112

Ventilator-Induced Lung Injury (VILI) in Acute Respiratory Distress Syndrome (ARDS): Volutrauma and Molecular Effects



R. Carrasco Loza^{*,1,2}, G. Villamizar Rodríguez² and N. Medel Fernández^{1,2}

¹Laboratorio de Investigación Biomédica, Hospital del Salvador, Facultad de Medicina, Universidad de Chile, Santiago, Chile

²Unidad de Cuidados Intensivos, Clínica Dávila, Santiago, Chile

Abstract: Acute Respiratory Distress Syndrome (ARDS) is a clinical condition secondary to a variety of insults leading to a severe acute respiratory failure and high mortality in critically ill patients. Patients with ARDS generally require mechanical ventilation, which is another important factor that may increase the ALI (acute lung injury) by a series of pathophysiological mechanisms, whose common element is the initial volutrauma in the alveolar units, and forming part of an entity known clinically as ventilator-induced lung injury (VILI).

Injured lungs can be partially protected by optimal settings and ventilation modes, using low tidal volume (VT) values and high positive-end expiratory pressure (PEEP). The benefits in ARDS outcomes caused by these interventions have been confirmed by several prospective randomized controlled trials (RCTs) and are attributed to reduction in volutrauma.

The purpose of this article is to present an approach to VILI pathophysiology focused on the effects of volutrauma that lead to lung injury and the 'mechanotransduction' mechanism. A more complete understanding about the molecular effects that physical forces could have, is essential for a better assessment of existing strategies as well as the development of new therapeutic strategies to reduce the damage resulting from VILI, and thereby contribute to reducing mortality in ARDS.

Keywords: Mechanotransduction, positive-end expiratory pressure, tidal volume, VILI, volutrauma.

1. INTRODUCTION

The acute respiratory distress syndrome (ARDS) is a relatively frequent complication in critically ill patients and responsible for significant co-morbidity and mortality [1]. For many years, multiple studies have aimed to determine the mechanisms of ARDS generation and progression as well as other aspects including the recognition of subtypes of ARDS, identification of risk and prognostic factors and others, all areas that have intended to contribute to the development of new preventive and therapeutic strategies. Most of the studies show strategies to prevent the occurrence or progression of ARDS, and only a few interventions have focused on to reverse the damage or accelerate the recovery period caused by the primary injury. However and unfortunately, only a few studies have shown significant effectiveness that contribute to better clinical outcomes [1], specifically those whose targets were based on to reduce the ARDS progression through the development of protective strategies that result in a reduction of volutrauma.

The purpose of this review is to present an approach to the VILI pathophysiology mainly focused on the initial importance of the volutrauma effects in a previously healthy lung of patients afflicted with a "first hit" with risk of developing ARDS, as well as the effects of volutrauma in a lung with ARDS. A more complete understanding of these mechanisms may allow for the development of new therapeutic strategies to prevent VILI and consequently achieve better clinical outcomes.

1.1. Ventilator-Induced Lung Injury

Mechanical ventilation (MV) by itself has been established as a factor that induces lung damage, which has been demonstrated even in previously healthy lungs, where the MV could aggravate the lung effects of a 'first inflammatory hit', a concept known as ventilator-induced lung injury (VILI) [2-4]. VILI caused total lung damage, which is the result of the sum of local lung damage with a heterogeneous distribution due to heterogeneous local lung susceptibility. These local events promote an anisotropic alveolar inflation in the opening phase and collapse at the end of the expiratory phase, which promotes greater damage. This initial anisotropic distribution promotes a local inflammatory response in areas that receive the higher volutrauma, but subsequently these areas trigger a generalized inflammatory response in the lung tissue.

The pathophysiological basis of VILI are complex and characterized by different overlapping interactions [4],

^{*}Address correspondence to this author at the Laboratorio de Investigación Biomédica, Hospital del Salvador, Facultad de Medicina, Universidad de Chile, Avenida Salvador 364, Providencia, Santiago, Chile; E-mail: rcarrascoloza@gmail.com

initially after the ARDS Network clinical trial [5], VILI was primarily linked to two phenomena: the first phenomenon occurs at the end of inspiratory cycle and is associated with the use of a high inspiratory pressure of tidal volume (VT) that promotes an alveolar over-distension, a process related to the risk of rupture of the airway and alveolar walls in the non-dependent lung regions (R-ND). The second phenomenon unfolds in dependent lung regions (RD), which are exposed to significant stress when the airway and alveoli open on inspiration and collapse on expiration [tidal recruitment]. This phenomenon is associated with the use of insufficient PEEP to prevent alveolar collapse and cyclical reopening. The two phenomena may overlap, since an unnecessarily high PEEP may lead to over-distension and an excessive VT drop, it can increase the effects caused by the cyclic alveolar collapse and reopening [5]. Moreover, some experimental studies have confirmed that the collapse of the airspace and the cyclical recruitment in lungs affected by a previous inflammatory conditions are crucial in the development of VILI [4, 6].

2. MECHANISM ASSOCIATED WITH INDUCTION OF VOLUTRAUMA

2.1. Physical Forces

The volutrauma effects over the lung parenchyma can be evaluated at different spatial scales, in this sense the response of its resident cells to the deforming stress take importance in order to provide the biological basis of lung protective care.

2.1.1. Regional Mechanics in Lung

Physical key events associated to volutrauma are alveolar collapse and lung distension [7]. Alveolar collapse is determined by cyclic recruitment and derecruitment ('opening' and 'closure') that could lead to damage of the epithelial lining by interfacial forces [8]. Indeed, in injured lung regions, as end-expiratory lung volume is quite low in the recumbent posture, the alveoli and conduction airways are particularly susceptible to closure induced by liquid plugs [9].

A protective approach that limits the lung damage must maintain sufficient end-expiratory volume to prevent alveolar collapse or open a collapsed/closed lung region [7]. To open a collapsed/closed lung region required a higher pressure than the pressure at which it was closed [10], then subsequent breaths preferentially inflate the remaining open units, so that gradually a 'shrinking baby lung' could accommodate the ventilatory demand [11]. The lung injury effects caused by the collapse and closing of alveolar could be reduced when lung areas may be permanently open [12, 13].

Mechanisms of biophysical injury associated with alveolar overdistension are responsible for lung injury through several effects including a decline in the surface tension due to reduction in surfactant properties [14], an increase of filtration coefficient and protein permeability in the endothelial and epithelial barriers [15], as well as wounding of alveolar resident cells by tensile and interfacial stress [8, 16]. In conclusion, a protective approach, must to reach an end-expiratory volume with a balance between preventing alveolar collapse and limit lung distension.

2.1.2. Mechanics of a Single Alveolus

At the alveolar level, the tidal stretches inflate the alveolus, therefore cells that lining the alveolus and their elastic linkages (the e-cadherins), become stretched. From physical perspective, the epithelial cells and their linkages through e-cadherin could be considered as springs in series. The capacity of the individual extension of these components depends on their relative stiffness, therefore if cells are too stiff, the intercellular linkages will drive the deformation during the inflation of the alveolus [7]. The stiffness of the epithelial cells is mostly determined by the actin cytoskeleton [17], whereas the total stiffness of e-cadherin linkages depends on their density around the cells being in parallel with each other's [7]. Therefore, any pathology that affects an adequate metabolic functioning of epithelial cells could result in inadequate distribution of the cytoskeleton, which in turn lead to increased cellular stiffness, determine a higher stress on the intercellular junctions and alteration of the alveolar permeability.

2.2. Lung Stress and Strain

Unfortunately the values of lung deformation (or strain) that can be safely reached during mechanical ventilation are poorly defined, especially in humans [18]. In the clinical practice usually the lung distension is commonly inferred from VT (mL/kg of ideal body weight) and end-inspiratory (plateau) airway pressure [5]. However, neither of these two variables reliably reflects tissue deformation, especially during ALI (acute lung injured)[18], when relationships between body weight and functional residual capacity (FRC) and between airway and transpulmonary pressure become unpredictable [19]. From the physical view, strain and stress are used to describe the responses of a body to a external overload; strain is the relative change in size and shape and stress is the internal tension [20]. The exact measurement of lung strain and stress is infeasible in clinical practice, but it is possible make some approximate measurements of these.

Lung strain is the ratio between the volume of gas inflated (change in volume) and FRC [18]. On the other hand, lung stress equals the corresponding change in transpulmonary (airway minus pleural) pressure [21]. However, as the lung stress-strain relationship is almost linear until reach the total capacity [19], lung strain can be derived from stress and viceversa. In this fact, the lung strain can be computed from FRC, which several ventilators automatically measure it, whereas over-imposed lung stress can be estimated from changes in the esophageal pressure [19]. In the assessed of the strain effects, we must consider different types of strain: static, dynamic and global strain. For example, during mechanical ventilation without PEEP, lung deformation is due to tidal ventilation, so this dynamic strain can be computed as VT/FRC. Moreover, when PEEP (and the corresponding volume of gas, VPEEP) is applied, lungs are also kept tonically inflated above their FRC and are exposed to an additional static strain (VPEEP/FRC). Finally the global strain is the sum of these two components: global strain = (VT + VPEEP)/FRC.

2.2.1. Dynamic Versus Static Strain: Animal Studies

Most of the studies have shown that ventilation with large VT and zero end-expiratory pressure is more injurious than ventilation with small tidal volume and some PEEP, for a given peak pressure [22]. Some studies have shown that medium-sized animals ventilated with dynamic strain of 2.5 (static strain equal to 0), cause lung damage and develop fatal pulmonary edema. By contrast, those animals ventilated with smallest dynamic (0.5) and largest static (2.0) strain ended the experiment with normal lungs, although they needed higher amount of fluids and high dose of norepinephrine to maintain an adequate cardiac output [23]. In these medium-sized animals, harms of mechanical ventilation depend not only on overall lung inflation, but also depend of the form that this is achieved: large cyclic deformations are more injurious than equivalent, mainly static.

On the other hand, other studies in small animals showed apparently contradictory results [24], because healthy rats developed pulmonary edema when they were ventilated with small VT(7 ml/kg of body weight) and high PEEP (15 cm H_2O). However, a methodological point of this latest study to consider was that lungs were inflated above their upper physiological limit [25]. These different results can be explained, being that the end-inspiratory volume does not exceed the total capacity, healthy lungs better tolerate static than dynamic, inflations [18]. However, when endinspiratory lung volume exceeds the total capacity, static or dynamic inflation invariably causes lung damage. This findings support the measuring of lung volumes during mechanical ventilation [26].

2.2.2. Dynamic Versus Static Strain: Human Studies

The major survival benefits evidence by clinical research has been represented by limiting the VT and by the increasing PEEP during acute respiratory insufficiency [18]. In patients with acute respiratory failure, the limitation of VT is beneficial either *per se* [5] or in conjunction with increasing PEEP [27, 28]. By contrast, the only increase in PEEP generally does not diminish hospital mortality [3, 29, 30], unless that the VT is concomitantly reduced [27, 28]. Moreover, in patients with healthy lungs, although few clinical studies have been conducted, the conclusions are very similar [18].

First, one meta-analysis suggests that MV with smaller VT and higher PEEPs is associated with a lower incidence of pulmonary complications in patients without ALI at enrolment [31]. These findings have been confirmed for two subsequently RCTs [32, 33].

Second, in patients with non-pulmonary disease was also found that increasing the PEEP without changing the tidal volume did not diminish the incidence of ARDS in mechanically ventilated [34].

Lung damage during ventilation mainly depends on the amplitude and the velocity of dynamic tissue deformation according to suggested by experimental and clinical evidence and could be explained by tissue viscoelasticity. However, if heat produced by the internal frictions can change the conformation of molecules forming lung parenchyma, which is rich in water, is uncertain and needs to be validated in the future [18]. For the previously mentioned reasons, it has tried to develop strategies to minimize tidal ventilation, including 'apnoea' with extracorporeal carbon dioxide removal [35].

Animal studies [36] and the failure in high-frequency jet ventilation (at low PEEP)[37] suggest that a high static inflation (up to total lung capacity) may be required to maintain the lung open. This effect is due to that during apnea, bronchial secretions are not efficiently cleared, distal airways get obstructed and re-absorption atelectasis leads to progressive alveolar de-recruitment [18]. However, in two clinical trials in which tried to evaluate this idea using highfrequency oscillatory ventilation, these did not report benefit in outcome patients with ARDS [38, 39].

These findings could be explained by a right heart dysfunction because of higher mean airway pressure (close to or even above 30 cm H_2O), with higher need for fluids and vasoactive drugs in the study population [40]. Therefore, it seems that interventions designed to protect the lung will hardly improve survival if they concomitantly harm the heart [18].

2.3. Influence of Ventilation Pattern

Beneficial effects have been found in animal models with an increased PEEP and a decreased VT by a reduction in the dynamic component of lung stretch [23]. These event is consistent with cellular studies suggesting that large stretch amplitudes and/or high strain rates are injurious to epithelial [41] and endothelial cells [42].

The risk of developing lung injury could be reduced with higher PEEP, which could open lung regions and make more homogeneous the stiffness at the length scales of small airways and alveoli [43]. In the intracellular level, structures could tolerate the static stretches of the PEEP by proper cytoskeletal remodeling, but are unable to adjust to the fast dynamically changing large strains [7]. Other factors that determine the effect of ventilation patterns in patients are high flow rates [44] and frequent deep inspirations [45], which can also worsen the ventilatory outcomes. In this regard, deep inspirations could help with recruitment and survival compared with conventional ventilation at 12 mL/kg [46], they could also have negative consequences [7]. The reason for these events could be since the integrity of the epithelium is compromised by deep inspirations resulting in increased e-cadherin levels in the lavage fluid [47]. Additionally, in lungs with edema, deep inspirations also caused cell apoptosis and alveolar capillary injury [48]. However, the mechanism that supports these findings are unclear, but they could also be related to the injurious effects of maximum strain and/or high strain rate on cells and intercellular junctions in the presence of high interfacial stresses [7].

In case of edema, deep inspirations also caused cell apoptosis and alveolar capillary injury [48]. The mechanisms behind these findings are unclear, but we speculate that they may also be related to the injurious effects of maximum strain and/or high strain rate on cells and intercellular junctions in the presence of high interfacial stresses. Novel ventilation strategies have emerged trying to reduce the effects already described that lead to lung injury [49]. In the variable ventilation, VT changes are from cycle to cycle [49], and it has shown those not only improves lung function [12, 50-53], but also reduce the epithelial injury with a reduction of e-cadherin levels in lavage fluid [47]. These events could be explained by the ability of this ventilatory modes to maintain a better alveolar recruitment than conventional ventilation [54], and an important mechanism could be due to the variable stretching of epithelial cells that directly stimulates surfactant secretion [55], which should have a protective effect against edema formation but this need to be confirmed experimentally [7].

2.3.1. Ventilation at High Lung Volumes

High (absolute) lung volumes may cause lung damage by several mechanisms such as alveolar rupture, air leaks, and gross barotrauma (e.g., pneumothorax, pneumomediastinum, subcutaneous emphysema)[56]. The critical variable leading to the air leaks is regional lung overdistension, not high airway pressure *per se*, so the term of barotrauma can be misleading [57].

Lung overdistension may also cause more subtle forms of damage manifested by pulmonary edema [57]. The overdistension has been demonstrated in animal models, where ventilated rats with very high peak airway pressures (and therefore overdistention) and zero PEEP developed hypoxemia and postmortem examination revealed perivascular and alveolar edema [58]. Conversely, in rats that underwent ventilation with the same peak airway pressure but with the addition of a PEEP of 10 cm of water did not develop edema, which showing an interaction between overdistension and low end-expiratory lung volume with respect to lung injury [58]. However, the precise mechanisms underlying this interaction have not been completely elucidated [57].

Other animal studies have shown that the volume would have a major importance than airway pressure in the development of lung injury [57]. In this view, animals undergoing ventilation with high VT developed pulmonary edema while animals undergoing ventilation with similar airway pressures but with straps around their abdomens and chests that reduced the tidal volumes, did not developed pulmonary edema [59]. Experimental approaches showed that the volume (i.e., lung stretching), not

Airway pressure, was the most important factor in determining injury, a finding that led them to coin the term "volutrauma" [57].

2.3.2. Ventilation at Low Lung Volumes

Ventilation at low (absolute) lung volumes could also cause injury through multiple mechanisms, such as repetitive opening and closing of airways and lung units [60, 61], effects on surfactant function [62] and regional hypoxia. In this case, epithelial sloughing, hyaline membranes and pulmonary edema characterize lung injury caused, and it has been termed "atelectrauma" [60].

Moreover, the atelectrauma is amplified in lungs, in which there are marked homogeneities in ventilation [57]. Since many decades ago, it was shown that the stretching forces in lung parenchyma at margins between aerated and atelectatic regions could be up to four to five times as high as those in other lung regions [63].

3. MECHANISM OF VENTILATOR-INDUCED LUNG INJURY IN HEALTHY AND ARDS LUNGS

3.1. General Concept

The pathophysiological principles of VILI are complex and characterized by different overlapping interactions of multiple factors, some of which were previously described. These interactions include [4]: (i) high VT causing over distension; (ii) cyclic closing and opening of peripheral airways during tidal breath resulting in damage of both the bronchiolar epithelium and the parenchyma (lung strain), mainly at the alveolar-bronchiolar junctions; (iii) lung stress by increased transpulmonary pressure); (iv) low lung volume associated with recruitment and de-recruitment of unstable lung units (atelectrauma);(v) inactivation of surfactant by large alveolar surface area oscillations associated with surfactant aggregate conversion, which increases surface tension [64]; (vi) local and systemic release of lung-borne inflammatory mediators, namely biotrauma [65].

3.2. New Pathophysiological Concepts

Some experimental and clinical studies have demonstrated two main mechanisms leading to VILI: direct trauma to the cell, which promotes releasing of cytokines to the alveolar space as well as to the systemic circulation and also, the so-called 'mechanotransduction' mechanism.

3.2.1. Direct Volutrauma

The direct effect caused by volutrauma result in a persistent cyclic stretch during mechanical ventilation, which stimulates alveolar epithelial and vascular endothelial cells through mechano-sensitive membrane-associated protein and ion channels [66]. This cyclic recruitment of unstable overdistended alveolar regions provoked by MV causes initially a local inflammatory response, that is associated with the cellular necrosis surrounding lung tissue [66], and then a systemic inflammatory response which ultimately can lead to severe MODS (multiple organ dysfunction syndrome) [67].

The tissue deformation secondary to volutrauma activates nuclear factor-kappa B (NF- κ B) signaling, which consequent leads to the production of interleukin (IL)-6, IL-8, IL-1 β and tumor necrosis factor [TNF]- α [66]. For example, high IL-6 levels have been detected in various experimental models of VILI [68], and in the serum and bronchoalveolar lavage fluid of patients with ARDS under MV [67]. Thus, cytokines and chemokines are potential effector molecules that modulate and regulate VILI initial mechanisms. In this view, the effects of high VT ventilation even have been demonstrated in models without previous lung injury, where for example a

high VT ventilation led to an increase in expression of intrapulmonary TNF- α and macrophage inflammatory protein-2 in mice [69].

3.2.2. Mechanotransduction

The term mechanotransduction is referred to the conversion of mechanical stimuli to a biochemical response when alveolar epithelium or vascular endothelium is stretched during mechanical ventilation [4]. Mechanical stimulus causes expansion of the plasma membrane and triggers cellular signaling thorugh various inflammatory mediators that influence a pulmonary and systemic cell dysfunction [66]. A high level of mechanical stretch is associated with increased epithelial cell necrosis, decreased apoptosis and increased of IL-8 levels [66]. Moreover, extracellular matrix (ECM), that determines the biomechanical behavior of the lung and plays a role in stabilizing lung matrix and fluid content, could also be affected [4]. This is because the mechanotransduction causes a mechanical force on ECM, which in turn causes a lung strain (the ratio between VT and functional residual capacity) [4]. Thus, high-VT ventilation causes ECM remodeling, influenced by the airway pressure gradient and the pleural pressure gradient [65, 70].

It has been in animal models that pulmonary edema formation in VILI develops when lung strain is greater than 1.5–2-fold [71]. Cyclic mechanical stress causes release and activation of matrix metalloproteinase (MMP), which play an important role in regulating ECM remodeling and VILI [4]. Moreover, lung strain could also lead to a modification of proteoglycan and glycosaminoglycans [4], which could cause different effects. For example, fragmentation of glycosaminoglycans may affect the development of the inflammatory response by interacting with various types of chemokine and acting as ligands for Toll-like receptors [70, 72].

Finally, mechanical strain induces ECM breakdown [70], which has demonstrated to be a signal for matrikines [4]. The relationship between pathophysiological events presented in VILI and mechanotransduction are shown in Fig. (1).

4. VENTILATION STRATEGIES AND MOLECULAR EFFECTS

Multiple ventilation strategies have been evaluated with the purpose to reduce lung injury, for example: low tidal volumes to limit overdistention, higher PEEPs to prevent injury at low lung volumes (atelectrauma), and recruitment maneuvers (procedures used to reinflate collapsed lung units) through the sustained application of an airway pressure higher than approximately 35 cm of water, which if achieved inflating atelectatic regions, than minimizes the ventilation heterogeneity [57]. The clinical evidence of each of these ventilation strategies are briefly described below.

4.1. Low Tidal Volumes

Usually, ARDS patients have two types of regions: relatively normally aerated non-dependent lung regions and relatively non-aerated dependent lung regions, and this last case refers to regions that are lower from a gravitational perspective than other regions and therefore are more likely to be collapsed [57]. For this reason, the volume available for ventilation is smaller, so that this condition has received the term of "baby lung" [11]. Therefore a decreased VT, emerges as a solution to ventilate this "baby lung" and prevent overinflation of the relatively small, normally aerated regions [57].

This strategy was initially assessed by clinical studies [27, 73], and their results provided the basis for the further development of ARDS Network, the main study that evaluated the use of low tidal volumes [5]. The ARDS Network investigators compared a control strategy using a tidal volume of 12 mL/Kg of predicted body weight with respect to a low-tidal-volume strategy that used 6 mL/ kg of predicted body weight. The low-tidal-volume strategy was associated with an absolute reduction of rate of death [39.8% *vs* 31.0%] [5].

4.2. High PEEP and Recruitment Maneuvers

Several forms of respiratory failure are characterized by pulmonary edema and end-expiratory alveolar collapse, thus the prevention of a potential atelectrauma is essential for reducing lung injury. In this view, a low PEEP may be insufficient to stabilize alveoli and keep them open, thereby increasing the probability VILI by atelectrauma [57]. A meta-analysis of randomized clinical trials in ARDS patients [74], concluded that a higher PEEP was associated with an absolute reduction of 5 percent in death rate among patients which had worse oxygenation, defined as a ratio of the partial pressure of arterial oxygen to the fraction of inspired oxygen $[PaO_2:FiO_2] \le 200 \text{ mm Hg}$ [74]. However, we should note that it is essential to find a value of PEEP not excessively high, because conversely, a too high PEEP could potentially cause adverse effects, including impairment of venous return and pulmonary overdistention [57]. For this reason, it is essential to develop a method to determine the most appropriate PEEP for each patient. Thereby, due to the importance of transpulmonary pressure in lung injury, an obvious approach would be to use transpulmonary pressure to set the PEEP, with the use of esophageal pressure as a surrogate for pleural pressure [57].

However, in this case is difficult obtain an adequate interpretation of absolute esophageal pressure for various reasons, including cardiac artifacts, the uneven distribution of pleural pressure, and esophageal distortion and contraction (especially in supine patients) [75]. However, a pilot study in ARDS patients evaluated this approach [76], setting the PEEP to achieve an end-expiratory transpulmonary pressure of 0 to 10 cm of water, while limiting end-inspiratory transpulmonary pressure to 25 cm of water. Thus, they found an improved oxygenation and a trend toward lower 28-day mortality [76]. However, it is necessary a larger clinical trial that shows significant clinical results, before this approach can be recommended in clinical practice. Finally, the recruitment maneuvers, that are based on the ability to achieve inflate atelectatic regions ventilation heterogeneity, minimizing the should theoretically reduce the lung injury [77]. Although some clinical trials that used such maneuvers have been included in a meta-analysis [74], and have been implemented as a



Fig. (1). Mechanisms associated with VILI development.

protective strategy increasing the number of lungs retrieved from heart-beating donors [78]. The role of recruitment maneuvers in clinical practice remains uncertain because of questions about its effect on outcomes and concerns regarding complications such as hemodynamic compromise or pneumothorax [3].

4.3. High-Frequency Oscillatory Ventilation

High-frequency oscillatory ventilation [HFOV] is a strategy characterized by use very small tidal volumes, applied at high frequencies (up to 15 per second) [57], which should theoretically be ideal to minimizing ventilatorinduced lung injury [79]. A meta-analysis which included eight clinical trials involving a total of 419 adults with ARDS, was found that HFOV-treated patients had significantly lower mortality than patients treated with conventional ventilation, which suggested that HFOV might improve survival and is unlikely to cause damage [80]. However, due to two recent large multicenter trials in ARDS patients did not show improved outcomes with HFOV [38, 39], this ventilatory strategy cannot be recommended as first-line therapy in such patients [81].

CONCLUSION

The acute respiratory distress syndrome (ARDS) persists as an important health problem in intensive care units, responsible of a significant co-morbidity and mortality [1]. The pathophysiology of VILI has initial physical mechanisms, which subsequently through the mechanotransduction lead to a molecular damage response. The physical mechanisms of injury include two main phenomena which may be overlapping, the over-distension in case of an unnecessarily high PEEP (volutrauma) and atelectrauma caused by the cyclic alveolar collapse and reopening in patients with excessive VT reduction (Fig. 1).

The mechanotransduction signals, lead to a series of events including development of inflammation, necrosis, apoptosis and remodeling of the extracellular matrix. Several ventilatory strategies have been evaluated to prevent lung injury, but so far only the reduction of VT to prevent volutrauma and an adequate PEEP to prevent atelectrauma have mostly been validated for use in routine clinical practice. Other potential ventilatory strategies need to be evaluated by larger clinical studies before extending its use in clinical practice.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

ACKNOWLEDGEMENTS

Author's Role: All authors helped to write the manuscript and have seen and approved the final version.

REFERENCES

- Koh Y. Update in acute respiratory distress syndrome. J Intensive Care 2014; 2: 2.
- [2] Bove PF, van der Vliet A. Nitric oxide and reactive nitrogen species in airway epithelial signaling and inflammation. Free Radic Biol Med 2006; 41: 515-27.
- [3] Brower RG, Lanken PN, MacIntyre N, et al. Higher versus lower positive end-expiratory pressures in patients with the acute respiratorydistress syndrome. N Engl J Med 2004; 351: 327-36.
- [4] Sutherasan Y, Vargas M, Pelosi P. Protective mechanical ventilation in the non-injured lung: review and meta-analysis. Crit Care 2014; 18: 211.
- [5] ARDSnetwork1. Ventilation with lower tidal volumes as compared with traditional tidal volumes for acute lung injury and the acute respiratory distress syndrome: the Acute Respiratory Distress Syndrome Network. N Engl J Med 2000; 342: 1301-8.
- [6] Sinclair SE, Chi E, Lin HI, Altemeier WA. Positive end-expiratory pressure alters the severity and spatial heterogeneity of ventilatorinduced lung injury: an argument for cyclical airway collapse. J Crit Care 2009; 24: 206-11.
- [7] Suki B, Hubmayr R. Epithelial and endothelial damage induced by mechanical ventilation modes. Curr Opin Crit Care 2014; 20: 17-24.
- [8] Hussein O, Walters BJ, Stroetz RW, et al. Biophysical determinants of alveolar epithelial plasma membrane wounding associated with mechanical ventilation. Am J Physiol 2013; 305: L478-84.
- [9] Washko GR, O'Donnell CR, Loring SH. Volume-related and volumeindependent effects of posture on esophageal and transpulmonary pressures in healthy subjects. J Appl Physiol 2006; 100: 753-8.
- [10] Otis DR Jr, Petak F, Hantos Z, et al. Airway closure and reopening assessed by the alveolar capsule oscillation technique. J Appl Physiol 1996; 80: 2077-84.
- [11] Gattinoni L, Pesenti A. The concept of 'baby lung'. Intensive Care Med 2005; 31: 776-84.
- [12] Carvalho AR, Pacheco SA, de Souza Rocha PV, et al. Detection of tidal recruitment/overdistension in lung-healthy mechanically ventilated patients under general anesthesia. Anesth Analg 2013; 116: 677-84.
- [13] Zannin E, Dellaca RL, Kostic P, et al. Optimizing positive endexpiratory pressure by oscillatory mechanics minimizes tidal recruitment and distension: an experimental study in a lavage model of lung injury. Crit Care 2012; 16: R217.
- [14] Veldhuizen RA, Welk B, Harbottle R, *et al.* Mechanical ventilation of isolated rat lungs changes the structure and biophysical properties of surfactant. J Appl Physiol 2002; 92: 1169-75.
- [15] Wray C, Mao Y, Pan J, Chandrasena A, Piasta F, Frank JA. Claudin-4 augments alveolar epithelial barrier function and is induced in acute lung injury. Am J Physiol Lung Cell Mol Physiol 2009; 297: L219-27.
- [16] Pecchiari M, Monaco A, Koutsoukou A, D'Angelo E. Plasma membrane disruptions with different modes of injurious mechanical ventilation in normal rat lungs. Crit Care Med 2012; 40: 869-75.
- [17] Van Citters KM, Hoffman BD, Massiera G, Crocker JC. The role of factin and myosin in epithelial cell rheology. Biophys J 2006; 91: 3946-56.

- [18] Protti A, Votta E, Gattinoni L. Which is the most important strain in the pathogenesis of ventilator-induced lung injury: dynamic or static? Curr Opin Crit Care 2014; 20: 33-8.
- [19] Chiumello D, Carlesso E, Cadringher P, et al. Lung stress and strain during mechanical ventilation for acute respiratory distress syndrome. Am J Respir Crit Care Med 2008; 178: 346-55.
- [20] Pilkey WD. Formulas for stress, strain and structural matrices. 2nd ed. John Wiley & Sons: Hoboken, New Jersey 2005; 89-143.
- [21] Gattinoni L, Carlesso E, Cadringher P, Valenza F, Vagginelli F, Chiumello D. Physical and biological triggers of ventilator-induced lung injury and its prevention. Eur Respir J Suppl 2003; 47: 15s-25s.
- [22] Dreyfuss D, Saumon G. Ventilator-induced lung injury: lessons from experimental studies. Am J Respir Crit Care Med 1998; 157: 294-323.
- [23] Protti A, Andreis DT, Monti M, et al. Lung stress and strain during mechanical ventilation: any difference between statics and dynamics? Crit Care Med 2013; 41: 1046-55.
- [24] Dreyfuss D, Saumon G. Role of tidal volume, FRC, and end-inspiratory volume in the development of pulmonary edema following mechanical ventilation. Am Rev Respir Dis 1993; 148: 1194-203.
- [25] Gattinoni L, Protti A, Caironi P, Carlesso E. Ventilator-induced lung injury: the anatomical and physiological framework. Crit Care Med 2010; 38: S539-48.
- [26] Dellamonica J, Lerolle N, Sargentini C, et al. PEEP-induced changes in lung volume in acute respiratory distress syndrome: two methods to estimate alveolar recruitment. Intensive Care Med 2011; 37: 1595-604.
- [27] Amato MB, Barbas CS, Medeiros DM, et al. Effect of a protectiveventilation strategy on mortality in the acute respiratory distress syndrome. N Engl J Med 1998; 338: 347-54.
- [28] Villar J, Kacmarek RM, Pérez-Méndez L, Aguirre-Jaime A. A high positive end-expiratory pressure, low tidal volume ventilatory strategy improves outcome in persistent acute respiratory distress syndrome: a randomized, controlled trial. Crit Care Med 2006; 34: 1311-8.
- [29] Meade MO, Cook DJ, Guyatt GH, et al. Ventilation strategy using low tidal volumes, recruitment maneuvers, and high positive end-expiratory pressure for acute lung injury and acute respiratory distress syndrome: a randomized controlled trial. JAMA 2008; 299: 637-45.
- [30] Mercat A, Richard JC, Vielle B, et al. Positive end-expiratory pressure setting in adults with acute lung injury and acute respiratory distress syndrome: a randomized controlled trial. JAMA 2008; 299: 646-55.
- [31] Serpa Neto A, Cardoso SO, Manetta JA, et al. Association between use of lung-protective ventilation with lower tidal volumes and clinical outcomes among patients without acute respiratory distress syndrome: a meta-analysis. JAMA 2012; 308: 1651-9.
- [32] Severgnini P, Selmo G, Lanza C, et al. Protective mechanical ventilation during general anesthesia for open abdominal surgery improves post-operative pulmonary function. Anesthesiology 2013; 118: 1307-21.
- [33] Futier E, Constantin JM, Paugam-Burtz C, et al. A trial of intraoperative low-tidal-volume ventilation in abdominal surgery. N Engl J Med 2013; 369: 428-37.
- [34] Pepe PE, Hudson LD, Carrico CJ. Early application of positive endexpiratory pressure in patients at risk for the adult respiratory-distress syndrome. N Engl J Med 1984; 311: 281-6.
- [35] Gattinoni L, Pesenti A, Kolobow T, Damia G. A new look at therapy of the adult respiratory distress syndrome: motionless lungs. Int Anesthesiol Clin 1983; 21: 97-117.
- [36] Gattinoni L, Iapichino G, Kolobow T. Hemodynamic, mechanical and renal effects during 'apneic oxygenation' with extracorporeal carbon dioxide removal, at different levels of intrapulmonary pressure in lambs. Int J Artif Organs 1979; 2: 249-53.
- [37] Carlon GC, Howland WS, Ray C, *et al.* High-frequency jet ventilation: a prospective randomized evaluation. Chest 1983; 84: 551-9.
- [38] Ferguson ND, Cook DJ, Guyatt GH, et al. High-frequency oscillation in early acute respiratory distress syndrome. N Engl J Med 2013; 368: 795-805.
- [39] Young D, Lamb SE, Shah S, et al. High-frequency oscillation for acute respiratory distress syndrome. N Engl J Med 2013; 368: 806-13.
- [40] Repesse' X, Charron C, Vieillard-Baron A. Right ventricular failure in acute lung injury and acute respiratory distress syndrome. Minerva Anestesiol 2012; 78: 941-8.
- [41] Tschumperlin DJ, Oswari J, Margulies AS. Deformation-induced injury of alveolar epithelial cells. Effect of frequency, duration, and amplitude. Am J Respir Crit Care Med 2000; 162: 357-62.
- [42] Birukova AA, Fu P, Xing J, et al. Mechanotransduction by gef-h1 as a novel mechanism of ventilator-induced vascular endothelial

permeability. Am J Physiol Lung Cell Mol Physiol 2010; 298: L837-L848.

- [43] Wellman TJ, Winkler T, Costa EL, *et al.* Effect of regional lung inflation on ventilation heterogeneity at different length scales during mechanical ventilation of normal sheep lungs. J Appl Physiol 2012; 113: 947-57.
- [44] Garcia CS, Abreu SC, Soares RM, et al. Pulmonary morphofunctional effects of mechanical ventilation with high inspiratory air flow. Crit Care Med 2008; 36: 232-9.
- [45] Steimback PW, Oliveira GP, Rzezinski AF, et al. Effects of frequency and inspiratory plateau pressure during recruitment manoeuvres on lung and distal organs in acute lung injury. Intensive Care Med 2009; 35: 1120-8.
- [46] Barbas CS, de Matos GF, Okamoto V, et al. Lung recruitment maneuvers in acute respiratory distress syndrome. Respir Care Clin N Am 2003; 9: 401-18.
- [47] Thammanomai A, Hamakawa H, Bartolak-Suki E, Suki B. Combined effects of ventilation mode and positive end-expiratory pressure on mechanics, gas exchange and the epithelium in mice with acute lung injury. PLoS One 2013; 8: e53934.
- [48] Santiago VR, Rzezinski AF, Nardelli LM, et al. Recruitment maneuver in experimental acute lung injury: the role of alveolar collapse and edema. Crit Care Med 2010; 38: 2207-14.
- [49] Lefevre GR, Kowalski SE, Girling LG, et al. Improved arterial oxygenation after oleic acid lung injury in the pig using a computercontrolled mechanical ventilator. Am J Respir Crit Care Med 1996; 154: 1567-72.
- [50] Arold SP, Mora R, Lutchen KR, et al. Variable tidal volume ventilation improves lung mechanics and gas exchange in a rodent model of acute lung injury. Am J Respir Crit Care Med 2002; 165: 366-71.
- [51] Arold SP, Suki B, Alencar AM, et al. Variable ventilation induces endogenous surfactant release in normal guinea pigs. Am J Physiol Lung Cell Mol Physiol 2003; 285: L370-5.
- [52] Funk DJ, Graham MR, Girling LG, et al. A comparison of biologically variable ventilation to recruitment manoeuvres in a porcine model of acute lung injury. Respir Res 2004; 22: 5.
- [53] Spieth PM, Carvalho AR, Pelosi P, et al. Variable tidal volumes improve lung protective ventilation strategies in experimental lung injury. Am J Respir Crit Care Med 2009; 179: 684-93.
- [54] Suki B, Alencar AM, Sujeer MK, et al. Life-support system benefits from noise. Nature 1998; 393: 127-28.
- [55] Arold SP, Bartolak-Suki E, Suki B. Variable stretch pattern enhances surfactant secretion in alveolar type ii cells in culture. Am J Physiol Lung Cell Mol Physiol 2009; 296: L574-81.
- [56] Macklin MT, Macklin CC. Malignant interstitial emphysema of the lungs and mediastinum as an important occult complication in many respiratory diseases and other conditions: an interpretation of the clinical literature in the light of laboratory experiment. Medicine 1944; 23: 281-358.
- [57] Slutsky AS, Ranieri VM. Ventilator-induced lung injury. N Engl J Med 2013; 369: 2126-36.
- [58] Webb HH, Tierney DF. Experimental pulmonary edema due to intermittent positive pressure ventilation with high inflation pressures: protection by positive end-expiratory pressure. Am Rev Respir Dis 1974; 110: 556-65.
- [59] Dreyfuss D, Soler P, Basset G, Saumon G. High inflation pressure pulmonary edema: respective effects of high airway pressure, high tidal volume, and positive end-expiratory pressure. Am Rev Respir Dis 1988; 137: 1159-64.
- [60] Slutsky AS. Lung injury caused by mechanical ventilation. Chest 1999; 116: 9S-15S.

Received: March 14, 2015

Revised: April 16, 2015

Accepted: April 16, 2015

© Carrasco Loza et al.; Licensee Bentham Open.

This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/) which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.

- [61] Bilek AM, Dee KC, Gaver DP III. Mechanisms of surface-tensioninduced epithelial cell damage in a model of pulmonary airway reopening. J Appl Physiol 2003; 94: 770-83.
- [62] Albert RK. The role of ventilation-induced surfactant dysfunction and atelectasis in causing acute respiratory distress syndrome. Am J Respir Crit Care Med 2012; 185: 702-8.
- [63] Mead J, Takishima T, Leith D. Stress distribution in lungs: a model of pulmonary elasticity. J Appl Physiol 1970; 28: 596-608.
- [64] De Prost N, Dreyfuss D: How to prevent ventilator-induced lung injury? Minerva Anestesiol 2012; 78:1054-66.
- [65] Pelosi P, Negrini D. Extracellular matrix and mechanical ventilation in healthy lungs: back to baro/volotrauma? Curr Opin Crit Care 2008; 14: 16-21.
- [66] Lionetti V, Recchia FA, Ranieri VM. Overview of ventilator-induced lung injury mechanisms. Curr Opin Crit Care 2005; 11: 82-6.
- [67] Ranieri VM, Suter PM, Tortorella C, et al. Effect of mechanical ventilation on inflammatory mediators in patients with acute respiratory distress syndrome: a randomized controlled trial. JAMA 1999; 282: 54-61.
- [68] Steinberg JM, Schiller HJ, Halter JM, et al. Alveolar instability causes early ventilator induced lung injury independent of neutrophils. Am J Respir Crit Care Med 2004; 169: 57-63.
- [69] Wilson MR, Choudhury S, Goddard ME, O'Dea KP, Nicholson AG, Takata M. High tidal volume upregulates intrapulmonary cytokines in an *in vivo* mouse model of ventilator-induced lung injury. J Appl Physiol 2003; 95: 1385-93.
- [70] Pelosi P, Rocco PR. Effects of mechanical ventilation on the extracellular matrix. Intensive Care Med 2008; 34: 631-9.
- [71] Protti A, Cressoni M, Santini A, et al. Lung stress and strain during mechanical ventilation: any safe threshold? Am J Respir Crit Care Med 2011; 183: 1354-62.
- [72] Souza-Fernandes AB, Pelosi P, Rocco PR. Bench-to-bedside review: the role of glycosaminoglycans in respiratory disease. Crit Care 2006; 10: 237.
- [73] Hickling KG, Henderson SJ, Jackson R. Low mortality associated with low volume pressure limited ventilation with permissive hypercapnia in severe adult respiratory distress syndrome. Intensive Care Med 1990; 16: 372-7.
- [74] Briel M, Meade M, Mercat A, et al. Higher vs lower positive endexpiratory pressure in patients with acute lung injury and acute respiratory distress syndrome: systematic review and meta-analysis. JAMA 2010; 303: 865-73.
- [75] Zin WA, Milic-Emili J. In: Esophageal pressure measurement. Hamid Q, Shannon J, Martin J, Eds. Physiologic basis of respiratory disease. Hamilton, ON, Canada: BC Decker 2005; pp. 639-47.
- [76] Talmor D, Sarge T, Malhotra A, et al. Mechanical ventilation guided by esophageal pressure in acute lung injury. N Engl J Med 2008; 359: 2095-104.
- [77] Lachmann B. Open up the lung and keep the lung open. Intensive Care Med 1992; 18: 319-21.
- [78] Mascia L, Pasero D, Slutsky AS, et al. Effect of a lung protective strategy for organ donors on eligibility and availability of lungs for transplantation: a randomized controlled trial. JAMA 2010; 304: 2620-7
- [79] Ferguson ND, Slutsky AS. High-frequency ventilation is the optimalphysiological approach to ventilate ARDS patients. J Appl Physiol 2008; 104: 1230-1.
- [80] Sud S, Sud M, Friedrich JO, *et al.* High frequency oscillation in patients with acute lung injury and acute respiratory distress syndrome [ARDS]: systematic review and meta-analysis. BMJ 2010; 340: c2327.
- [81] Malhotra A, Drazen JM. High-frequency oscillatory ventilation on shaky ground. N Engl J Med 2013; 368: 863-5.