

Biological Activity of Phenolic Compounds from Argentinean Herbs Infusions

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Abstract: Argentinean herbal infusions were studied for their beneficial biological properties and secondary metabolites content, such as phenolic compounds in order to find new natural products beneficial. The modifications of antihypertensive and antioxidant activities by the addition of commonly used additives (lemon, sugar and sweetener) were studied also.

Ilex paraguaiensis infusion showed the highest concentration of phenolic compounds and flavonoid fraction, the supplementation with lemon juice increased the phenolic compound content. All infusion showed high DPPH radical scavenging assays and additives did not modify significantly this activity. The infusion of *I. paraguaiensis* presents the maximum viability reduction of *Escherichia coli* and *Staphylococcus aureus* and the herbs combination *Lippia integrifolia-Ilex paraguaiensis* was the most effective to reduce *S. aureus* and *E.coli* viability. The phenolic compounds concentration as well as the qualitative composition of each infusion plays an important role in the antimicrobial activity. Because the high correlation between the concentration of total phenolic compounds and antioxidant and antihypertensive activities, we can infer that the phenolic compounds are responsible for such activities. The results obtained allow us to propose the herbs studied as potential natural antioxidants and antihypertensive compounds that could be used in pharmaceutical and food industries.

Keywords: Antibacterial capacity, antihypertensive properties, antioxidant activity, herbs infusions, phenolic compounds.

INTRODUCTION

Tea (*Camellia sinensis*) is one of the most consumed beverages worldwide after water and as a result, a variety of herbs or herb mixtures have emerged.

Herbs and their derivatives are used in nutrition, medicine, cosmetics fragrances, repellents, flavouring and beverages [1]. The herb infusions are an important source of phenolic compounds [2-4]. Herbs contain phenolic compounds, which are bioactive substances in vegetables, food and derived products because of their human health protecting capacity.

The chemical structure and the biodistribution of the phenolic compounds in the body are responsible for the biological properties and bioavailability of these compounds [5, 6]. No studies about the antihypertensive activity of Argentinean herb infusions or the modification produced in antihypertensive or antioxidant activities due to the use of additives in herb infusions were reported till now.

The aim of this study was to investigate the antihypertensive, antioxidant and antibacterial properties of different Argentinean herbs infusions consumed traditionally; and the modifications for the use of three

additives (sugar, sweetener and lemon) commonly used during herbal infusion consumption in Argentine during these activities.

MATERIALS AND METHODS

Argentinean Herb Samples

Herbs samples were obtained commercially from pharmacy in Argentinean. *Lippia integrifolia*, *Mentha piperita*, *Lippia turbinata*, *Wendtia Calysina*, *Chenopodium ambrosioides*, *Minthostachys verticillata*, *Peumus boldus*, *Aloysia citriodora* and *Ilex paraguaiensis* herbs were selected. A combination of herbs called digestive, contain: *Lippia integrifolia* 20%, *Lippia turbinata* 20%, *Wendtia calysina* 20%, *Chenopodium ambrosioides* 20% and *Minthostachys verticillata* 20%. Lemon, sugar and sweetener were obtained from local commercial.

Herbs Infusions Elaboration

Herbal infusions were prepared as consumed in Argentinean. Individual herbs (2 g) were added to (250 mL) of water heated to 90°C for 10 min. The infusions were filtered using cotton. An aliquot of the infusions was clarified using activated charcoal (30 mg/L) and sterile filtered.

Selected herb mixtures were elaborated by the same procedure, weighing 1 g of each herb and then adding 250

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mL boiling water. A spoonful of lemon, sugar or sweetener was added to the infusion.

For the tests of antimicrobial activity, different herbs infusions were concentrated, weighing 4 g of each herb and dissolved in 250 mL boiling water.

Concentration of Phenolic Compounds in Herb Infusions

To determine the concentration of total phenolic compounds, the method described by Singleton and Rossi [7] was used. The measurements were carried out in triplicate and results were expressed as milligram of gallic acid equivalents per liter (GAE) using a standard curve of gallic acid.

To discriminate the concentration of phenolic acids and flavonoid compounds in infusions, 10 mL of infusion was mixed with 10 mL of diluted solution of HCl (1:3) and 5 mL formaldehyde solution (8 mg/mL). The mixture was incubated 24 hours at room temperature in order to precipitate the flavonoid fraction [8]. The concentration of phenolic acids in the supernatant was determined using the technique of Singleton and Rossi, while the concentration of flavonoid compounds was determined by difference between the concentration of total phenolic compounds and phenolic acids. The results were expressed as mg/L of gallic acid equivalents (GAE). The phenolic compounds concentrations were confirmed by HPLC analysis, using a Knauer Smartline system chromatograph. The chromatographic conditions were as follows: repositil-Pur ODS-3 (250 x 4.6 mm), flow rate 1.0 mL/min, injection volume 20 µL, detection wavelength 280 nm. Phase A (phosphoric acid 0,1%) and phase B (CH₃CN).

ACE-Inhibitory (ACEI) Activity

ACEI activity was determined using the method described by Cushman and Cheung [9] and later modified by Hernández-Ledesma *et al.* [10]. This technique is based on the quantification of hippuric acid formed by the reaction of hippuryl-histidyl-leucine (HHL) with ACE in the presence and absence of an inhibitor. An aliquot of 110 µL of substrate, 10 mM HHL dissolved in buffer (0.2 M phosphate and 0.3 M NaCl, pH 8.3), and 25 µL of ACE (26 mU dissolved in 50 % glycerol) were added to 15 µL of samples (source of ACE inhibitor). The reaction mixture was incubated at 37°C for 80 min and then the enzyme reaction was stopped by the addition of 110 µL of 1N HCl. The hippuric acid formed was extracted with 1 mL of ethyl acetate, shaken and subsequently centrifuged at 5000 g for 10 min. A 750-µL of the organic layer was dried at 45°C in a vacuum chamber (-60 cm Hg) for 60 min. The residue was redissolved in 1 mL of distilled water and absorbance was measured at 228 nm. A reaction blank was obtained by the addition of HCl before ACE enzyme activity. Interference of compounds with absorbance at this wavelength was eliminated with a sample blank. ACEI activity is expressed as follows:

$$\% \text{ of ACEI} = 100[(A-B) - (C-D)] / (A-B)$$

Where, A represents the absorbance in the presence of ACE, B the absorbance of the reaction blank, C the absorbance in

the presence of ACE and inhibitor and D absorbance of the sample blank.

Antioxidant Activity

Ferric-reducing Antioxidant Power Assay (FRAP).

The ferric-reducing ability (FRAP) of the samples was determined using the proposed method by Benzie and Strain [11] with some modifications. The method measures the ability of the sample to reduce a complex of ferric iron with tripyridyl-s-triazine molecule (TPTZ) in the ferrous form. The intensity of color is proportional to the reducing activity of the sample, which is measured spectrophotometrically at 593 nm. A standard curve of ferrous sulfate (100-3500 µM FeSO₄) was used. The FRAP reagent was freshly prepared daily by mixing acetate buffer (pH 3.6), 10 mmol TPTZ solution in 40 mmol HCl and 20 mmol Fe (III) chloride solution in the proportions of 10:1:1 (v/v), respectively. The reagent was warmed to 37°C in a water bath prior to use. Fifty microliters of the samples were added to 1.5 mL of FRAP reagent and incubated for 4 minutes. Measurements were made in triplicate and the results expressed as µmol/L FeSO₄.

Antioxidant activity of the samples was measured according to the free radical scavenging capacity DPPH (1,1-diphenyl-2-picrylhydrazyl), using the method proposed by Von Gadow *et al.* [12]. An aliquot of 50 µL of sample was mixed with 2 ml of a methanol solution of DPPH radical. Absorbance measurements were performed immediately and after 15 minutes of incubation at 517 nm. Methanolic solutions of trolox were tested. All measurements were made in triplicate.

Antioxidant activity of the samples was also measured according to the free radical scavenging capacity ABTS-(2,2'-azinobis-(3-ethylbenzothiazoline-6-sulfonate)), using the method proposed by Re *et al.* [13], with slight modifications. Absorbance was measured after 6 minutes incubation at 734 nm. Standard curve of ascorbic acid (0 - 20 mmol/L) was also evaluated against the radical. Results were expressed as scavenged activity of ABTS radical (%), equivalent in ascorbic acid.

Antibacterial Activity

Microorganisms and culture conditions. The bacterial strain used was *Escherichia coli* ATCC 35218. The bacteria used in each experiment were cultured at 37°C in nutrient broth and agar broth containing in g/L: beef extract, 3.0; peptone, 5.0; sodium chloride, 8.0 and agar 15.0. Before each experiment the bacteria were cultured in liquid medium for 24 h and used as source of inoculums.

The minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) were determined following the guidelines recommended by the CLSI (Clinical and Laboratory Standard Institute) normative [14].

To determine the MIC and MBC Mueller Hilton broth (MHB) were used. The final concentration of bacteria in each macrobroth dilution tube was approximately 5×10^5 cfu/ml of MHB. Serial dilutions of herbal infusions were carried out. MIC is interpreted as the lowest concentration of

Table 1. Antioxidant activity of herb infusions and the modification by the addition of Lemon, sugar or sweetener.

	FRAP ($\mu\text{mol/L FeSO}_4$)				DPPH (% depuration radical DPPH)				ABTS (% depuration radical ABTS)			
	Herb infusion	Lemon	Sugar	Sweetener	Herb infusion	Lemon	Sugar	Sweetener	Herb infusion	Lemon	Sugar	Sweetener
<i>Lippia integrifolia</i>	1520	1850	980	920	72.0	77.9	71.1	70.4	70.0	76.9	69.1	69.4
<i>Mentha piperita</i>	1250	1450	1240	1230	72.5	78.1	71.6	71.4	70.5	77.1	70.6	70.4
<i>Lippia turbinata</i>	0	0	0	0	25.9	31.0	20.4	21.3	25.9	32.0	19.4	20.3
<i>Wendtia calysina</i>	880	1240	850	830	55.3	62.2	53.9	51.8	44.3	49.2	43.9	43.8
<i>Chenopodium ambrosioides</i>	0	0	0	0	21.5	26.3	15.3	15.4	16.5	21.3	12.3	10.4
<i>Peumus boldus</i>	0	0	0	0	30.2	36.4	29.7	28.1	29.2	36.4	27.7	28.1
<i>Aloysia citriodora</i>	600	800	480	490	71.8	79.5	71.1	69.5	71.8	80.5	71.1	69.5
<i>Ilex paraguayensis</i>	940	1100	700	920	73.0	78.8	72.8	71.3	73.0	79.8	72.8	71.3
<i>digestive herbs</i>	3100	3400	1860	1950	86.5	94.6	80.7	79.1	85.5	94.0	82.7	82.0
<i>LI / MP</i>	0	0	0	0	20.4	27.4	21.5	20.2	22.4	30.4	25.5	25.2
<i>LI / IP</i>	1380	1720	940	1040	74.2	80.4	66.5	70.5	76.2	82.4	66.5	70.5
<i>MP / IP</i>	1900	2000	1720	1680	75.9	80.6	71.1	73.1	77.9	82.6	71.1	73.1

phenolic compounds which produce growth inhibition of the microorganism (no evident turbidity) after 24 h of incubation at 37°C. From the dilutions in which no growth was observed, an aliquot of 50 μL are inoculated MHB-agar plates. The plates are incubated for 24 h at 37°C. The MBC is the minimum concentration of phenolic compounds present in herbal infusions needed to produce 99.9 % kill of the initial inoculums. Assays were performed in triplicate and the MIC and MBC determined in herbal infusions treated with activated charcoal were used as control.

Statistical Analysis

The experiments were performed in triplicate and statistical analysis was carried out using MS-Excel software.

RESULTS AND DISCUSSION

Phenolic Compounds Concentrations

Fig. (1) shows the differences in the total phenolic compounds (a), phenolic acid (b) and flavonoid (c) compounds concentrations between herb infusions and herbs infusions added with additives such as lemon, sugar or sweetener. In a previous work, Rodríguez Vaquero *et al.* [15] described that the total phenolic concentration in herb infusions ranged from 50.3 to 925.0 mg GAE/L, *Ilex paraguayensis*, *Lippia integrifolia* and *Mentha piperita*

had the highest concentration, whereas *Lippia turbinata* and *Chenopodium ambrosioides* had the lowest phenolic compounds concentration.

In all infusions concentration, majority flavonoid compounds were compared to the concentration of phenolic acids. The removal of phenolic compounds from the infusions was effective after the clarification process. Our results showed that the addition of lemon increased around 10 % the phenolic compound concentration in all cases. The addition of sweetener slightly decreased the concentration of phenolic compounds in all herbs infusions, but this decrease was not significant and it could be some interference between compounds. The addition of sugar did not modify significantly the phenolic concentration of herb infusions.

Antihypertensive activity. Fig. (2) shows the antihypertensive activity of individual herb infusions and mixtures of herb infusions. Results show that ACEI activity of *Ilex paraguayensis*, *Lippia integrifolia*, *Mentha piperita* and the three selected combinations was higher than 80%. The lowest activity was shown in the presence of *Chenopodium ambrosioides* and *Mintostachys verticillata* (between 15 and 37%). The extraction of phenolic compounds of herb infusions produced the total loss of the ACEI activity, demonstrating that phenolic compounds present in Argentinean herb infusions were the responsible of this activity. The addition of lemon increased the ACEI activity to the herb infusions preparation around 10 %; this

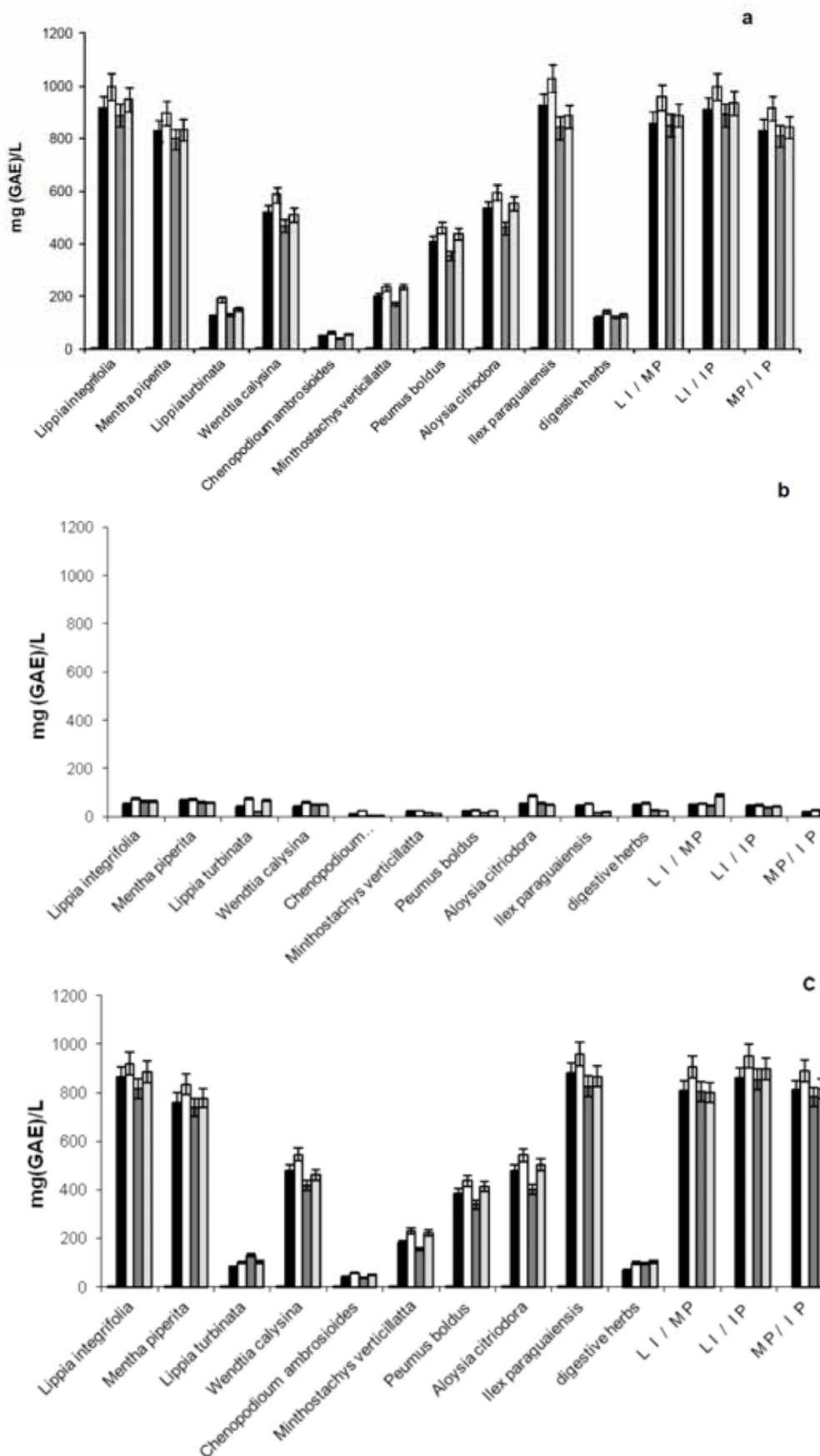


Fig. (1). Total phenolic compounds (a), phenolic acids (b) and flavonoids (c) concentrations in Argentinean herb infusions without additives (■), with lemon (□), sugar (■) or sweetener (□). All determination was at least at triplicate.

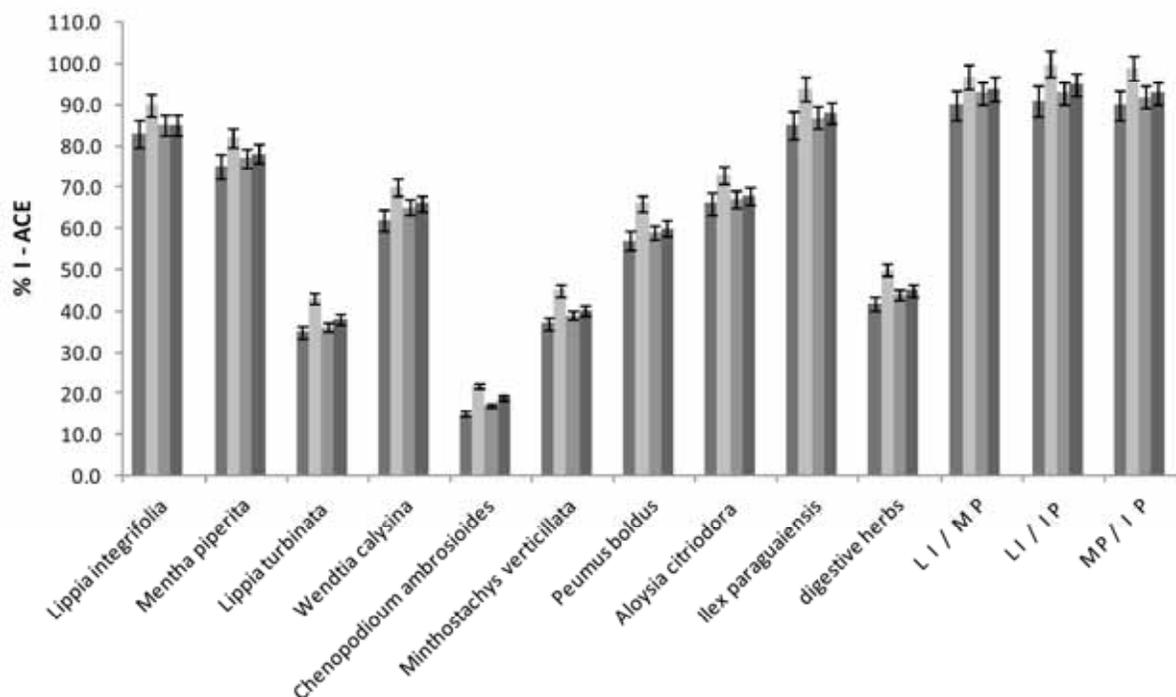


Fig. (2). Angiotensin I-converting enzyme (ACE) inhibition (%) by herb infusions (■), herb infusions added with lemon (□), sugar (▒) or sweetener (▓). All determination was at least at triplicate.

could be due to the phenolic compounds found in lemon, but the addition of sugar or sweetener did not modify this activity.

Our results show a high correlation between the phenolic concentration and the antihypertensive activity and this fact was in agreement with Vallejo *et al.* [18] that demonstrated a good correlation between the phenolic compounds concentrations in juice fruit and the antihypertensive activity.

Antioxidant capacity. Three methods (FRAP, DPPH and ABTS radical scavenging assays) have been used to measure the effect of the additives in herb infusions on the antioxidant activity (Fig. 3).

In a previous work, Rodríguez Vaquero *et al.* [15] reported that *Ilex paraguayensis* possesses the highest ferric reducing power and *Peumus boldus* the lowest ferric reduced power values, whereas it was not detected in *Lippia turbinata*, *Chenopodium ambrosioides* and *Minthostachys verticillata* infusions. The greatest DPPH free radical scavenging activity was found with *Ilex paraguayensis*, whereas *Lippia turbinata*, *Chenopodium ambrosioides* and *Minthostachys verticillata* showed a lowest radical scavenging activity. The radical scavenging activity on clarified infusions was lower than 8%.

Our results showed that infusions prepared with *Ilex paraguayensis*, *Lippia integrifolia*, *Mentha piperita*, *Aloysia citriodora*, *Peumus boldus*, and the three combinations selected possess a great free ABTS radical scavenging activity (higher than 70%). *Lippia turbinata*, *Wendtia calysina*, *Chenopodium ambrosioides*, *Minthostachys verticillata* and *digestive herbs* show the lowest a free ABTS radical scavenging activity (lower than 50%).

The antioxidant activity determinants by the three methods increased this activity with the addition of lemon to herb infusions preparation and the addition of sugar or sweetener did not modify this activity.

We found a high correlation between phenolic compounds concentration in herb infusions and beneficial so we are in agreement with results observed by Li *et al.* [16] with phenolic compounds of Chinese medicinal plants and with those of Yoo *et al.* [17] that found a higher correlation between antioxidant capacity and total phenolic and total flavonoids content in commercial herbs.

Antibacterial property. The modifications in MIC and MBC against *E. coli* of selected herb infusions by additives are presented in Table 2. *Ilex paraguayensis*, *Lippia integrifolia*, *Mentha piperita* and *Peumus boldus* infusions produced inhibition in the growth of *E. coli*, whereas *Wendtia calysina* and *Aloysia citriodora* did not modify the growth of this bacterium. The addition of lemon produces an increase in the growth inhibition observed with *Ilex paraguayensis* and *Lippia integrifolia* infusion, but did not modify the inhibition observed with *Mentha piperita* or *Peumus boldus* infusions. Only *Ilex paraguayensis* and *Lippia integrifolia* infusions produced cellular death and this effect was increased by 50% with the addition of lemon. With the addition of sugar or sweetener a slight increase in the growth of the bacterium was observed, may be due to the consumption of sugar or sweetener.

With clarified infusions, inhibition of the growth of *E. coli* was not observed, so the antibacterial effect was attributed to the phenolic compounds in the infusions.

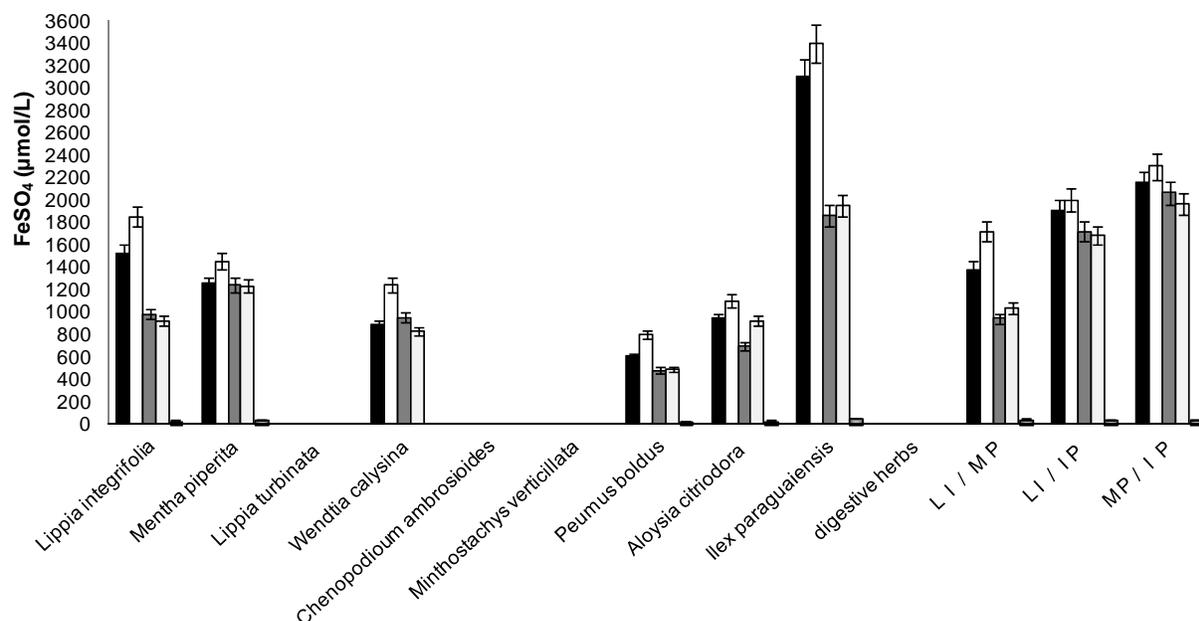


Fig. (3). Antioxidant capacity of herbs infusions. Ferric Reducing Antioxidant Power of herb infusions ($\mu\text{mol/L FeSO}_4$) (a), Percentage inhibition of DPPH radical (%) (b) and Percentage inhibition of DPPH radical (%).

Table 2. Modification of the MIC and MBC of selected infusions against *E. coli* by the addition of additives to herb infusions.

	Herb Infusions								Clarified Herb Infusions							
	MIC				MBC				MIC				MBC			
	Herb infusion	Lemon	Sugar	Sweetener	Herb infusion	Lemon	Sugar	Sweetener	Herb infusion	Lemon	Sugar	Sweetener	Herb infusion	Lemon	Sugar	Sweetener
<i>Lippia integrifolia</i>	916*	468	nd	nd	916	468	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd
<i>Mentha piperita</i>	830	830	nd	nd	830	415	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd
<i>Wendtia calysina</i>	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd
<i>Peumus boldus</i>	409	409	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd
<i>Aloysia citriodora</i>	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd
<i>Ilex paraguayensis</i>	925	462.5	nd	nd	925	462.5	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd

*mg of equivalent of gallic acid (GAE) / L, nd: not detected.

We are in agreement with Vallejo *et al.* [18] who reported that the antioxidant, antihypertensive and antimicrobial activity of Argentinean juice fruit was related with the phenolic compound concentration, but also with the phenolic profile. Rodríguez Vaquero *et al.* [19] reported that *E. coli* was sensible to commercial phenolic compound and to phenolic compounds from red wines.

Our results indicate that the addition of a spoonful of lemon is recommended, this fact increases the phenolic concentration in herb infusions and increases beneficial

activities of the infusion to human health such as the antioxidant, antihypertensive activities and inhibits the growth of *E. coli* on herb infusions.

The biological properties of the Argentinean herbs infusions demonstrated in this study, particularly from *Ilex paraguayensis* and *Lippia integrifolia*, could enable their use as potential natural antioxidants and antihypertensive agents with application in the food and pharmaceutical industries, to favoring and/or replacing the biological activities of the synthetic drugs.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflicts of interest.

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REFERENCES

- [1] Djeridane, A.; Yousfi, M.; Nadjemi, B.; Boutassouna, D.; Stocker, P.; Vidal, N. Antioxidant activity of some Algerian medicinal plants extracts containing phenolic compounds. *Food Chem*, **2006**, *97*, 654-660.
- [2] Moraes-De-Souza, R.A.; Oldoni, T.L.C.; Cabral, I.S.R.; Alencar, S.M. Compostos fenólicos totais e atividade Antioxidante de chás comercializados no Brasil. *Boletimdo Centro Pesquisa Processamento de Alimentos*, **2011**, *29*, 229-236.
- [3] Ruiz-Terán, F.; Medrano-Martínez, A.; Navarro-Ocaña, A. Antioxidant and free radical scavenging activities of plant extracts used in traditional medicine in Mexico. *Afri J Biotechnol*, **2008**, *7*, 1886-1893.
- [4] Silva, E.M.; Souza, J.N.S.; Rogez, H.; Rees, J.F.; Larondelle, Y. Antioxidant activities and polyphenol contents of fifteen selected plant species from the Amazonian region. *Food Chem*, **2007**, *101*, 1012-1018.
- [5] Moller, P.; Loft, S. Dietary antioxidants and beneficial effects on oxidatively damaged DNA. *Free Radic Biol Med*, **2006**, *41*, 388-415.
- [6] Monteiro, M.; Farah, A.; Perrone, D.; Trugo, L.C.; Donangelo, C. Chlorogenic acid compounds from coffee are differentially absorbed metabolized in humans. *Nutrition*, **2007**, *37*, 2196-2201.
- [7] Singleton, V.L.; Rossi, J.A. Colorimetry of total phenolics with phosphomolybdic phosphotungstic acid reagents. *Am J Enol Vitic*, **1965**, *16*, 144-158.
- [8] Zoecklein, B.W.; Fugelsang, K.C.; Gump, B.H.; Nury, F.S. Phenolic compounds and wine color. In: Production wine analysis. Ed. Zoecklein, Bruce. Van Nostrand Reinhold. New York, **1990**, 129-168.
- [9] Cushman, D.W.; Cheung, H.S. Spectrophotometric assay and properties of the angiotensin I-converting enzyme of rabbit lung. *Biochem Pharmacol*, **1971**, *20*, 1637-1648.
- [10] Hernández-Ledesma, B.; Martín-Alvarez, P.J.; Pueyo, E. Assessment of the spectrophotometric method for determination of angiotensin-converting-enzyme activity: influence of the inhibition type. *J Agric Food Chem*, **2003**, *51*, 4175-4179.
- [11] Benzie, I.F.; Strain, J.J. The ferric reducing ability of plasma (FRAP) as measurement of "antioxidant power": The Frap assay. *Anal Biochem*, **1996**, *239*, 70-76.
- [12] Von Gadow, A.; Joubert, E.; Hansmann, C.F. Comparison of antioxidant activity of aspalathin with that of other plant phenols of Rooibosed tea (*Aspalathon linearis*), α -tocopherol, BHT, and BHA. *J Agric Food Chem*, **1997**, *45*, 632-638.
- [13] Re, R.; Pellegrini, N.; Proteggente, A.; Pannala, A.; Yang, M.; Rice-Evans, C. Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radic Biol Medic*, **1999**, *26(9-10)*, 1231-1237.
- [14] Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial susceptibility testing, *seventeenth informational supplement*, M100-S17, **2007**, Wayne, PA: CLSI.
- [15] Rodríguez-Vaquero, M.J.; Tomassini-Seravalle, L.R.; Manca de Nadra, M.C.; Strasser de Saad, A.M. antioxidant capacity and antibacterial activity of phenolic compounds from argentinean herbs infusions. *Food Control*, **2010**, *21*, 779-785
- [16] Li, H.B.; Wong, C.C.; Cheng, K.W.; Chen, F. Antioxidant properties in vitro and total phenolic contents in methanol extracts from medicinal plants. *Food Sci Technol*, **2008**, *41*, 385-390.
- [17] Yoo, K.M.; Lee, C.H.; Lee, H.; Moon, B.; Lee, C.Y. Relative antioxidant and cytoprotective activities of common herbs. *Food Chem*, **2008**, *106*, 929-936.
- [18] Vallejo, C.V.; Aredes Fernández, P.A.; Farias, M.E.; Rodríguez-Vaquero, M.J. biofilm inhibition of spoilage bacteria by argentinean fruit juices with antihypertensive activity. *Current Pharm Biotechnol*, **2013**, *14*, In press.
- [19] Rodríguez-Vaquero, M.J.; Manca de Nadra, M.C. Growth parameter and viability modifications of *Escherichia coli* by phenolic compounds and argentine wine extracts. *Appl Biochem Biotechnol*, **2008**, *151* (2), 342-352.

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