

CD4 T Cells Treated with gp120 Acquire a CD45RO+/CD45RA+ Phenotype

Sergey A. Trushin^{1,2}, Gary D. Bren¹, Andrew D. Badley^{*,1,2}

¹Division of Infectious Diseases, ²Program in Translational Immunovirology and Biodefense, Mayo Clinic, Rochester, MN 55905, USA

Abstract: HIV-infected patients exhibit quantitative and qualitative defects in CD4 T cells, including having increased numbers of CD4+CD45RO+/CD45RA+ T cells, although it remains unclear how these cells arise. Here we demonstrate that gp120 treatment of activated but not resting primary human CD4 T cells decreases number of cells with single positive CD45RO+/CD45RA- effector memory phenotype while proportionally increasing the subset of cells with double positive CD45RO+/CD45RA+ mixed phenotype. We found that double positive CD45RO+/CD45RA+CD4 T cells preferentially undergo apoptosis while single positive CD45RO+/CD45RA- and CD45RO-/CD45RA+ do not. Blocking gp120-CD4 interaction with sCD4 or inhibition Lck activity reverses gp120 induced increase in double positive CD45RO+/CD45RA+CD4 T cells and subsequently diminishes the apoptosis of double positive CD45RO+/CD45RA+ cells. Altogether these data indicate that gp120 ligation of the CD4 receptor increases the number of double positive CD45RO+/CD45RA+ CD4 T cells which subsequently undergo apoptosis in a CD4 dependent manner.

INTRODUCTION

One hallmark of HIV pathogenesis is a decline of CD4 T cell number that results from the death of both HIV-1 infected CD4 T cells, as well as uninfected cells. Both HIV expressed proteins, as well as immune activation, contribute to this T cell death [1]. Immune activation leads to an increase in T cell turnover from enhanced proliferation which in turn is associated with high rates of apoptosis [2, 3]. The magnitude of immune activation correlates with the level of HIV viremia [4]. Increased expression of immune activation markers (HLA-DR+, CD38+, CD45RO+, and CD95) also correlates with higher apoptosis rates of CD4 T cells [5]. Importantly, suppression of viral replication with HAART treatment reduces immune activation [6, 7], normalizes expression of activation markers, and decreases CD4 T cell apoptosis [8-10].

As a result of chronic immune activation, central memory and naïve CD4 T cells are constantly recruited into the effector pool [11, 12] resulting in dramatic changes in populations of naïve, effector and central memory CD4 T cells [13]. The mechanisms of how the ratio of these CD4 T cells is altered during HIV infection are not completely understood.

In particular, chronic HIV infection results in an increased number of CD4 T cells with a peculiar double positive CD45RO+/CD45RA+/CD25+CD4+ phenotype, and this subset is further increased following intermittent IL-2 therapy [14]. The origin of this subset was previously ascribed to be transition phase of naïve CD45RO-/CD45RA+CD4 T cells transitioning to a memory CD45RO+/CD45RA-CD4 T cell phenotype [15]. Furthermore, HAART with IL-2 therapy selectively increases the number of activated CD4 T cell expressing CD45RO+/CD25+ [16]. Here we describe a

novel effect of HIV-1 gp120 signaling through CD4 resulting in altered CD45 isoform expression by CD4 T cells.

MATERIALS AND METHODOLOGY

Cell Culture and Reagents

This protocol was reviewed and approved by the Mayo institutional review board. CD4+ T cells were isolated from the blood of healthy volunteer blood donors by using RosetteSep CD4 enrichment cocktail (StemCell Technologies, Vancouver, British Columbia, Canada), producing 98% pure CD4+ T cells as determined by flow cytometry. CD4 T cells were maintained in RPMI 1640 supplemented with 10% fetal bovine serum (Invitrogen, Carlsbad, CA), 2 mM L-glutamine, and antibiotics (penicillin 100 U/ml, streptomycin 100 µg/ml) at 0.5x10⁶ cells/ml. CD4+ T cells used in the various experiments were stimulated with PHA (1 µg/ml) for 24 hours, and then cells were washed twice with RPMI 1640 and maintained in media supplemented with 50 U/ml of IL-2 for 48-72 hours. CD4 T cells were incubated with HIV-1 X4 gp120IIIB (Immuno Diagnostics, Inc. Woburn, MA) or gp120 IIIB pretreated with soluble CD4 (1:2 ratio) (Immuno Diagnostics, Inc. Woburn, MA) at concentrations of 1 µg/ml/2x10⁶ cells for 24 hours at 37°C. AMD3100 (NIH AIDS Research and Reference program) was used at 2 µM for 30 minutes at 37°C. PP2 was purchased from CalBiochem (La Jolla, CA). Anti-CD4-PE, anti-CD25-FITC, anti-CD69-PE, anti-CD4PerCP, anti-CD62L-PE, anti-CD45RO-FITC, anti-CD45RA-PE-Cy-7, anti-HLA-DR-PE, AnnexinV-Cy-5, AnnexinV-APC, IgG1κ-PE-Cy7, IgG2a-FITC, IgG1κ-PE and propidium iodine were purchased from BD Biosciences (San Jose, CA).

Cell Death Analysis and Flow Cytometry

CD4 T cells were untreated or pre-incubated with specific inhibitors and stimulated with either BSA or with soluble gp120IIIB (10µg/ml) overnight. The following day, cell death was analyzed by staining with AnnexinV-Cy-5 and propidium iodine following the manufacturer's instructions

*Address correspondence to this author at the Division of Infectious Diseases, Mayo Clinic, 200 First Street SW, Rochester, MN 55905, USA; Tel: 507-266-5065; Fax: 507-284-3757; E-mail: badley.andrew@mayo.edu

from BD Biosciences (San Jose, CA). All experiments were performed at least three times.

T cell phenotyping studies were performed by using four-color Flow analysis on FACSCanto cytometer and using FACSDiva 6.0 software. Briefly, 2×10^6 cells were resuspended in 200 μ l of PBS+0.5%BSA, stained with the indicated primary conjugated antibodies for 20 minutes (anti-CD45RO-FITC, anti-CD45RA-PE-Cy-7, anti-CD27 APC and anti-CD62L-PE), washed, fixed and then analyzed. In some cases, for analysis of cell death, cells were stained in binding buffer (140 mM NaCl, 10 mM HEPES/NaOH (pH 7.4), 2.5 mM CaCl₂) as described above except that anti-CD62L-PE was substituted with AnnexinV-PE.

RESULTS

Gp120 Decreases CD45RO+ Memory Phenotype in Activated But Not Resting CD4 T cells

We and others have previously demonstrated that gp120 induces death of resting CD4 T cell in a CXCR4-p38 dependent manner [17]. However, the effect of gp120 on activated CD4 T cells is largely unknown. To examine the effect of gp120, resting (CD4+CD25-CD69-HLA-DR-) and activated (CD4+CD25+CD69+HLA-DR+) primary human CD4 T cells were treated with gp120 IIIB (10 μ g/ml) for 24 hours and then CD4 T cells were analyzed for CD45RO and CD45RA expression by flow cytometry. As shown in Fig. (1A), gp120 treatment results in a 50% decrease in the subset of single positive CD45RO+/CD45RA- (31.85 \pm 0.63% with

BSA, 13.80 \pm 3.73% with gp120, $p=0.01$) memory cells when activated but not resting CD4 T cells are treated with gp120 (34.25 \pm 1.1% with BSA, 28.95 \pm 1.96% with gp120, $p=0.08$). In parallel, there is a proportional increase in the subset of double positive CD45RO+/CD45RA+ mixed phenotype CD4+ T cells (Fig. 1B) as well as an increase in the proportion of Annexin-V positive CD4 T cells (Fig. 1C). While the increased apoptosis of resting cells following gp120 is due to CXCR4 signaling [17], the mechanisms by which gp120 causes death in activated cells are unknown.

The finding that gp120 treatment of activated cells increases the lymphoid homing marker, CD27 (13.8% of total CD4 T cells) preferentially in double positive CD45RO+/CD45RA+ cells suggests that these cells might be the specific cell type which dies following gp120 treatment of activated CD4 T cells (Fig. 2A, B). We therefore measured absolute number and proportion of Annexin-V positivity of double positive CD45RO+/CD45RA+, single positive CD45RO-/CD45RA+ and CD45RO+/CD45RA- subsets by using four color flow cytometry analysis. First we observed that CD45RO+RA- proportion decreased (39.42 \pm 3.44% with BSA, 27.31 \pm 6.38% with gp120) while CD45RO+RA+ population increased (35.97 \pm 2.70% with BSA, 49.42 \pm 10.05% with gp120), suggesting that the former transitioned to the latter (Fig. 3A). Next we observed that only double positive CD45RO+/CD45RA+ CD4 T cell subset contain a significant proportion of Annexin positive cells (~40%) in contrast to single positive CD45RO+/CD45RA- and CD45RO-/CD45RA+ subsets that do not (Fig. 3B). Therefore, gp120 signaling both

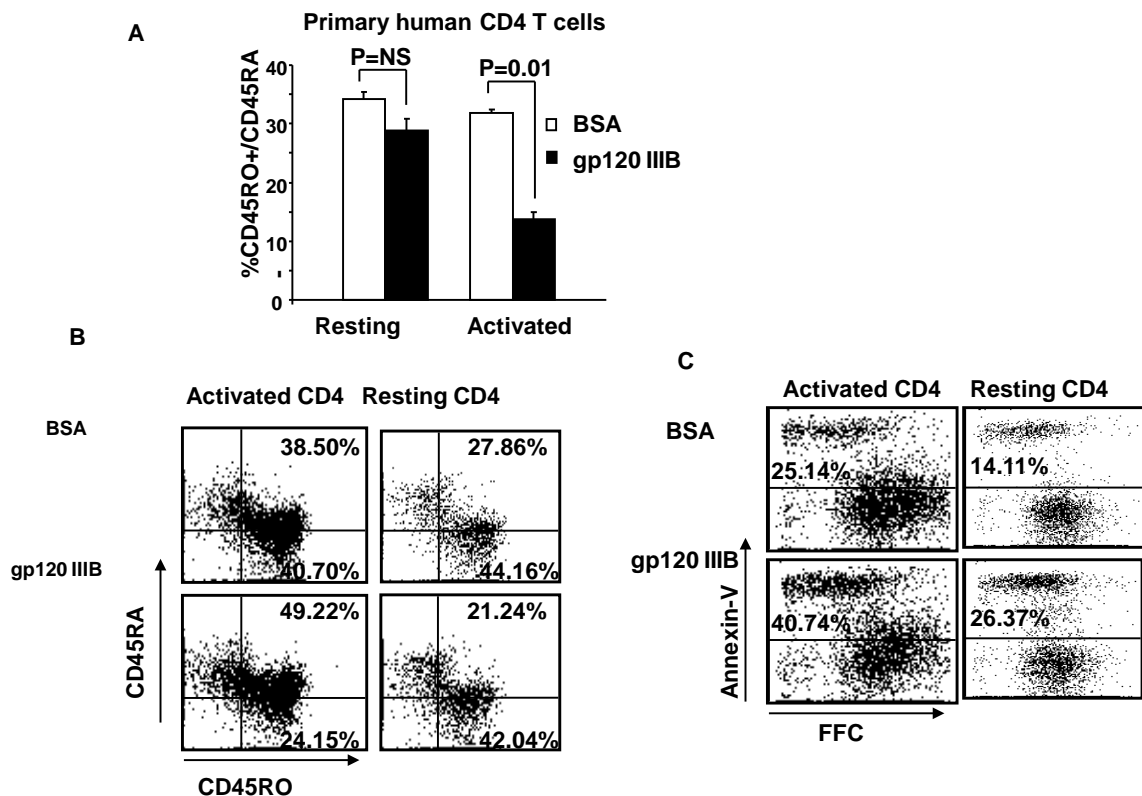


Fig. (1). Gp120 decreases CD45RO+/CD45RA- memory phenotype in activated but not resting primary human CD4 T cells. (A and B) CD4+ T cells (98% pure) were stimulated with PHA for 24 hours or left rested, and treated with either p120 (IIIB) (1 μ g/ml) or BSA control (1 μ g/ml) and analyzed for CD45RO and CD45RA (gated on CD4 population) by flow cytometry. (C) Resting and activated primary human CD4 T cells were treated with gp120 (IIIB) as above and analyzed for Annexin-V positive cells. Data is representative of three independent experiments.

activates cells towards a double positive phenotype and subsequently promotes their apoptotic death. Of interest, these Annexin positive CD45R0+/CD45 RA+ cells are also CD27+. Altogether these results indicate that gp120 treatment of activated CD4 T cells result in decrease memory CD45R0+/CD45RA- CD4 T cells due to accumulation of CD45R0+/CD45RA+ /CD27+ T cell subset that subsequently and selectively undergo apoptosis.

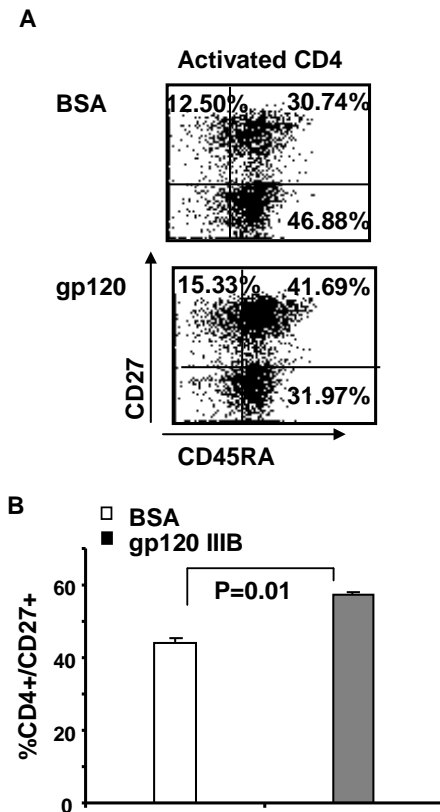


Fig. (2). Gp120 increases subset of CD27+/CD45RO+/CD4+ T cells. (A) Activated primary human CD4 T cells (98% pure) were treated with gp120 IIIIB and the expression of CD27 and CD45RA were analyzed by flow cytometry. (B) Pooled data from three independent experiments. P values were determined by Student's paired t test.

Gp120 Stimulated Accumulation of CD45R0+/CD45 RA+ CD4 T Cells Requires CD4 and Lck But Not CXCR4

To investigate whether gp120 effect on CD45R0+/CD45RA+ CD4 T cell is due to ligation to CD4 or CXCR4, we used either soluble CD4 or the CXCR4 inhibitor, AMD3100. Independent experiments verified the activity of AMD3100 by blocking SDF-induced chemotaxis (data not shown). As shown in Fig. (4A, B), inhibition of CD4 ligation by sCD4 [17] blocks the increase in double positive CD45R0+/CD45RA+ memory cells. Consistent with this observation, inhibition of Lck activity with PP2 inhibitor also blocks increase in CD45R0+/CD45RA+ CD4 T cell subset. Conversely inhibition of gp120-CXCR4 interaction with AMD3100 [17] does not block completely gp120 mediated increase of CD45R0+/CD45RA+ CD4 T cells. Therefore, our results demonstrate that gp120 ligation to CD4 and subsequent activation of Lck results in accumulation of dou-

ble positive CD45R0+/CD45RA+ memory CD4 T cells which subsequently and selectively undergo apoptosis in a CD4 dependent and CXCR4 independent manner.

DISCUSSION

In the current report we have shown that gp120 treatment of activated CD4 T cells decreases the subset of single positive CD45R0+/CD45RA- CD4 T cells and increases the subset of double positive CD45R0+/CD45RA+ CD4 T cells which selectively undergo apoptosis. These results imply the gp120 may play a role in depletion of memory CD45R0+/CD45RA- CD4 T during HIV.

Previously, the CD45R0+/CD45RA+ subset of CD4 T cells was characterized as a transition from naïve to effector phenotype during CD4 priming [15, 18] with increased expression of activation markers [19, 20]. The generation of this subset can occur in two ways: (i) the transition of recently activated of CD45RA+ T naïve cells to CD45R0+/CD45RA+ phenotype [15, 21] and; (ii) re-expression of CD45RA isoform in the absence of Ag stimulation [15]. The fact that only few CD45R0+/CD45RA+ cells express markers of recently activated cells [19, 20] and upregulation of CD45RA+ (up to 10%) by CD45R0+/CD45RA- cells [15] suggest that gp120 rather upregulates CD45RA+ on single positive CD45R0+/CD45RA- memory CD4 T cells rather than promoting CD45RO expression on naïve CD4 T cells. Indeed, increase of double positive CD45R0+/CD45 RA+ CD4 T cells is followed by proportional decrease in CD45R0+/CD45RA- memory but not decrease in naïve CD45RO-/CD45RA+ CD4 T cells.

Others have shown that PBMC stimulated with staphylococcal enterotoxin A (SEA) in the presence of V3-derived gp120 peptides results in increase of CCR5+/CXCR4+/CD45RO+ CD4 T cells followed by increased levels of apoptosis [22]. Our findings are in good agreement with this observation: first, both results demonstrate the gp120 mediated increase in transient state from naïve to activated CD4 T cells; second, both studies observe the increased apoptosis of activated CD4 T cells in the presence of gp120 or V3-derived peptide. However, our results highlight that gp120 priming of previously activated CD4 T cells reverses single positive CD45R0+/CD45RA- effector phenotype to double positive CD45R0+ /CD45RA+ transient phenotype.

The observation that double positive CD45R0+/CD45 RA+ preferentially undergo apoptosis presents one potential scenario of how inappropriately activated CD4 T cells die during HIV disease, and why IL-2 augments this process [16]. It is of further interest that gp120 treatment results in apoptosis of CD27+/CD45R0+/CD45RA+ cells. In fact, CD27 expression is rapidly upregulated following TCR stimulation [23] and CD27 signaling is essential for survival of Ag-primed CD4 T cells [24]. Therefore, either the lack of antigen specific signaling or the lack of CD27 signaling may explain the increased apoptosis of double positive CD45R0+/CD45RA+ following gp120 treatment. Finally, the observation that CD4 ligation by gp120 and subsequent activation of Lck is required for generation of double positive CD45R0+/CD45RA+ CD4 T cells is of relevance and suggests that inhibitors of CD4-gp120 interactions may reverse the expansion of double positive CD45R0+/CD45RA+ population and subsequent increased apoptosis during HIV infection.

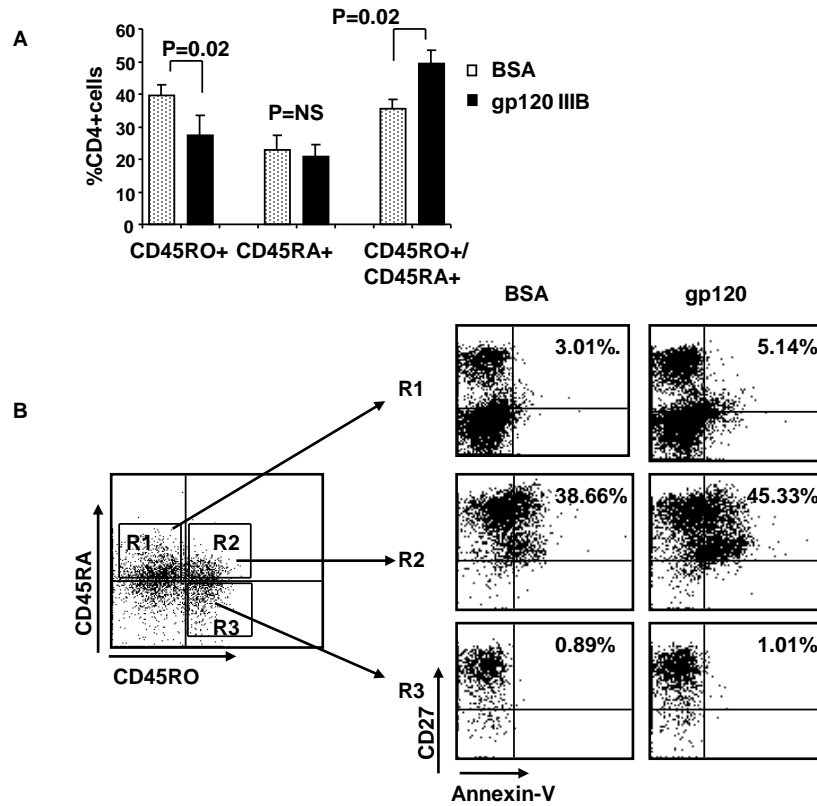


Fig. (3). Gp120 increases subset of double positive CD45RA+/CD45RO+ CD4 T cells that undergo apoptosis. (A) Activated primary human CD4 T cells were treated with gp120 IIIIB and CD45RA+/CD45RO-, CD45RA-/CD45RO+ and double positive CD45RA+/CD45RO+ cells were quantified by flow cytometry. (B) Same as Fig. (3A) except that CD27 expression and Annexin-V positivity were measured by four color flow cytometry. The data is representative of three independent experiments.

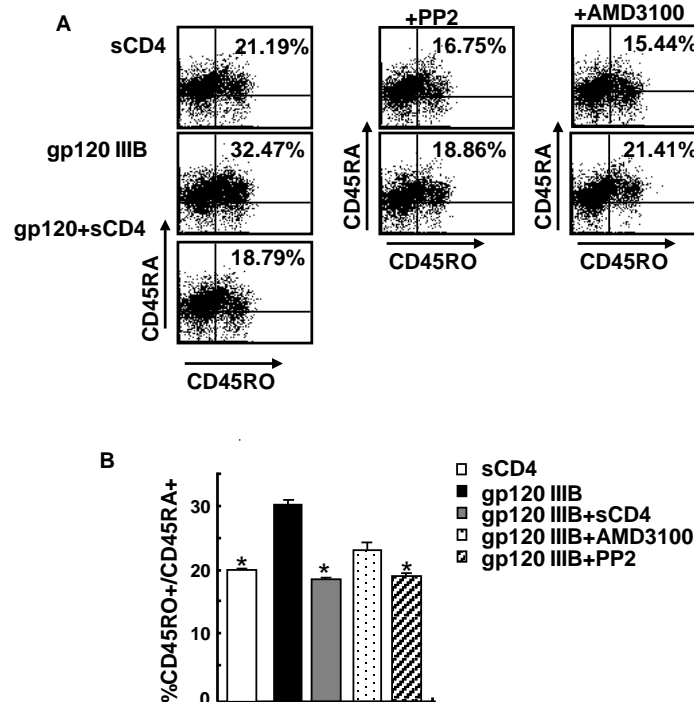


Fig. (4). CD4 and Lck are required for gp120 mediated increase of double positive CD45RA+/CD45RO+ CD4 T cells. (A) Activated primary human CD4 T cells were left untreated or were pre-treated with either Lck inhibitor, PP2 (5µM) or CXCR4 inhibitor, AMD3100 (2µM) or sCD4 (2µg/ml) followed by gp120. Then number of single positive CD45RA+/CD45RO-, CD45RA-/CD45RO+ and double positive CD45RA+/CD45RO+ cells were analyzed by flow cytometry. (B) The data represents three independent experiments as described above. P values were determined by Student's paired t test. *=P<0.05 compared to gp120 alone.

ACKNOWLEDGEMENTS

Grant Support: Dr. Andrew Badley is supported by the following grants: NIH R01 AI62261, R01 AI40384 and a Burroughs Wellcome Award ID #1005160.

REFERENCES

- [1] Lum J, Badley AD. In: Badley AD, Ed. Cell Death During HIV Infection. Boca Raton: Taylor and Francis Press 2006; pp. 109-126.
- [2] Douek DC, Betts MR, Hill BJ, *et al.* Evidence for increased T cell turnover and decreased thymic output in HIV infection. *J Immunol* 2001; 167: 6663-8.
- [3] Hellerstein M, Hanley MB, Cesar D, *et al.* Directly measured kinetics of circulating T lymphocytes in normal and HIV-1-infected humans. *Nat Med* 1999; 5: 83-9.
- [4] Perelson AS, Neumann AU, Markowitz M, Leonard JM, Ho DD. HIV-1 dynamics *in vivo*: virion clearance rate, infected cell life-span, and viral generation time. *Science* 1996; 271: 1582-6.
- [5] Gougeon ML, Lecoer H, Dulioust A, *et al.* Programmed cell death in peripheral lymphocytes from HIV-infected persons: increased susceptibility to apoptosis of CD4 and CD8 T cells correlates with lymphocyte activation and with disease progression. *J Immunol* 1996; 156: 3509-20.
- [6] Hazenberg MD, Stuart JW, Otto SA, *et al.* T-cell division in human immunodeficiency virus (HIV)-1 infection is mainly due to immune activation: a longitudinal analysis in patients before and during highly active antiretroviral therapy (HAART). *Blood* 2000; 95: 249-55.
- [7] Lempicki RA, Kovacs JA, Baseler MW, *et al.* Impact of HIV-1 infection and highly active antiretroviral therapy on the kinetics of CD4+ and CD8+ T cell turnover in HIV-infected patients. *Proc Natl Acad Sci USA* 2000; 97: 13778-83.
- [8] Badley AD, Dockrell DH, Algeciras A, *et al.* *In vivo* analysis of Fas/FasL interactions in HIV-infected patients. *J Clin Invest* 1998; 102: 79-87.
- [9] Bohler T, Walcher J, Holzl-Wenig G, *et al.* Early effects of antiretroviral combination therapy on activation, apoptosis and regeneration of T cells in HIV-1-infected children and adolescents. *AIDS* 1999; 13: 779-89.
- [10] Giorgi JV, Majchrowicz MA, Johnson TD, Hultin P, Matud J, Detels R. Immunologic effects of combined protease inhibitor and reverse transcriptase inhibitor therapy in previously treated chronic HIV-1 infection. *AIDS* 1998; 12: 1833-44.
- [11] Hazenberg MD, Hamann D, Schuitemaker H, Miedema F. T cell depletion in HIV-1 infection: how CD4+ T cells go out of stock. *Nat Immunol* 2000; 1: 285-9.
- [12] Silvestri G, Feinberg MB. Turnover of lymphocytes and conceptual paradigms in HIV infection. *J Clin Invest* 2003; 112: 821-4.
- [13] McCune JM. The dynamics of CD4+ T-cell depletion in HIV disease. *Nature* 2001; 410: 974-9.
- [14] Sereti I, Martinez-Wilson H, Metcalf JA, *et al.* Long-term effects of intermittent interleukin 2 therapy in patients with HIV infection: characterization of a novel subset of CD4(+)/CD25(+) T cells. *Blood* 2002; 100: 2159-67.
- [15] Hamann D, Baars PA, Hooibrink B, van Lier RW. Heterogeneity of the human CD4+ T-cell population: two distinct CD4+ T-cell subsets characterized by coexpression of CD45RA and CD45RO isoforms. *Blood* 1996; 88: 3513-21.
- [16] Sereti I, Herpin B, Metcalf JA, *et al.* CD4 T cell expansions are associated with increased apoptosis rates of T lymphocytes during IL-2 cycles in HIV infected patients. *AIDS* 2001; 15: 1765-75.
- [17] Trushin SA, Algeciras-Schimmich A, Vlahakis SR, *et al.* Glycoprotein 120 binding to CXCR4 causes p38-dependent primary T cell death that is facilitated by, but does not require cell-associated CD4. *J Immunol* 2007; 178: 4846-53.
- [18] Picker LJ, Treer JR, Ferguson-Darnell B, Collins PA, Buck D, Terstappen LW. Control of lymphocyte recirculation in man. I. Differential regulation of the peripheral lymph node homing receptor L-selectin on T cells during the virgin to memory cell transition. *J Immunol* 1993; 150: 1105-21.
- [19] Baars PA, Maurice MM, Rep M, Hooibrink B, van Lier RA. Heterogeneity of the circulating human CD4+ T cell population. Further evidence that the CD4+CD45RA-CD27- T cell subset contains specialized primed T cells. *J Immunol* 1995; 154: 17-25.
- [20] Prince HE, York J, Jensen ER. Phenotypic comparison of the three populations of human lymphocytes defined by CD45RO and CD45RA expression. *Cell Immunol* 1992; 145: 254-62.
- [21] Wallace DL, Beverley PC. Phenotypic changes associated with activation of CD45RA+ and CD45RO+ T cells. *Immunology* 1990; 69: 460-7.
- [22] Porichis F, Vlata Z, Hatzidakis G, Spandidos DA, Krambovitis E. HIV-1 gp120/V3-derived epitopes promote activation-induced cell death to superantigen-stimulated CD4+/CD45RO+ T cells. *Immunol Lett* 2007; 108: 97-102.
- [23] Watts TH. TNF/TNFR family members in costimulation of T cell responses. *Annu Rev Immunol* 2005; 23: 23-68.
- [24] Hendriks J, Xiao Y, Borst J. CD27 promotes survival of activated T cells and complements CD28 in generation and establishment of the effector T cell pool. *J Exp Med* 2003; 198: 1369-80.

Received: February 26, 2009

Revised: March 6, 2009

Accepted: March 12, 2009

© Trushin *et al.*; Licensee Bentham Open.

This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.