats fed with seeds extract showed nearly normal beta cells is less number of vacuoles with swollen mitochondria, the cytoplasmic granules having an electron dense core surrounded by a clear zone and the nucleus is fairly regular in outline with two well defined nucleoli (Fig. 11).

CONCLUSION

It is important to conclude the role of plant tissue culture used in this experiment as an alternative source for large scale production of bioactive compounds. Methyl jasmonate
MJ used in this experiment affected positively the accumulation of both trigonelline and 4-hydroxyisoleucine in cell suspension cultures of \textit{Trigonella foenum graecum} L. The noticeable improvement in the histology of pancreas of STZ-diabetic rats fed with extract of cells treated with MJ is coincided with significant hypoglycemic activity than that for seeds extract.

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