

Design, Synthesis and Evaluation of 1,3,2-Diazaphosphorin[4,5-b]Quinoxaline-5,10-di-N-oxide Derivatives as Novel VEGFR-2 and SRC Kinase Inhibitors in the Treatment of Prostate Cancer

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Abstract: Prostate cancer remains a significant public health problem, with limited therapeutic options. Recently, new approaches in prostate cancer treatment have been developed, inhibition of the proangiogenic VEGFR kinase and SRC kinase, involved in progression and metastasis of prostate cancer were investigated. A new series of 1,3,2-diazaphosphorin[4,5-b]quinoxaline-5,10-di-N-oxide was synthesized using Lawson's reagent in a reaction that provides a rare example of the incorporation of phosphorous. The structure of the new compounds was confirmed by spectroscopic data. The new synthesized compounds were evaluated for their *in-vitro* prostate cancer cell line (PC3). Compounds **7a** and **8a** possessed remarkable antitumor activity more active than the known drug Doxorubicin with IC₅₀= 0.12, 0.36 and 0.63 μ M respectively. Evaluation of the new synthesized compounds on VEGFR-2 and SRC kinases showed that compound **7a** was the most effective with IC₅₀= 18 μ M. On the other hand, weak inhibitory effect was observed by the compounds on SRC kinase. Docking study was performed for compound **7a** into ATP binding site of VEGFR-2 kinase.

Keywords: Diazaphosphorin, quinoxaline 1,4-di-N-oxides, prostate cancer, VEGFR, SRC kinase, molecular modeling.

1. INTRODUCTION

Prostate cancer remains a significant public health problem, with limited therapeutic options in the setting of castrate-resistant metastatic disease [1, 2]. Until recently, there were only four FDA-approved chemotherapeutic agents (estramustine [3], itoxantrone [4, 5], docetaxel [3, 5], cabazitaxel [6-9] with docetaxel and prednisone being a standard for patients requiring chemotherapy. However, the responses are not durable and the disease ultimately progresses [10]. A direct strategy depending on targeting tumor cells with chemotherapeutic agents has dominated the field of cancer therapy for several years [11] however, this strategy was also toxic to the non-malignant cells and would eventually develop resistance [12]. Recently, there was an improvement in cancer therapies aimed at the underlying cancer pathology [13]. One of these indirect anti-cancer therapies was based on the fact that tumor growth beyond a few millimeters and metastatic dissemination within the host depended on the induction of angiogenesis, or the process of new blood vessel formation [14-18]. One of the most important pro-angiogenic factors involved in tumor angiogenesis is vascular endothelial growth factor (VEGF) [15, 19]. Hence, blockade of the VEGF/VEGFR-signaling is regarded as an attractive therapeutic target for inhibition of tumor angiogenesis [13, 20]. In addition, several reports

have implicated VEGF in prostate carcinogenesis and in its metastatic spread [21-26]. Clinical trials, phase II and III, investigating VEGF-targeting therapy in combination with chemotherapeutic agents in prostate cancer treatment concluded that this combination is reasonably safe and effective [26-28]. Moreover, quinoxaline 1,4-di-N-oxides derivatives proved to be potential cytotoxic agents against prostate cancer [29-31], two novel quinoxaline 1,4-di-N-oxides derivatives Q39 [32] and TX-2098 [33], Fig. (1), were reported to exert their activity possibly through VEGF inhibition. Furthermore, the lead compound of this series of compounds, tirapazamine, has been successfully used in combination with low-dose metronomic cyclophosphamide in treatment of human prostate cancer PC-3 [34]. On the other hand, Src is an example of non-receptor TKs [35] involved in multiple signaling pathways central to prostate cancer development, progression and metastasis in addition to its osteoclast activities [36, 37] leading to increased VEGF expression [38]. As such, Src inhibition represents a valid therapeutic strategy for the treatment of advanced metastatic prostate cancer. In light of this data, it seemed clear that there is a great need to exploit novel targets and new agents in the search for more effective treatment [39-41]. The reported inhibitory effect of quinoxaline 1,4-di-N-oxide on VEGF expression [32, 33] combined with the cytotoxic activity of compounds derived from the 1,3,2-diazaphosphorine [42], encouraged us to synthesize and evaluate the inhibitory effect of a new series of 1,3,2-diazaphosphorine[4,5-b]quinoxaline-5,10-di-N oxides against VEGFR-2 and Src kinases.

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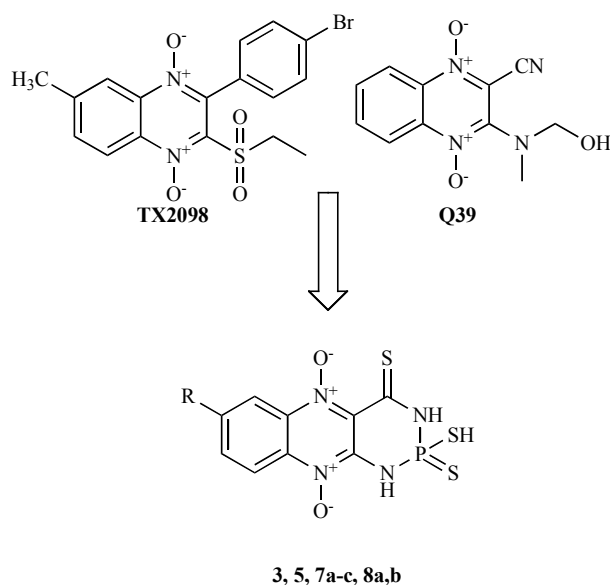


Fig. (1). New design of the synthesized compounds.

2. RESULTS AND DISCUSSION

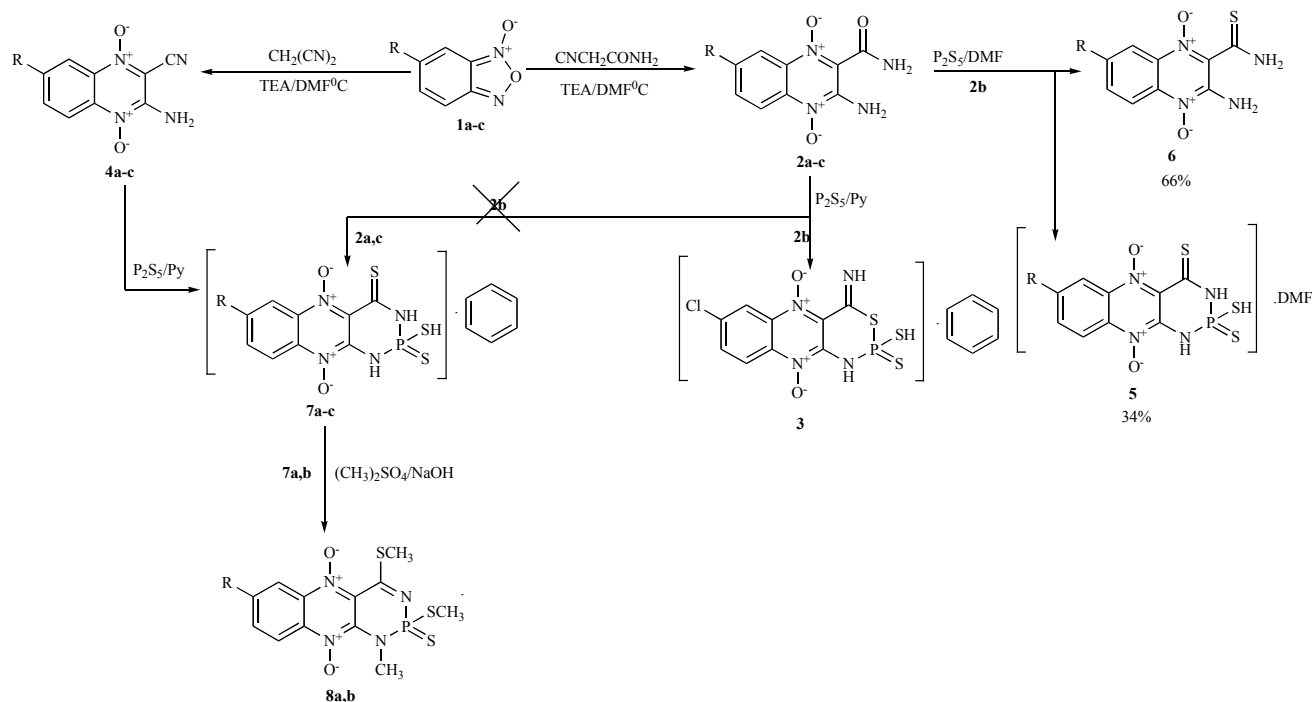
2.1. Chemistry

Attempts to convert carbamoyl or carbonitrile groups using phosphorous pentasulphide in pyridine by previous research groups, proved that the reaction did not stop at the thionation step but rather proceeded all the way to the formation of annelated heterocycles and 1,3,2-diazaphosphorin rings [43-47]. This reaction provides a rare example of the incorporation of phosphorous derived from P_2S_5 , into a heterocyclic system. Scheme 1 outlines a new synthetic pathway to prepare the tricyclic diazaphosphorin compounds **3**, **5**, **7a-c** and **8a, b** two key substrates were prepared, 2-amino-3-carbamoyl-6-substituted quinoxaline-1,4-dioxide **2a-c**

and 3-amino-7-substituted-2-quinoxalinecarbonitrile-1,4 di-N-oxides **4a-c** [48].

They were obtained by the well-known Beirut [49, 50] reaction between 5-substitutedbenzo [c] [1, 2, 5]oxadiazol-1-oxides (benzofuroxanes,**1a,b**) and cyanoacetamide for derivatives **2a-c** or malononitrile for **4a-c**. The reaction of either 3-carbamoyl derivatives **2a,c** or 2-carbonitrile derivatives **4a-c** with P_2S_5 in pyridine yielded 2-mercapto-7-substituted [1, 3, 2] diazaphosphorin[4, 5-b]quinoxaline-5,10-di-N-oxide-2,4(1H,3H)-dithione in the form of pyridine solvates **7a-c**, Scheme 1. While, compound **2b**, under the same reaction conditions, gave the azathiaphosphorine derivative **3**. The synthesized compounds were assigned their structures based on the spectral and microanalytical data. Infrared spectrum of the products **3**, **7a-c** (isolated as pyridine solvates) was characterized by the disappearance of bands corresponding to cyano and carboxamide groups in addition to the bands at cm^{-1} 3364-3231(2 NH); 1544, 1484 (NO); 1228(C=S). In the 1H NMR spectra, the appearance of new signals corresponding to pyridine protons characterized the spectra in addition to the typical pattern of proton signals of quinoxaline-1,4-di-N-oxide, a broad NH signal was also observed. An SH proton signal is not clearly seen in the 1H NMR spectra, which is consistent with the data concerning a study of the spectra for such compounds [44]. When this reaction was carried out in DMF, two products were obtained, the diazaphosphorine **5** and in a larger portion 2-amino-3-carbamothioyl-6-chloroquinoxaline 1,4-dioxide **6**, proved by spectral and microanalytical data.

Furthermore, the removal of the pyridine in ether and methylation of the aqueous solution with dimethyl sulphate gave the trimethyl derivatives **7a,b** compound **7c** was excluded from this reaction due to its low yield. The 1H NMR spectra of these compounds displayed singlets in the range 2.07-3.28 ppm corresponding to the methyl groups.



Scheme (1). Synthetic route for the preparation of the newly synthesized compounds.

Table 2. % Activity Change of Src and VEGFR-2 Kinases Tested Against the Target Compounds

Compound	Src Kinase			VEGFR Kinase		
	% Activity Change 5 μ M	% Activity Change 20 μ M	% Activity Change 50 μ M	% Activity Change 5 μ M	% Activity Change 20 μ M	% Activity Change 50 μ M
3	2	11	-27	-5	-23	-43
5	-8	-5	-21	-6	-26	-58
7a	-7	-20	-17	-23	-54	-71
7b	-4	-8	-18	0	9	-17
7c	6	3	-10	-8	-22	-41
8a	5	2	-19	-1	-21	-35
8b	-6	-25	-31	-1	-14	-12

Table 3. IC₅₀ Values of the Tested Compounds Against VEGFR-2 and Src Kinases

Compound	IC ₅₀ VEGFR-2 Kinase μ M	IC ₅₀ Src Kinase μ M
3	64.11	ND
5	40.32	209.4
7a	17.8	3133
7b	ND ^a	303.9
7c	74.69	ND
8a	86.25	ND
8b	1587	142.8

^aND denotes the compounds where the IC₅₀ could not be determined.

assay format for profiling evaluation of protein kinase targets. The results observed as percentage activity change compared to control are presented in Table 2. The intra-assay variability was determined to be less than 10%. Inhibition of target activity by the compound gives negative values, while activation of target activity gives positive values. It has been considered that only values of >15% change in activity compared to control to be significant. The profiling data for the compounds against Src and VEGFR-2 target is presented in Table 2. The profiling data for the compounds against VEGFR-2 target ranged from good to weak inhibition of its activity, Table 2. The 2-mercapto-[1, 3, 2] diazaphosphorin [4, 5-b] quinoxaline 5,10-di-N-oxide -2,4(1H,3H)-dithione **7a** showed the best inhibitory effect out of all the compounds where VEGFR-2 activity was inhibited by 71% at 50 μ M concentration, compared to control. Compound **5** also showed moderate inhibition of VEGFR-2 activity by 58% at 50 μ M concentration, compared to control. The other compounds showed weak inhibition of VEGFR activity. Moreover, all the tested compounds displayed weak inhibition of Src activity.

Furthermore, IC₅₀ determinations were done for the tested compounds being assayed by Src and VEGFR-2 Kinases, the IC₅₀ concentrations against Src were quite high, ranging from 143 μ M to more than 3130 μ M, Table 3. On the other hand, compounds being assayed by VEGFR-

2 Kinase displayed IC₅₀ values much lower than those with the Src kinase, ranging from 18 μ M to more than 1580 μ M. The best result observed was for the diazaphosphorin derivative **7a**, Table 3. However, activity of this compound was significantly reduced upon substitution of the hydrogens of thiol and amino groups with methyl groups. Contrary to this observation the methylated derivative **8b** showed an inhibitory effect on Src kinase greater than its analogue **8a**.

2.3. Docking Studies

The vascular endothelial growth factor and its receptor tyrosine kinase VEGFR-2 or kinase insert domain receptor (KDR) are attractive targets for the development of novel anticancer agents [53]. The goal for researchers would be to find a selective inhibitor for the VEGFR-2 (KDR), because it is expressed on almost all endothelial cells and the majority of the effects in angiogenesis, including cell proliferation, micro-vascular permeability, invasion, migration, and survival are mediated by VEGFR-2 [54, 55]. In the present work, docking studies were performed to explore the binding mode between the 1,3,2-diazaphosphorine derivatives and KDR. The process was started by retrieving crystal structure of VEGFR-2 (PDB ID: 1VR2) from Protein Data Bank [56]. Docking studies of compounds with combination of VEGFR-2 (PDB ID: 1VR2) was performed using Molecular Operating Environment (MOE.10.2012) [57]. After

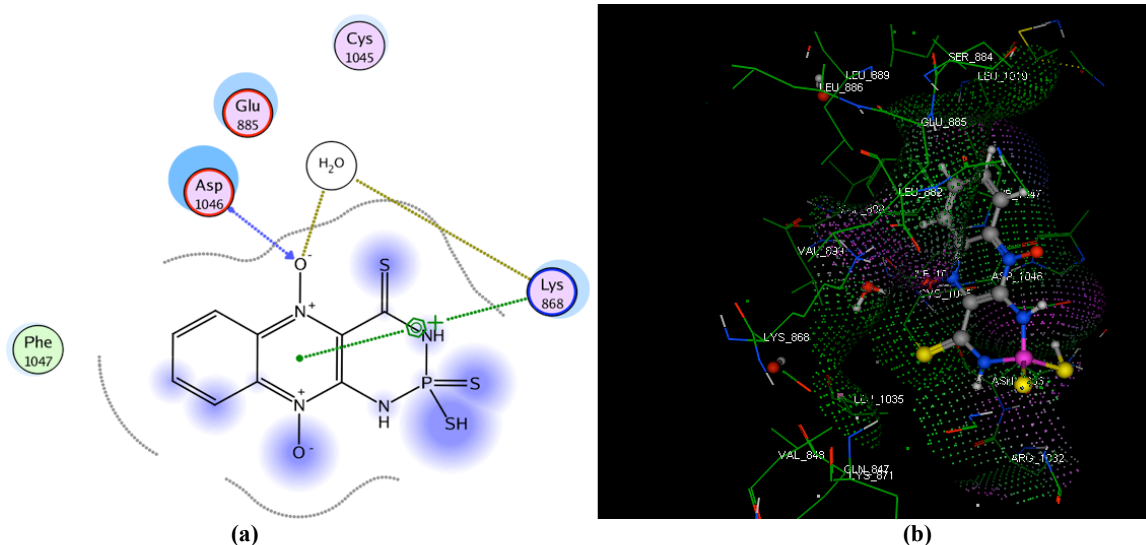


Fig. (3) (a) Two-dimensional representation of the interacting mode of **7a** with VEGFR-2 kinase. (b) Three-dimensional structural models of compound **7a** into VEGFR-2 kinase derived from the docking simulations.

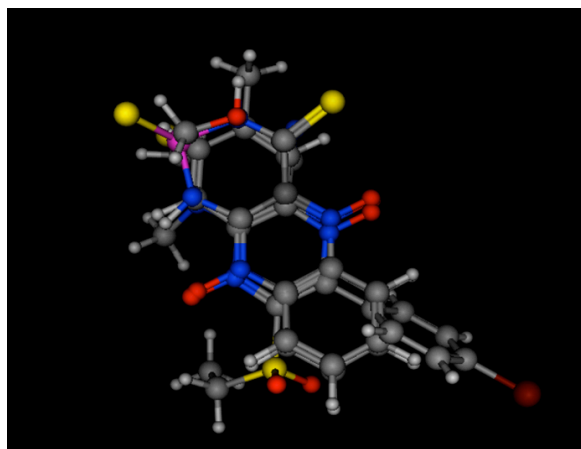


Fig. (4). Comparison of conformational alignments of compound **7a** with Q39 and TX-2098.

inspecting the kinase active site of several VEGFR-2 structures, 4 residues were selected: Lys868, Glu885, Cys919 and Asp1046, these residues interact consistently, by forming hydrogen bonds (H-bonds) with the inhibitors that are co-crystallized in the inspected VEGFR-2 structures [58, 59]. Thus, the compounds were docked in their most stable conformation into the ATP-binding site of VEGFR-2 and subjected to energy minimization. The best-fitted conformer (docking score -3.75 Kcal/mol) was observed for 2-mercapto- [1, 3, 2] diazaphosphorin [4, 5-b] quinoxaline-5, 10-di-N-oxide-2, 4 (1H,3H)-dithione **7a**. The docking interaction of compound **7a** with the active site of VEGFR-2 is represented in Fig. (3). When **7a** was bound to VEGFR-2, it formed hydrogen bonds with backbone-NH of Asp1046 (2.87 Å) and Lys 868 (2.30 Å), through a water molecule, as well as a strong ionic contact with Lys 868. The diazaphosphorin seems to be extended toward the back hydrophobic pocket created by the conformational rearrangement of Phe1047 and Cys 1045 [60].

Moreover, comparison of conformational alignments of compound **7a** and the quinoxalinederivatives Q39 and TX-2098 were also performed, Fig. (4).

3. EXPERIMENTAL

3.1. Chemistry

3.1.1. Chemical Methods

All the solvents used were commercially available and distilled before use. Reactions were monitored by thin-layer chromatography (TLC) on silica gel plates (60 F254), visualizing with ultraviolet light. Infra-red spectra (KBr) were recorded on FT-IR 5300 spectrophotometer and Perkin Elmer spectrum RXIFT-IR system (ν , cm^{-1}). ^1H NMR spectra were recorded on Varian Gemini spectrophotometer (300 MHz) in DMSO-d_6 as solvent. Proton chemical shifts (δ) are relative to tetramethylsilane (TMS, $\delta = 0.00$) as internal standard and expressed in ppm. Spin multiplicities are given as s (singlet), d (doublet), t (triplet), and m (multiplet) as well as b (broad). Coupling constants (J) are given in hertz. Melting points were determined by using melting point apparatus and are uncorrected. MS spectra were obtained on a GC Ms-QP 1000 EX mass spectrometer at 70 eV. Microanalyses were performed using a C H N S/O analyzer. Elemental data are within $\pm 0.4\%$ of the theoretical values. All yields reported are unoptimized. The chemical reagents

used in synthesis were purchased from Fluka, Sigma and Aldrich.

3.1.2. Preparation of 3-amino-7-substituted-2-carbamoylquinoxaline 1, 4-dioxide (2a-c).

A mixture of 5(6)-substituted benzofuroxane **1a-c** and cyanoacetamide (10 mmol) was stirred for 10 min at 0°C. Over the cooled suspension was added a solution of triethylamine (5 drops) in dimethylformamide. The mixture was allowed to stand at room temperature over 24 h and then filtered off. The solid product was filtered and recrystallized from ethanol.

Compound 2a Yield 71%; mp. 260-265°C. IR (KBr, cm⁻¹): 1660(C=O), 3388, 3224 (NH,NH₂). ¹H NMR spectrum (DMSO-d₆), δ, ppm 5.6 (s, 2H, NH₂, D₂O-exchangable), 8.53 (s, 2H, CO-NH₂, D₂O-exchangable), 7.26-8.2 (m, 4H, quinox-H). Elemental Analysis: calcd, C, 49.09; H, 3.66; N, 25.45. C₉H₈N₄O₃ found C, 49.47; H, 2.96; N, 25.40.

Compound 2b Yield 85%; mp. IR (KBr, cm⁻¹): 1715(C=O), 1586, 1455 (NO), 3356, 3454, 3512 (NH,NH₂). ¹H NMR spectrum (DMSO-d₆), δ, ppm 5.76 (s, 2H, NH₂, D₂O-exchangable), 8.60 (s, 2H, CO-NH₂, D₂O-exchangable), 7.36-8.32 (m, 3H, quinox-H). Elemental Analysis: calcd, C, 42.45; H, 2.77; N, 22.00. C₉H₇ClN₄O₃ found C 42.47; H 2.66; N, 22.12.

Compound 2c Yield 41%; mp 248°C. IR (KBr, cm⁻¹): 1651(C=O), 3200, 3413 (NH,NH₂). ¹H NMR spectrum (DMSO-d₆), δ, ppm 3.93 (s, 1H, OCH₃), 4.9(s, 2H, NH₂-D₂O exchangable), 7.85(s, 2H, CO-NH₂, D₂O-exchangable), 6.97-7.32 (m, 3H, quinox-H). Elemental Analysis: calcd, C, 48.00; H, 4.03; N, 22.39. C₁₀H₁₀N₄O₄ found C 48.27; H 4.46; N, 22.12.

3.1.3. Preparation of 7-chloro-2-mercapto[1,3,2]azathia-phosphorin[4,5-b]quinoxaline-5,10-di-N-oxide-4(1H)-one-2(3H)-thione, Pyridine solvate (3)

Dry pyridine (3 ml) was added slowly and carefully with stirring to carefully ground and stirred mixture of 2-amino-3-carbamoyl-6-chloro quinoxaline 1, 4-dioxide **2b** (2.18g, 0.01 mol) and phosphorus pentasulfide (3.33 g, 0.015 mol). The mixture was heated and changed to oil. After addition of pyridine, the mixture was boiled for 5 min and then washed with boiling benzene (2 × 100 ml); water (200 ml) was added and this was boiled for 2 min and then cooled down. The precipitate was filtered off and carefully washed with water and ethanol. Yield (83%); mp 360°C (DMF-water, 3:1). IR (KBr, cm⁻¹): 1655 (CO), 1625(C=N), 1500, 1358 (NO), 3408(NH), 1244 (C=S). ¹H NMR spectrum (DMSO-d₆), δ, ppm 7.39(1H, d, H₈), 7.50(1H, t, γ-H), 7.58 (1H, s, H₈), 7.73(1H, t, β-H), 7.95(2H, d, α-H), 7.99(1H, d, H₆), 10.18 (1H, br. s, NH, D₂O-exchangable). MS (m/z): 365 (M⁺), 204 (M⁺ - SH, - O₂ - PS₂), 195 (M⁺ - O₂, -NH₂, -CO, - PS₂), 188 (M⁺ - SH, - O₃, - PS₂), 63 (PS⁺). Elemental Analysis: calcd, C, 37.80; H, 2.27; N, 12.59; S, 21.62. C₁₄H₁₀ClN₄O₃PS₃ found C, 37.78; H, 2.31; N, 12.65; S, 21.73

3.1.4. Preparation of 3-amino-7-substituted-2-quinoxaline-carbonitrile -1, 4-di-N-oxides (4a-c) [48].

A mixture of 5(6)-substituted benzofuroxane **1a-c** (10 mmol) and malononitrile (10.6 mmol) was stirred for 10 min

at 0°C. Over the cooled suspension was added a solution of triethylamine (5 drops) in dimethylformamide. The mixture was allowed to stand at room temperature over 24 h and then filtered off. The solid product was washed with diethylether and recrystallized from dioxane.

Compound 4a Yield 65%; m.p. 190°C.

Compound 4b Yield 80%; m.p. 263-264°C.

Compound 4c Yield 31%; m.p. 248-249°C.

3.1.5. Preparation of 7-chloro-2-mercapto[1,3,2]diazaphosphorin[4,5-d]quinoxaline-5,10-di-N-oxide-2,4(1H, 3H)-dithione (5)

A mixture of 2-amino-3-carbamoyl-6-chloro quinoxaline 1,4-dioxide **2b** (2.18g, 0.01 mol) and phosphorus pentasulfide (3.33g, 0.015 mol) in dimethylformamide were carefully ground and stirred. The mixture was boiled for 5 min and then washed with boiling benzene (2 × 100 ml); water (200 ml) was added and this was boiled for 2 min and then cooled down. The precipitate was filtered off and carefully washed with water and ethanol. Yield (34%); mp 160°C. IR (KBr, cm⁻¹): 1655 (CO), 1625(C=N), 1500, 1490 (NO), 3408(NH), 1244(C=S). ¹H NMR spectrum (DMSO-d₆), δ, ppm 7.57(1H, d, H₈), 7.70(1H, s, H₆), 7.96 (1H, d, H₉), 9.18 (1H, br. s, NH, D₂O-exchangable). MS (m/z): 365 (M⁺), 204 (M⁺ - SH, - O₂ - PS₂), 195 (M⁺ - O₂, -NH₂, -CO, - PS₂), 188 (M⁺ - SH, - O₃, - PS₂), 63 (PS⁺). Elemental Analysis: calcd, C, 37.80; H, 2.27; N, 12.59; S, 21.62. C₁₄H₁₀ClN₄O₃PS₃ found C, 37.78; H, 2.31; N, 12.65; S, 21.73.

3.1.6. Preparation of 2-amino-3-carbamothioyl-6-chloro-quinoxaline 1,4-dioxide (6)

The same procedure for preparation of **5**, the major product obtained **6**; Yield (66%); mp 250°C. (ethanol). IR (KBr, cm⁻¹): 3125, 3422 (NH₂), 1254(C=S). ¹H NMR spectrum (DMSO-d₆), δ, ppm 7.48 (1H, d, H₈), 7.70 (1H, s, H₆), 7.96 (1H, d, H₉), 5.42 (1H, br. s, NH₂, D₂O exchangeable). MS (m/z): **270** (M⁺), 255 (M⁺ - NH₂), 163 (M⁺ - O₂, -NH₂, -CS). Elemental Analysis: calcd, C, 39.93; H, 2.61; N, 20.70; S, 11.85. found C, 39.89 ; H, 2.59; N, 20.73; S, 11.79

3.1.7. Preparation of 2-mercapto-7-substituted [1, 3, 2] diazaphosphorin [4,5-b] quinoxaline 5, 10-di-N-oxide - 2,4(1H,3H)-dithione, Pyridine solvate (7a-c)

Dry pyridine (3 ml) was added slowly and carefully with stirring to carefully ground and stirred mixture of 3-amino-7-substituted-2-quinoxalinecarbonitrile-1, 4-di-N-oxides **4a-c** (10 mmol) and phosphorus pentasulfide (15 mmol). The mixture was heated and changed to oil. After addition of pyridine, the mixture was boiled for 5 min and then washed with boiling benzene (2 × 100 ml); water (200 ml) was added and this was boiled for 2 min and then cooled down. The precipitate was filtered off and carefully washed with water and ethanol. **Compound 7a** Yield (63%); mp 160°C (DMF-water, 3:1). IR (KBr, cm⁻¹): 1635 (C=N), 3364, 3317, 3231 (2 NH), 1544, 1484 (NO), 1228(C=S). ¹H NMR spectrum (DMSO-d₆), δ, ppm 10.0 (1H, br. s, NH-P, D₂O exchangable), 10.18 (1H, br. s, NH, D₂O-exchangable), 7.39-7.83 (7H, m, Ar-H and pyridine-H), 7.8(2H, d, H₆, H₉). MS (m/z): 330 (M⁺), 265 (M⁺ - SH, -O₂), 205 (M⁺ - SH, -O₂, -CSNH₂), 170 (M⁺ - SH, - O₂ - PS₂), 144 (M⁺ - O₂, CS-NH-

PS2), 95 (PS2⁺). Found, %: C; H; N; S.C 39.47; H 2.66; N, 13.12; S 30.01. C₁₄H₁₁N₄O₂PS₃. Calc., %: C 39.43; H 2.60; N 13.14; S 30.07.

Compound 7b. Yield (77%); mp 210°C (DMF–water, 3:1). IR (KBr, cm⁻¹): 1615(C=N), 1538, 1496 (NO), 3124,3249,3385(2 NH), 1226 (C=S). ¹H NMR spectrum (DMSO-d₆), δ, ppm 7.41-7.84(6H, m, H₈,H₉ and pyridine-H), 7.85 (1H, s, H₆), 8.59(2H, d, α-H), 10.11 (1H, br. s, NH-P, D₂O-exchangeable), 10.43(1H, br. s, NH, D₂O-exchangeable). MS (m/z): 364 (M⁺), 299 (M⁺-SH, -O₂), 238 (M⁺-SH,-O₂, -CSNH₂), 204 (M⁺-SH, -O₂-PS₂), 95 (PS₂⁺). Elemental Analysis: calcd C, 36.48; H, 2.19; N, 12.15; S, 27.83. C₉H₆ClN₄O₂PS₃ found C, 36.52; H, 2.18; N, 12.33; S, 27.91

Compound 7c Yield (23%); mp 130°C (DMF–water, 3:1). IR (KBr, cm⁻¹): 1623(C=N), 1557, 1490(NO), 3350,3221(2 NH), 1251 (C=S). ¹H NMR spectrum (DMSO-d₆), δ, ppm 3.9(3H, s, OCH₃), 7.39(1H, d, H₈), 7.59(1H, d, H₉), 7.79 (2H, t, β-H), 7.8(1H, s, H₆), 8.06(1H, t, γ-H), 8.59(2H, d, α-H), 10.11 (1H, br. s, NH-P, D₂O-exchangeable), 10.43(1H, br. s, NH, D₂O exchangeable). MS (m/z): 364 (M⁺), 299 (M⁺-SH, -O₂), 238 (M⁺-SH,-O₂, -CSNH₂), 204 (M⁺-SH,-O₂-PS₂), 95 (PS₂⁺). Elemental Analysis: calcd, C, 39.46; H, 2.87; N, 12.27; S, 28.10. C₉H₆ClN₄O₂PS₃ found C, 39.48; H, 2.90; N, 12.32; S, 27.81.

3.1.8. Preparation of 2, 4-dimethylthio-5-methyl-7-substituted [1, 3, 2] diazaphosphorin [4, 5-b]quinoxaline 5,10-di-N-oxide-2- thione (8a,b)

Pyridine solvate **7a, b** (0.01mol) was dissolved in aqueous (1 mol/l) solution of NaOH (80 ml). The solution obtained was washed twice with benzene and cooled down to 0-5°C, and then dimethyl sulfate (5 ml) was added. The reaction mixture was stirred for 30 min, the precipitate was filtered off and washed with water and ethanol.

Compound 8a Yield (54%); mp 240-242°C(DMF–water, 3:1).IR (KBr, cm⁻¹): 1643(C=N), 1243 (P=S). ¹H NMR spectrum (DMSO-d₆), δ, ppm 2.07 (3H, d, P–SCH₃); 2.49 (3H, s, 4-C–SCH₃); 3.22 (3H, d, N¹-CH₃), 7.39(1H, t, H₈), 7.42(1H, d, H₆), 7.63(1H, t, H₇), 7.85(1H, d, H₉), Elemental Analysis: Found, %: C 37.65; H 4.94; N 17.46; S 30.12. C₁₀H₁₅N₄PS₃. Calculated, %: C 37.72; H 4.75; N 17.60; S 30.21.

Compound 8b Yield (43%); mp 262-265°C (DMF–water, 3:1). IR (KBr, cm⁻¹): 1633(C=N), 1444, 1371 (NO), 1243 (P=S). ¹H NMR spectrum (DMSO-d₆), δ, ppm 2.08 (3H, s, P–SCH₃); 2.51 (3H, s, 4-C–SCH₃); 3.28 (3H, s, N¹-CH₃), 7.39(1H, d, H₉), 7.68(1H, d, H₈), 7.73(1H, s, H₆), Elemental Analysis: Found, %: C 37.65; H 4.94; N 17.46; S 30.12. C₁₀H₁₅N₄PS₃. Calculated, %: C 37.72; H 4.75; N 17.60; S 30.21.

3.2. Biological Studies

3.2.1. Cytotoxicity against PC-3 Prostate Adenocarcinoma Cell Line

The cytotoxic inhibitory effect of the new synthesized compounds on human prostate cancer PC-3 was performed at the pharmacology department, Ain Shams University, Cairo, Egypt.

Materials:

RPMI-1640 medium, fetal bovine serum (FBS) and phosphate buffered saline were purchased from Lonza (Basel, Switzerland). Dimethyl sulfoxide and SulphoRhodamine-B (SRB) were obtained from Sigma-Aldrich (St Louis, MO, USA). Dulbecco's Modified Eagle Medium (DMEM) was purchased from Gibco (Grand Island, NY, USA).

Cell culture:

PC3, human prostate cancer cell line was grown in RPMI-1640 medium supplemented with 10% heat inactivated FBS, 100 units/mL of penicillin and 100 mg/mL of streptomycin and maintained at 37° in a humidified atmosphere containing 5% CO₂. The cells were maintained as “monolayer culture” by serial subculturing.

SRB cytotoxicity assay:

Cytotoxicity was determined using SRB method as previously described by Skehan *et al.* (1). Exponentially growing cells were collected using 0.25% Trypsin-EDTA and seeded in 96-well plates at 1000-2000 cells/well in RPMI-1640 supplemented medium. After 24 h, cells were incubated for 72 h with various concentrations of the tested compounds. Following 72 h treatment, the cells will be fixed with 10% trichloroacetic acid for 1 h at 4 °C. Wells were stained for 10 min at room temperature with 0.4% SRB dissolved in 1% acetic acid. The plates were air dried for 24 h and the dye was solubilized with Tris-HCl for 5 min on a shaker at 1600 rpm. The optical density (OD) of each well was measured spectrophotometrically at 564 nm with an ELISA microplate reader (ChroMate-4300, FL, USA).

Data Analysis

The dose response curve of compounds was analyzed using Emax model. Where R is the residual unaffected fraction (the resistance fraction), [D] is the drug concentration used, K_d is the drug concentration that produces a 50% reduction of the maximum inhibition rate and m is a Hill-type coefficient. IC₅₀ was defined as the drug concentration required to reduce fluorescence to 50% of that of the control (i.e., K_d = IC₅₀ when R=0 and Emax =100-R) (2).

$$\% \text{ Cell viability} = (100 - R) \times \left(1 - \frac{[D]^m}{K_d^m + [D]^m} \right) + R$$

3.2.2. Profiling Against the Protein Kinases VEGFR and Src

The evaluation of the new synthesized compounds were performed at Kinexus compound profiling services and evaluation of compounds against protein kinase targets, order no° 2618.

Kinase Activity Assay Protocol

• Materials

Quality Control & Reagents

The various protein kinase targets to be employed in the compound profiling process were cloned, expressed and purified using proprietary methods. Quality control testing is

routinely performed on each of the targets to ensure compliance to acceptable standards. Protein substrates employed in the compound profiling process were synthesized internally, KDR, active (aliases include FLK1 and VEGFR) recombinant human protein expressed in Sf9 cells and Src, active (aliases include ASV, SRC1, c-SRC, and p60-Src) full length recombinant protein expressed in E.coli cells. 33P-ATP was purchased from PerkinElmer. All other materials were of standard laboratory grade. The highly purified active enzymes are generated from the full-length human genes and are mutation free.

• Methods

The assay condition for the various protein kinase targets were optimized to yield acceptable enzymatic activity. In addition, the assays were optimized to give high signal-to-noise ratio.

• Protein Kinase Assays

A radioisotope assay format was used for profiling evaluation of protein kinase targets and all assays are performed in a designated radioactive working area. Blank control was set up for each protein kinase target which included all the assay components except the addition of the appropriate substrate (replaced with equal volume of assay dilution buffer). The corrected activity for each protein kinase target was determined by removing the blank control value.

3.3. Molecular Modeling

All the molecular modeling calculations and docking simulation studies were performed using Molecular Operating Environment (MOE[®]) [57] version 10.2010, Chemical Computing Group Inc., Montreal, Canada. All the interaction energies and different calculations were automatically calculated.

The target compounds were constructed into a 3D model using the builder interface of the MOE program, the target compounds were subjected to a conformational search and all conformers were subjected to energy minimization, all the minimizations were performed. The X-ray crystallographic structures of VEGFR-2 receptor (PDB ID: 1VR2) was obtained from the Protein Data Bank.

CONCLUSIONS

Recently, the concept of angiogenesis was introduced as a pathway in cancer treatment. New derivatives of 1, 3, 2-diazaphosphorin [4, 5-b] quinoxaline-5, 10-di-N-oxide were synthesized and evaluated for the treatment of prostate cancer through inhibition of the tyrosine kinases VEGFR-2 and SRC, which are involved in angiogenesis and metastases of certain tumors. The results of cytotoxicity against human prostate cell line PC-3 showed that, compounds **7a**, **8a** and **5** were the most potent cytotoxins. In particular, 2-mercapto-1, 3, 2-diazaphosphorin [4, 5-b] quinoxaline-5,10-di-N-oxide-2,4-dithione **7a** demonstrated good inhibitory effect on VEGFR-2. However, the activity of this series of compounds did not seem to be mediated through SRC kinase inhibition.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflicts of interest.

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REFERENCES

- [1] Hwang, C.; Heath, E.I. Angiogenesis inhibitors in the treatment of prostate cancer. *J. Hematol. Oncol.*, **2010**, 3(26), 1-12.
- [2] Hubert, R.K.; Randenborgh, H.; Treiber, U.; Wutzler, S.; Battistel, C.; Lehmer, A.; Wagenpfeil, S.; Hartung, R.; Paul, R. *In vitro* cytotoxic effects of imatinib in combination with anticancer drugs in human prostate cancer cell lines. *Prostate*, **2005**, 63, 385-394.
- [3] Petrylak, D.P.; Tangen, C.M.; Hussain, M.H.; Lara, P.N.Jr; Jones J.A.; Taplin, M.E.; Burch, P.A.; Berry, D.; Moynour, C.; Kohli, M.; Benson, M.C.; Small, E.J.; Raghavan, D.; Crawford, E.D. Docetaxel and estramustine compared with mitoxantrone and prednisone for advanced refractory prostate cancer. *N. Engl. J. Med.*, **2004**, 351, 1513-1520.
- [4] Kantoff, P.W.; Halabi, S.; Conaway, M.; Picus, J.; Kirshner, J.; Hars, V.; Trump, D.; Winer, E.P.; Vogelzang, N.J. Hydrocortisone with or without mitoxantrone in men with hormone-refractory prostate cancer: results of the cancer and leukemia group B 9182 study. *J. Clin. Oncol.*, **1999**, 17, 2506-2513.
- [5] Tannock, I.F.; De Wit, R.; Berry, W.R.; Horti, J.; Pluzanska, A.; Chi, K.N.; Oudard, S.; Théodore, C.; James, N.D.; Tureson, I.; Rosenthal, M.A.; Eisenberger, M.A. Docetaxel plus Prednisone or Mitoxantrone plus Prednisone for advanced prostate cancer. *N. Engl. J. Med.*, **2004**, 351, 1502-12.
- [6] De Bono, J.S.; Oudard, S.; Ozguroglu, M.; Hansen, S.; Machiels, J.P.; Kocak, I.; Gravis, G.; Bodrogi, I.; Mackenzie, M.J.; Shen, L.; Roessner, M.; Gupta, S.; Sartor, A. O. Prednisone plus cabazitaxel or mitoxantrone for metastatic castration-resistant prostate cancer progressing after docetaxel treatment: a randomised open-label trial. *Lancet*, **2010**, 376(9747), 1147-1154.
- [7] "Jevtana (cabazitaxel) Injection Approved by U.S. FDA after Priority Review"(Press release). Available from: <http://sanofiaventis.mediaroom.com/index.php?s=43&item=288>. sanofi-aventis. [Access on: 17th June 2010]
- [8] Available from: http://www.centerwatch.com/drug-information/fda-approvals/drug_areas.aspx?AreaID=12
- [9] Drugs@FDA, U.S. Food and Drug Administration. Available from: <http://www.accessdata.fda.gov/scripts/cder/drugsatfda/index.cfm>
- [10] Aragon-Ching J.B.; Dahut, W.L. VEGF inhibitors and prostate cancer therapy. *Curr. Mol. Pharmacol.*, **2009**, 2(2), 161-8.
- [11] Ferrara, N.; Gerber, H.P.; Lecouter, J. The biology of VEGF and its receptors. *Nat. Med.*, **2003**, 9(6), 669-676.
- [12] Ferrara, N. VEGF as a therapeutic target in cancer. *Oncology*, **2005**, 69(3), 11-16.
- [13] Moreira, I.S.; Fernandes, P.A.; Ramos, M.J. Vascular Endothelial Growth Factor (VEGF) Inhibition -A Critical Review. *Anti-Cancer Agents Med. Chem.*, **2007**, 7, 223-245.
- [14] Arora, N.; Rizwan, M.; Zheng, T.; Cai, J.; Smith, D.L.; Parkash, S.G. Vascular endothelial growth factor chimeric toxin is highly active against endothelial cells. *Cancer Res.*, **1999**, 59, 183-188.
- [15] Dreves, J.; Kondering, M.A.; Wolloscheck, T.; Wedge, S. R.; Ryan, A.J.; Ogilvie, D.J.; Esser, N. The VEGF receptor tyrosine kinase inhibitor, ZD6474, inhibits angiogenesis and affects microvascular architecture within an orthotopically implanted renal cell carcinoma. *Angiogenesis*, **2004**, 7(4): 347-354.
- [16] Folkman, J. Angiogenesis in cancer, vascular, rheumatoid and other disease. *Nat. Med.* **1995**, 1(1), 27-31.
- [17] Ruggeri, B.; Singh, J.; Gungir, D.; Angeles, T.; Albom, M.; Chang, H.; Robinson, C.; Hunter, K.; Dobrzanski, P.; Jones-Bolin, S.; Aimone, L.; Klein-Szanto, A.; Herbert, J.; Bono, F.; Schaeffer, P.; Casellas, P.; Bourie, B.; Pili, R.; Isaacs, J.; Ator, M.; Hudkins, R.; Vaught, J.; Mallamo, J.; Dionne, C. A novel, orally active pan inhibitor of vascular endothelial growth factor receptor tyrosine kinases with potent antiangiogenic activity and antitumor efficacy in preclinical models. *Cancer Res.*, **2003**, 63, 5978-5991.
- [18] Carmeliet, P.; Ng, Y.S.; Nuyens, D.; Theilmeier, G.; Brusselmanns, K.; Cornelissen, I.; Ehler, E.; Kakkar, V.V.; Stalmans, I.; Mattot, V.; Perriard, J.C.; Dewerchin, M.; Flameng, W.; Nagy, A.; Lupu, F.; Moons, L.; Collen, D.; D'Amore, P.A.; Shima, D.T. Impaired myocardial angiogenesis and ischemic cardiomyopathy in mice lacking the vascular endothelial growth factor isoforms VEGF164 and VEGF188. *Nat. Med.*, **1999**, 5(5), 495-502.

- [19] Bold, G.; Altmann, K.H.; Frei J.; Lang, M.; Manley, P. M.; Traxler, P.; Wietfeld, B.; Buchdunger, E.; Cozens, R.; Ferrari, S.; Furet, P.; Hofmann, F.; Martiny-Baron, G.; Mestan, J.; Rosel, J.; Sills, M.; Stover, D.; Acemoglu, F.; Boss, E.; Emmenegger, R.; Lasser, L.; Masso, E.; Roth, R.; Schlachter, C.; Vetterli, W.; Wyss, D.; Wood, J. W. New anilinothalazines as potent and orally well absorbed inhibitors of the VEGF receptor tyrosine kinases useful as antagonists of tumor-driven angiogenesis. *J. Med. Chem.*, **2000**, *43*(12), 2310-2323.
- [20] Jain, R.K.; Duda, D.G.; Clark, J.W.; Loeffler, J.S. Lessons from phase III clinical trials on anti-VEGF therapy for cancer. *Nat. Clin. Pract. Oncol.*, **2006**, *3*, 24-40.
- [21] Aragon-Ching, J.B.; William, L. Dahut. VEGF inhibitors and prostate cancer therapy. *Curr. Mol. Pharmacol.*, **2009**, *2*(2), 161-168.
- [22] Huss, W.J.; Barrios, R.J.; Greenberg, N.M. SU5416 selectively impairs angiogenesis to induce prostate cancer-specific apoptosis. *Mol. Cancer Ther.*, **2003**, *7*, 611-616.
- [23] Mohamedali, K.A.; Poblens, A.T.; Sikes, C.R.; Navone, N.M.; Thorpe, P.E.; Darnay, B.G.; Rosenblum, M.G. Inhibition of prostate tumor growth and bone remodeling by the vascular targeting agent VEGF121/rGel. *Cancer Res.*, **2006**, *66*(22), 10855-10860.
- [24] Bok, R.A.; Halabi, S.; Fei, D.T.; Rodriguez, C.R.; Hayes, D.F.; Vogelzang, N.J.; Kantoff, P.; Shuman, M.A.; Small, E.J. Vascular endothelial growth factor and basic fibroblast growth factor urine levels as predictors of outcome in hormone-refractory prostate cancer patients: a cancer and leukemia group B study. *Cancer Res.*, **2001**, *61*, 2533-2536.
- [25] Ferrer, F.A.; Miller, L.J.; Andrawis, R.I.; Kurtzman, S.H.; Albertsen, P.C.; Laudone, V.P.; Kreutzer, D.L. Vascular endothelial growth factor (VEGF) expression in human prostate cancer: in situ and *in vitro* expression of VEGF by human prostate cancer cells. *J. Urol.*, **1997**, *157*, 2329-2333.
- [26] Eun-mi, Y.; Maneesh J.; Jeanny B.A. Angiogenesis inhibitors in prostate cancer. *Discov. Med.*, **2010**, *10*(55), 521-30.
- [27] Wu, H.; Huang, C.; Chang, D. Anti-angiogenic therapeutic drugs for treatment of human cancer. *J. Cancer Mol.*, **2008**, *4*(2), 37-45.
- [28] Merino, M.; Pinto, A.; Gonzá'lez, R.; Espinos, E. Antiangiogenic agents and endothelial antagonists in advanced castration resistant prostate cancer. *Eur. J. Cancer*, **2011**, *47*, 1846-1851.
- [29] Solano, B.; Junntola, V.; Mari'n, A.; Villar, R.; Burguete, A.; Vicente, E.; Pe'rez-Silanes, S.; Aldana, I.; Monge, A.; Dutta, S.; Sarkar, U.; Gates, K.S. Synthesis and biological evaluation of new 2-Arylcarbonyl-3-trifluoromethylquinoxaline 1,4-Di-N-oxide derivatives and their reduced analogues. *J. Med. Chem.* **2007**, *50*, 5485-5492.
- [30] Hu, Y.; Xia, Q.; Shangguan, S.; Liu, X.; Hu, Y.; Sheng, R. Synthesis and biological evaluation of 3-Aryl-quinoxaline-2-carbonitrile 1,4-Di-N-oxide derivatives as hypoxic selective anti-tumor agents. *Molecules*, **2012**, *17*, 9683-9696.
- [31] Sheng, R.; Xu, Y.; Weng, Q.; Xia, Q.; He, Q.; Yang, B.; Hu Y. Synthesis and cytotoxic activity of 3-phenyl-2-thio-quinoxaline 1,4-dioxide derivatives in hypoxia and in normoxia. *Drug Discov. Ther.*, **2007**, *1*(2), 119-123.
- [32] Weng, Q.; Wang, D.; Guo, P.; Fang, L.; Hu, Y.; He, Q.; Yang, B. Q39, a novel synthetic Quinoxaline 1, 4-Di-N-oxide compound with anti-cancer activity in hypoxia. *Eur. J. Pharmacol.*, **2008**, *581*(3), 262-9.
- [33] Miyake, K.; Nishioka, M.; Imura, S.; Batmunkh, E.; Uto, Y.; Nagasawa, H.; Hori, H.; Shimada, M. The novel hypoxic cytotoxin, TX-2098 has antitumor effect in pancreatic cancer; possible mechanism through inhibiting VEGF and hypoxia inducible factor-1 α targeted gene expression. *Exp. Cell. Res.*, **2012**, *318*(13), 1554-1563.
- [34] Emmenegger, U.; Morton, G. C.; Francia, G.; Shaked, Y.; Franco, M.; Weirnerman, A.; Man, S.; Kerbel, R.S. Low-dose metronomic daily cyclophosphamide and weekly tirapazamine: a well-tolerated combination regimen with enhanced efficacy that exploits tumor Hypoxia. *Cancer Res.*, **2006**, *66*(3), 1630-1639.
- [35] Roskoski, R.Jr. Src protein-tyrosine kinase structure and regulation. *Biochem. Biophys. Res. Commun.*, **2004**, *324*(4), 1155-1164.
- [36] Saad, F.; Lipton, A.; SRC kinase inhibition: targeting bone metastases and tumor growth in prostate and breast cancer. *Cancer Treat. Rev.*, **2010**, *36*(2), 177-184.
- [37] Chang, Y.; Kung, H.; Evans, C.P. Nonreceptor tyrosine kinases in prostate cancer. *Neoplasia*, **2007**, *9*(2), 90-100.
- [38] Gray, M.J.; Zhang, J.; Ellis, L.M.; Semenza, G.L.; Evans, D.B.; Watowich, S.S.; Gallick, G.E. HIF-1 α , STAT3, CBP/p300 and Ref-1/APE are components of a transcriptional complex that regulates Src-dependent hypoxia-induced expression of VEGF in pancreatic and prostate carcinomas. *Oncogene*, **2005**, *24*, 3110-3120.
- [39] Edwards, J. Src kinase inhibitors: an emerging therapeutic treatment option for prostate cancer. *Expert Opin. Invest. Drugs*, **2010**, *5*, 605-614.
- [40] Park, S.I.; Zhang, J.; Phillips, K.A.; Araujo, J.C.; Najjar, A.M.; Volgin, A.Y.; Gelovani, J.G.; Kim, S.J.; Wang, Z.; Gallick, G.E. Targeting SRC family kinases inhibits growth and lymph node metastases of prostate cancer in an orthotopic nude mouse model. *Cancer Res.*, **2008**, *68*(9), 3323-3333.
- [41] Nam, S.; Kim, D.; Cheng, J.Q.; Zhang, S.; Lee, J.H.; Buettner, R.; Mirosevich, J.; Lee, F.Y.; Jove, R. Action of the Src family kinase inhibitor, dasatinib (BMS-354825), on human prostate cancer cells. *Cancer Res.*, **2005**, *65*(20), 9185-9189.
- [42] John Bull, E.O.; Naidu, M.S.R. Isoquino [2, 1-c] [1, 3, 2] benzodiazaphosphorine derivatives: new potential agents for cancer chemotherapy. *Phosphorus Sulfur Silicon Relat. Elem.*, **2000**, *162*(1), 231-243.
- [43] Acheson, R.M.; Lines, C.T.; Bryce, M.R.; Dauter, Z.; Reynolds, C.D.; Schmidpeter, A. Synthesis and X-ray crystal structure of 2,3-dihydro-2-mercapto-2,1,3-benzophosphadiazine-4(1H)-thione 2-sulphide derivatives. *J. Chem. Soc. Perkin. Trans. II*, **1985**, 1913-1917.
- [44] Bryce, M.R.; Mathews, R.S. Synthetic and NMR spectroscopic studies on the 2,1,3-benzophosphodiazine ring system. *J. Organomet. Chem.*, **1987**, *325*, 153-157.
- [45] Nilov, D.B.; Kadushkin, A.V.; Soloveva, N.P.; Grank, V.G. An unexpected synthesis of 7,8-polymethyleneimidazo-1,3,2-diazaphosphorines-heteroanalogues of mercaptopurine derivatives. *Mendeleev Commun.*, **1995**, *2*, 67.
- [46] Nilov, D.B.; Solov'eva, N.P.; Nikolaeva, S.; Peters, V.V.; Krylova, L.Y.; Gus'kova, T.A.; Granik, V.G. Synthesis and antiviral activity of Pyrazolo [3,4-d]-1,3,2-Diazaphosphorin. *Pharm. Chem. J.*, **1998**, *32*(7), 16-19.
- [47] Nilov, D.B.; Kadushkin, A.V.; Solov'eva, N.P.; Sheinker, Y.N.; Granik, V.G. Synthesis and study of the properties of 7,8-Polymethyleneimidazo [4,5-d]-1,3,2-Diazaphosphorin-2-thiones. *Chem. Heterocycl. Comp.*, **2004**, *40*(1), 106-113.
- [48] Monge, A.; Palop, J.A.; Lopez de Cerain, A.; Senador, V.; Martínez Crespo, F.J.; Sainz, Y.; Narro, S.; DeMiguel, Garcia, E.C.; Gonzalez, M.; Hamilton, E.; Barker, A.J.; Clarke, E.D.; Greenhow, D.T. Hypoxia-selective agents derived from quinoxaline 1,4-di-N-oxides. *J. Med. Chem.*, **1995**, *38*, 1786-1792.
- [49] Monge, A.; Palop, J.A.; Gonzalaz, M.; Martínez Crespo, F.J.; Lopez de Cerain, A.; Sainz, Y.; Narro, S. New Hypoxia-Selective Cytotoxins Derived From Quinoxaline 1,4-dioxides. *J. Heterocycl. Chem.*, **1995**, *32*(4), 1213-1217.
- [50] Haddadin, M.J.; Agopian, G.; Issidorides, C.H. Synthesis and photolysis of some substituted quinoxaline di-N-oxides. *J. Org. Chem.*, **1971**, *36*, 514-518.
- [51] Du, J.; Lei, B.; Qin, J.; Liu, H.; Yao, X.; Molecular modeling studies of vascular endothelial growth factor receptor tyrosine kinase inhibitors using QSAR and docking. *J. Mol. Graph. Model.*, **2009**, *27*(5), 642-654.
- [52] Skehan, P.; Storeng, R.; Scudiero, D.; Monks, A.; McMahon, J.; Vistica, D.; Warren, J.T.; Bokesch, H.; Kenney, S.; Boyd M.R. New colorimetric cytotoxicity assay for anticancer-drug screening. *J. Natl. Cancer Inst.*, **1990**, *82*, 1107-1112.
- [53] Al-Abd, A.M.; Lee, J.H.; Kim, S.Y.; Kun, N.; Kuh. H.J. Novel application of multicellular layers culture in situ evaluation of cytotoxicity and penetration of paclitaxel. *Cancer Sci.*, **2008**, *99*, 423-443.
- [54] Neaz, M.M.; Pasha, F.A.; Muddassar, M.; Lee, S.H.; Sim, T.; Hah, J.-M.; Cho, S.J. Pharmacophore based 3D-QSAR study of VEGFR-2 inhibitors. *Med. Chem. Res.*, **2009**, *18*, 127-142.
- [55] Du, J.; Lei, B.; Qin, J.; Liu, H.; Yao, X. Molecular modeling studies of vascular endothelial growth factor receptor tyrosine kinase inhibitors using QSAR and docking. *J. Mol. Graph. Model.*, **2009**, *27*, 642-654.
- [56] RCSB Protein Data Bank. Available from: <http://www.pdb.org>

- [57] Molecular Operating Environment 2008.10 (MOE), Chemical Computing Group Inc., Montreal, Quebec, Canada. Available from: <http://www.chemcomp.com>
- [58] Jun, C.; Chun-Quan, S.Z.; Hui, C.; Yao-Wu, L.; Jia-Guo, L.; Wan-Nian, Z.; You-Jun, Z.; Ju, Z. Study of properties of VEGFR2 active site and binding mode of VEGFR2 and its inhibitors. *Acta Chim. Sin.*, **2007**, *65*(6), 547-552.
- [59] Oguro, Y.; Miyamoto, N.; Okada, K.; Takagi, T.; Iwata, H.; Awazu, Y.; Miki, H.; Hori, A.; Kamiyama, K.; Imamura, S. *Design, synthesis, and evaluation of 5-methyl-4-phenoxy-5H-pyrrolo-[3,2-d] pyrimidine derivatives: Novel VEGFR2 kinase inhibitors binding to inactive kinase conformation.* *Bioorg. Med. Chem.*, **2010**, *18*, 7260-7273.
- [60] Gangjee, A.; Yang, J.; Ihnatb, M.A.; Kamatb, S. Antiangiogenic and antitumor agents: design, synthesis, and evaluation of novel 2-Amino-4-(3-bromoanilino)-6-benzylsubstituted Pyrrolo [2,3-d] pyrimidines as inhibitors of receptor tyrosine kinases. *Bioorg. Med. Chem.*, **2003**, *11*, 5155-5170.

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