

Human Papillomavirus Infections and Cancer Stem Cells of Tumors from the Uterine Cervix

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Abstract: Different rate of development of productive infections (as low grade cervical intraepithelial neoplasias), or high grade lesions and cervical malignant tumors associated with infections of the Transformation zone (TZ) by High-Risk Human Papillomavirus (HR-HPV), could suggest that different epithelial host target cells could exist. If there is more than one target cell, their differential infection by HR-HPV may play a central role in the development of cervical cancer. Recently, the concept that cancer might arise from a rare population of cells with stem cell-like properties has received support in several solid tumors, including cervical cancer (CC). According to the cancer stem cell (CSC) hypothesis, CC can now be considered a disease in which stem cells of the TZ are converted to cervical cancer stem cells by the interplay between HR-HPV viral oncogenes and cellular alterations that are thought to be finally responsible for tumor initiation and maintenance. Current studies of CSC could provide novel insights regarding tumor initiation and progression, their relation with viral proteins and interplay with the tumor micro-environment. This review will focus on the biology of cervical cancer stem cells, which might contribute to our understanding of the mechanisms responsible for cervical tumor development.

Keywords: Cancer stem cells, cervical cancer, human papillomavirus, squamous cell carcinoma, transformation zone.

INTRODUCTION

Based on morphological observations carried out by Rudolf Virchow and Julius Cohnheim in the late 19th century, it has been since then clearly recognized that tumors exhibit a varied cellular heterogeneity. Within a tumor, cell populations with diverse phenotypic features coexist, manifested in their morphology, surface antigens expression, differentiation proteins, among others. Also, tumor populations display functional properties reflected in different levels of proliferation as observed *in vivo* by colony formation assays, tumorigenicity as manifested *in vivo* by xeno-transplantations, different metastatic potential, and response to treatment, among the most outstanding [1].

Attempts have been made to explain this cellular heterogeneity mainly based on two models of carcinogenesis. A stochastic model stating that cellular heterogeneity arises from biologically equivalent populations as a product of selection and expansion of clones with

growth advantages. Thus, based on these assumptions, any cell within a tumor will be able to form new tumor [1]. Contrariwise, a hierarchical model stating that cellular heterogeneity arises from different phenotypic and functional populations within a tumor that contains a particular subpopulation, denominated cancer stem cells (CSC), which is the only subpopulation able to form new tumors and metastases [2]. In 2006, the consensus definition of CSC from the Workshop of the American Association for Cancer Research is a cell within a tumor that possesses the capacity to self-renew and originate all of the heterogeneous lineages of cells that comprise a tumor. Thus, CSC can only be defined experimentally by their ability to generate a continuously growing tumor. The implementation of this approach explains the use of alternative terms in the literature, such as “tumor-initiating cells” and “tumorigenic cells” to describe putative cancer stem cells [3] (for a detailed review of the biology and model of CSC see Dalerba *et al.*, 2007[4] and Lobo *et al.*, 2007 [5]). Since the mid-1990's, sufficient experimental evidence has been generated that supports the existence of CSC in different cancers.

Identification and isolation of CSC began in 1994; Lapidot *et al.* identified human Acute myeloid leukemia

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(AML)-initiating cells and fractionated them on the basis of cell-surface-marker expression [6]. Bonnet *et al.*, using purified cell populations found that leukemia-initiating cells were exclusively CD34⁺⁺ CD38⁻ [7]. Concerning solid tumors, Al-Hajj *et al.* identified and isolated tumorigenic cells from breast cancer, as CD44⁺ CD24^{-/low} lineage⁻ cells [8]. Following this pioneer work, CSC have been identified in many other tumors such as brain [9], prostate [10], colon [11-13], neck and head [14], pancreas [15, 16], liver [17] and melanoma [18] (for a detailed review, see O'Brien *et al.* [11] and Ailles *et al.* [19]).

Additional to surface antigens, other methodological strategies have been widely used to identify and isolate CSC. Goodell *et al.*, experimenting with Hoechst 33342 dye in murine bone marrow cells, found, by measuring the fluorescence of the dye at two wavelengths simultaneously, a small side population enriched with hematopoietic stem cells markers and reconstitution activity *in vivo*, which was termed Side Population (SP) [20]. In addition, formation of floating cell aggregates or spheres comprises another approach. In 1992, cells isolated from the striatum of the adult mouse brain were induced to proliferate *in vitro*, as spheres, by epidermal growth factor [21]. To date, several cancer models have been evaluated for their ability to generate CSC-enriched spheres; some of these include breast [22], melanoma [23], ovarian [24], and prostate [25].

Similar to other tissues, stem cells from normal female reproductive tract are thought to play a pivotal role in its physiology and are likely to be involved in its response to injury and disease, and could be at the origin of many tumors of the uterine cervix, by generating CSC. Cell surface antigens such as CD44, CD117, CD133, and MYD88 have been used to isolate ovarian CSC; these cells also exclude Hoechst 33342. Clonogenic epithelial and stromal cells, side population, and label-retaining cells with stem-like features from endometrial cancer tissues have been recently described (for a detailed review, see López *et al.* [26] and Table 1). With regard to cervical cancer (CC), over the past years, some studies have attempted to isolate CSC from primary tumors and cell lines based on cell surface markers, Hoechst staining, formation of spheres, and embryonic stem markers. However, few studies have evaluated the tumorigenic potential or have performed a more complete characterization (Table 2). The ability to prospectively identify CSC will facilitate the elucidation of pathways that regulate their growth and survival. Furthermore, because these cells drive tumor development, strategies designed to target this population may lead to more effective therapies.

Stem cells are defined as cells that possess the ability to perpetuate themselves by means of self-renewal, to generate mature cells of a particular tissue through differentiation of a progeny, consisting of both transiently amplifying cells and differentiated cells, and responding to homeostatic controls. These cells have been found to respond differently to therapy. Importantly stem cells, although with an unlimited capacity to self-renew, usually exhibit a limited replicating activity. Signaling pathways that normally regulate stem cell self-renewal can frequently lead to carcinogenesis when deregulated; thus, stem cells are now believed to constitute

Table 1. Markers of Human Uterine and Ovarian Cancer Stem Cells

Uterine Cancer Stem Cells	Clonogenic cells	Hubbard, 2009 [71]
	SP ⁺	Friel, 2008 [72]
		Kato, 2010 [73]
	CDs	Rutella, 2009 [74]
		Afify, 2006 [75, 76]
Ovarian Cancer Stem Cells	Clonogenic cells	Bapat, 2005 [77]
	SP ⁺	Szoket, 2006 [78]
	CD44 ⁺ , CD117 ⁺	Zhang, 2008 [24]
	CD133 ⁺	Curley, 2009 [79]
		Baba, 2009 [80]
		Ferrandina, 2008 [81]
		Kusumbe, 2009 [82]
	CD44 ⁺ , MYD88 ⁺	Alvero, 2009 [83]
		Kusumbe, 2009 [82]
CD44 ⁺ , MYD88 ⁺	Alvero, 2009 [83]	

Abbreviations: CD, Cluster of differentiation; SP, Side population.

in the majority of the cases, the targets of transformation. Arguments supporting this include the fact that stem cells often persist for long periods of time, many years or the whole individuals life, in contrast to mature cells in highly proliferative tissues, which die after short periods of time (days or weeks). This means that there is an exponential higher opportunity for mutations to accumulate in individual stem cells than in mature cell types [27]. Therefore, together with the fact that these cells may be target cells for HR-HPV infections, it is easy to imagine that this could represent a major step toward cervical carcinogenesis. HR-HPV oncogenes are well known to influence cell cycle controls leading to malignant transformation. The fact that incorporation of the viral genome into the cell nucleus requires the entry of the cell into mitosis [28]; this makes us propose that it could be possible for a viral genome to persist in a "latent" state in stem cells that do not divide for years. It is plausible that since this is an undifferentiated, multiplying cell, it makes it possible for a viral genome to incorporate into its DNA during the cell cycle (Fig. 1). A stem or progenitor cell can be the origin of tumor growth by disturbance of the asymmetric division process and can therefore be a candidate for the target cell of HR-HPV infection [2]. This particularly could explain the long latency period between viral infection and CC development.

MARKERS OF CERVICAL CANCER STEM CELL

The well-known process of squamous metaplasia occurring in the TZ of the cervix also suggests an important role for the reserve cell as the basal cell for columnar as well as squamous epithelial regeneration. Morphological studies have shown that reserve cells are undifferentiated, omnipotent cells that possess the capacity to undergo squamous differentiation (metaplasia). Basal cells of the ectocervical squamous epithelium are more differentiated,

Table 2. Selective Markers of Human Epithelial Stem Cells in Cervical Cancer

Sample	CSCs Markers	% Tumor Cells with CSCs Marker	Min. CSCs to Engraft	Recapitulation of Original Tumor Phenotype	Serial Transplant Performed	Additional Assays	Ref.
Normal cervix and CIN I-II-III	p63 CK17	*	-	-	-	-	Martens, 2004 [32]
Normal cervix and CSCC	Nanog Musashi-1 Nucleostemin	Normal cervix < 65% and CSCC > 90%, with all markers**	-	-	-	Correlation with clinicopathological features	Ye, 2008 [39]
Normal cervix and CSCC	PSCA PIWIL1 TBX2	Normal cervix < 9% and CSCC > 50%	-	-	-	Correlation with clinicopathological features and HPV16	Liu, 2010 [40]
Normal cervix, CIN-III, CSCC, cell lines and spheres	Sox2	SiHa and spheres from primary cancer tissues: positive. Normal cervix: 25%, CIN III: 83%, CSCC: 77%	10 ⁶ SiHa-Sox2 cells	Yes	No	Proliferation and clonogenicity of exogenous Sox2 in HeLa and SiHa cell lines	Ji, 2010 [42]
Invasive carcinomas and adenocarcinomas	ALDH	Invasive squamous carcinoma 25% and adenocarcinomas 20%	-	-	-	CD133 expression	Yao, 2011 [45]
SiHa and Ca Ski cell lines	Spheres ALDH	CaSki: 32% SiHa: 0% Ca Ski: 30% SiHa: 24%	-	-	-	A431 sphere characterization	Bortolomai, 2010 [46]
HeLa cell line	Spheres ALDH1 CD44	HeLa-Twist elevate the level of SFE, ALDH1 and CD44 in HeLa	-	-	-	Activation of β -catenin and Akt pathways by Twist and suppression of CD44 expression by inhibition of signaling pathways.	Li, 2011 [51]
CSCC	Spheres	-	10 ⁵	Yes	No	Chemoresistance assay Expression of embryonic and stemness genes	Feng, 2009 [52]
HeLa, SiHa, Ca Ski and C33A cell lines	Spheres	HeLa: 18%	10 ⁴	-	No	Chemoresistance assay and gene silencing of E6 in HeLa sphere-forming cells	Gu, 2011 [53]
HeLa, SiHa, Ca Ski and C-41 cell lines	Spheres-CD49f	4-12%	10 ³	Yes	Yes	Gene expression analysis and radioresistance assay	López, 2012 [44]

Abbreviations: CSCs, Cancer stem cells; CIN, Cervical intraepithelial neoplasia; CK, Cytokeratin; CSCC, Cervical squamous cell carcinoma; PSCA, Prostate stem cell antigen; PIWIL1, Piwi-like protein 1; ALDH, Aldehyde dehydrogenase. *In normal ectocervical epithelium, p63 was restricted to the basal compartment in 76-100% of cells in all cases and CK17, was in 26-50%. In endocervical epithelium, p63 and CK17 were not expressed. Subcolumnar reserve cells in the transitional zone and endocervix showed p63 and CK17 expression in 76-100% of cells in all cases. In CIN I and -II, the basal compartment showed p63 expression in 76-100% of cells in all cases. In CIN III lesions, staining was noted throughout the entire epithelial thickness. CK17 expression was noted in the basal compartment of all CIN lesions, irrespective of their grade, in 26-50% of cells in all cases. ** Expression >5%

devoted to the formation of squamous cells, and therefore less suitable as stem cells of the cervical epithelium [29]. Previously, transcription factor p63 was described as marker for basal squamous cells and subcolumnar reserve cells in cervical epithelium [30, 31]. Martens *et al.* described the identification of a uterine cervical epithelium stem-cell population using monoclonal antibodies against p63 and cytokeratin 17 (CK17) in normal ectocervical, endocervical, and cervical intraepithelial neoplasia (CIN) grades I, II and III. Reserve cells and reserve cell hyperplasia showed strong

staining of p63 (nuclear) and CK17 (cytoplasmatic). Cytoplasmatic staining with the monoclonal antibody against CK17 was shown in the basal compartment of ectocervical epithelium and of CIN lesions irrespective of their grade. Reserve cells and reserve cell hyperplasia showed strong expression in all cases [32]. Additionally, in the adult cervix, reserve cells demonstrate a specific phenotype that expresses, among others, the BCL-2 gene [33]. In all cases, p63 was strongly expressed in the basal layer of the

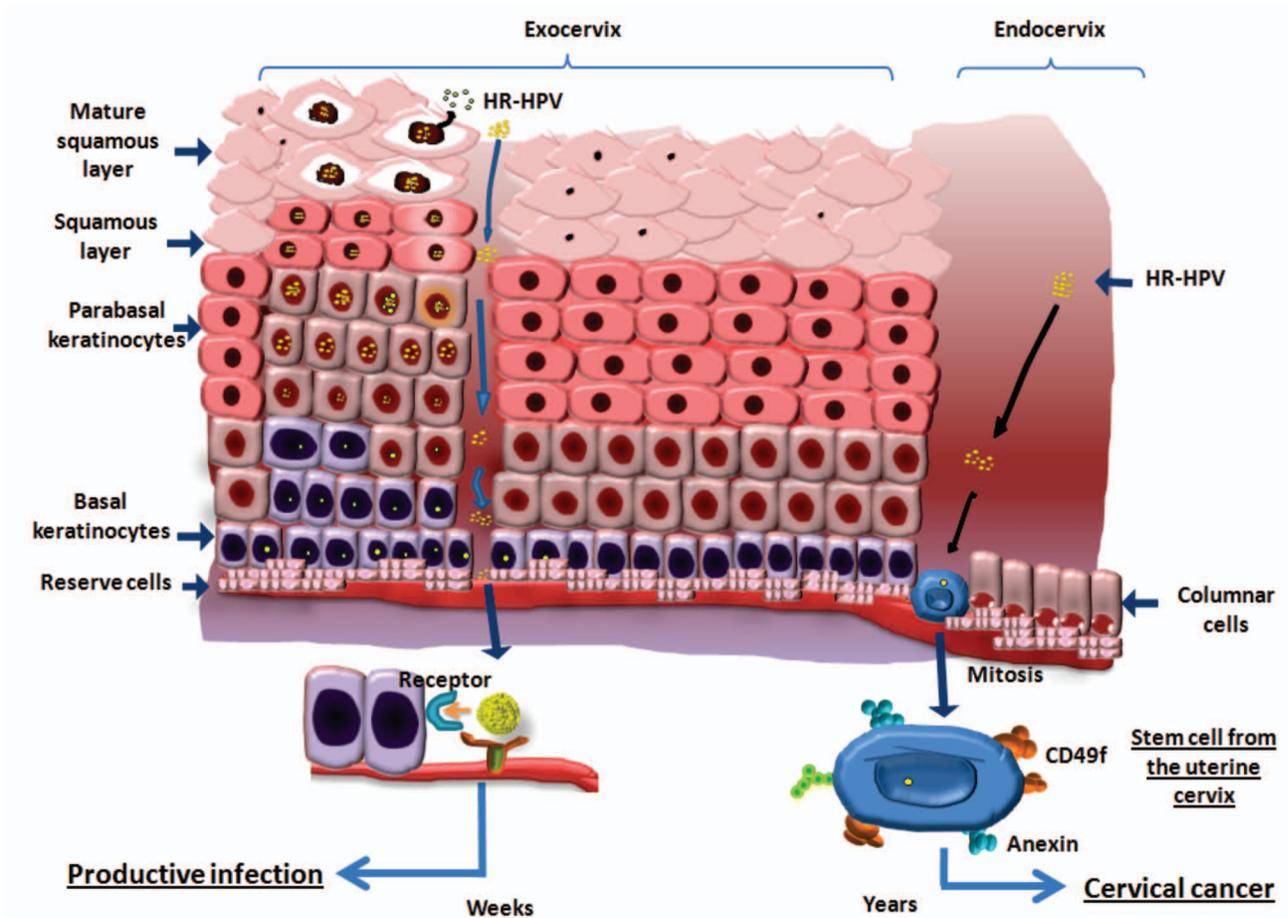


Fig. (1). Two different consequences proposed for infection of basal cells or cervical stem cells with High Risk-Human Papillomavirus (HR-HPV). One area where cervical stem cells are likely to be found comprises the squamocolumnar junction, where the ecto- and the endocervix meet and where reserve cells are frequently found. We propose that HR-HPV infect in the cervix both basal and stem cells, in the former leading to a productive infection within weeks and in the latter, probably generating cancer years later.

ectocervical epithelia and in the basal layers of CIN lesions irrespective of grade.

According to other studies [34, 35], p63 was expressed in basal and suprabasal cells of mature and immature metaplastic squamous cells and in subcolumnar reserve cells of the TZ in the uterine cervix. In normal cervix, the transition from squamous to columnar cells coincided with loss of p63 expression, whereas its expression was inversely correlated with both squamous cell maturation and non-squamous cell differentiation. Recently, it was shown that p63 induces a stem cell phenotype in the MCF7 breast carcinoma cell line and that it could be a marker of breast CSC; in this cell line, p63 led to increased cancer cell proliferation, clonogenicity, anchorage-independent growth, and the incidence of tumor xenografts formed *in vivo* in immunosuppressed mice [36]. These results suggest that p63 might be a tumor-initiating transcription factor in CC and that p63 is one of the possible markers of CSC in epithelial tissues. Recently, Mighty *et al.* found that, upon differentiation of normal keratinocytes, p63 levels decreased rapidly, while higher levels were retained in HPV-positive cells, and that late p63 gene expression regulates the

expression of cell cycle regulators, such as, cyclin A, cyclin B1, cdk1, and cdc25c. In addition, activation of the DNA repair pathway was necessary for genome amplification, and the expression of two members, BRCA2 and RAD51, was altered in the absence of p63 in HPV-positive cells [37].

Furthermore, Martens *et al.* focused on the distribution pattern and marker profile of reserve cells along the entire length of the adult endocervical canal. The authors employed sagittal sections from 10 normal uteri, comprising the region from ectocervix to lower uterine cavity; these sections were histologically examined and immunostained for p63, bcl-2, and CK 5, 7, 8, and 17. They observed that reserve cells are present in all investigated cervixes along the entire cervical canal. The concentration of subglandular reserve cells was highest near the squamocolumnar junction and in the upper third of the cervix. These authors found a discontinuous distribution pattern of reserve cells in all of the uterine cervix; on the basis of their marker profile, they defined two subpopulations of reserve cells in the adult human uterine cervical epithelium: a CK17-positive subpopulation in the lower part of the cervical canal with progenitor cells for the squamous and columnar epithelium, and a subpopulation of

CK17-negative reserve cells with progenitor cells only for columnar epithelium [38].

The stem cell-abundant proteins Nanog, Nucleostemin (NS) and Musashi1 (Msi1) are highly expressed in undifferentiated embryonic stem cells and negatively regulate stem-cell differentiation, proliferation, and asymmetric division, respectively. Ye *et al.* examined the expression of Nanog, NS, and Msi1 in cervical epithelial lesions of varying severity and in cervical carcinomas by immunohistochemical analysis and assessed their association with several prognostic variables. These findings indicate that Nanog, NS, and Msi1 may be involved in carcinogenesis of the cervix and the progression of cervical carcinoma. However, there were no positive correlations between Nanog, NS, and Msi1 expression levels and the clinical pathological prognostic factors analyzed, indicating that overexpression of these three stem cell-abundant proteins in the cervical epithelium is not related with the prognosis of cervical carcinoma [39].

Few reports have emerged concerning the expression of stem cell-associated genes *PSCA* and *PIWIL1* and transcriptional factor *TBX2* in cervical squamous cell carcinoma (CSCC). For example, Liu *et al.* investigated the expression of these CSC-associated gene products in CSCC and their relationship with HPV16 in order to provide some basic information on potential molecular mechanisms of the effect of HPV16 infection on cervical carcinogenesis. They evaluated the expression of *PSCA*, *PIWIL1*, *TBX2* and HPV16 E7 in 59 CSCC and Matched adjacent normal cervix (MANC), employing the streptavidin-peroxidase method. *PSCA*, *PIWIL1* and *TBX2* expression rates in MANC were 6 (3/52), 8 (4/52), and 2% (1/52), respectively, but those in CSCC comprised 62 (32/52), 75 (39/52), and 52% (27/52), respectively. However, the increased expression of *PSCA*, *PIWIL1*, and *TBX2* observed in CSCC had no correlation with the patient's age or histological grade. Elevated expression of *PSCA* and *PIWIL1* was associated with invasion of CSCC, and up-regulated expression of *TBX2* had a positive association with lymph node metastasis. HPV16 had a statistical positive correlation with *PSCA*, *PIWIL1*, and *TBX2* in CSCC, which raised the question of whether HPV16 viral oncoproteins might be involved in up-regulating the expression of CSC-associated genes and suggests a complex molecular mechanism for the role of HPV16 infection during cervical carcinogenesis of CSCC [40]. Recently, Ye *et al.* identified a *Piwil2*-like (PL2L) protein that is predominantly expressed in human primary and metastatic cervical cancers in association with Nuclear factor-kappa beta (NF- κ B) that promote tumorigenesis [41]. Further researches should be conducted utilizing a cell or animal model to demonstrate a definite role for HPV16 in the overexpression of *PSCA*, *PIWIL1*, PL2L, and *TBX2*.

SOX2 is a member of the SOX gene family that encodes transcription factors expressed in several phases of embryonic development, which affects cell fate and differentiation. Ji *et al.* examined the expression of SOX2 in normal and pathologic cervical tissues. They also compared SOX2 expression in CC tumorspheres and its differentiated cells. They demonstrated that SOX2 may play an important role in tumorigenesis and suggest a possible therapeutic target molecule in CC [42].

Another marker associated with the pluripotent state of cells comprising the inner cell mass of pre-implantation embryos and that has been known to play a critical role in the maintenance of the pluripotency of embryonic stem cells is the POU-domain transcription factor OCT4. Cantz *et al.* were unable to detect staining of the OCT4 transcription factor in the nucleus in the cervical carcinoma cell line HeLa by immunofluorescence using two different monoclonal antibodies [43].

Another useful CSC marker is the Aldehyde dehydrogenase (ALDH1), a detoxifying enzyme responsible for the oxidation of retinol into retinoic acid and which plays a role in the early stem-cell differentiation. When expression of ALDH1 in cervical carcinoma was evaluated by Yao *et al.* by immunohistochemistry of 23/89 samples of invasive squamous carcinoma and 4/20 samples of adenocarcinomas, they were found to exhibit immunoreactivity to ALDH1. They found lower levels of CD133 in HeLa, SiHa and CaSki cervical cell lines similar to showed by López *et al.* (2012) [44], define ALDH1 as a marker for identifying CSC and suggest that cervical carcinoma contains a small subpopulation of cells with a cancer stem cell-like phenotype [45].

CHARACTERIZING CERVICAL STEM CELLS USING SPHERE FORMATION ASSAY

Bortolomai *et al.* compared CaSki, SiHa and A431 cancer cell lines for sphere-forming efficiency and intrinsic ALDH enzymatic activity. A431 cells exhibited the highest fraction of ALDH-positive cells and good correlation between ALDH positivity and sphere forming efficiency. Therefore, they selected the A431 cell line as a model with which to evaluate the experimental procedures adopted for CSC characterization in tumor cell lines [46]. However, to the best of our knowledge, A431 cells were established from an epidermoid carcinoma in the vulva of an 85-year-old female patient [47].

Studies deriving from many laboratories indicate that Epithelial-mesenchymal transition (EMT) can endow cells with stem cell-like characteristics [48-50]. Li *et al.* induced EMT in breast cancer MCF7 and CC HeLa cells with expression of Twist, a key transcriptional factor of EMT. They also found that cancer stem cell-like traits, such as tumorsphere formation and the expression of ALDH1 and CD44 were significantly elevated in Twist-overexpressing cells. Interestingly, the authors showed that β -catenin and Akt pathways were activated in these Twist-overexpressing cells. Activation of β -catenin correlated with CD44 expression. Knockdown of β -catenin expression and inhibition of the Akt pathway greatly suppressed the expression of CD44. This study provides several new insights into the regulation of EMT and the cell differentiation program. First, these results indicate that activation of β -catenin and Akt pathways is critical for the maintenance of EMT-associated stem cell-like properties. Second, these results suggest that targeting the β -catenin and Akt pathways can suppress EMT-associated stem cell-like properties [51].

A CSC population from primary carcinoma of the cervix uteri was recently identified. Cervical carcinoma from 19 patients was disaggregated and subjected to stem cell-

selective growth conditions. Eight of 19 tumor-derived cultures encompassed CSC capable of self-renewal and extensive proliferation as clonal non-adherent spherical clusters. Cell markers of spheroids were identified as CD44⁺ CK17⁺. Cell survival assays showed that sphere-forming cells were only 48% inhibited by doxorubicin compared with non-sphere forming cells, which were 78% inhibited by this drug. To investigate the tumorigenicity of these CSC, xenograftment of 1×10^5 dissociated spheroid cells allowed full recapitulation of the original tumor, whereas the same amount of tumor cells without non-adherent spheroid selection remained non-tumorigenic. Stem properties of these spheroid cells were further established by RT-PCR and WB, demonstrating the expression of embryonic and adult stemness-related gene products (OCT4, PIWIL2, C-MYC, STAT3, and SOX2) [52].

Employing a sphere culture method that favors the growth of self-renewal cells, Gu *et al.* isolated Sphere-forming cells (SFC) from CC cell lines HeLa and SiHa. HeLa-SFC were more tumorigenic, as evidenced by the growth of tumors following injection of immunodeficient mice with 1×10^4 cells, compared with 1×10^6 parental HeLa cells that were required to grow similarly sized tumors within the same time frame. They further demonstrated that HeLa-SFC expressed a higher level (6.9-fold) of the HPV E6 oncogene, compared with that of parental HeLa cells. Gene silencing of E6 with a lentiviral-short-hairpin RNA inhibited HeLa-SFC sphere formation and cell growth. The authors then measured the expression of self-renewal genes, Transformation growth factor- β (TGF- β), and the leukemia-inhibitory factor in short-hairpin RNA-transduced HeLa-SFC and found that expression of all three TGF- β isoforms was significantly down-regulated, while the leukemia-inhibitory factor remained unchanged. This result suggests that E6 silencing exerts a specific effect on the expression of self-renewal gene TGF- β [53]. To prove down-regulation of TGF- β after E6 silencing, the authors measured the expression of both H-RAS and K-RAS genes, downstream components of the TGF- β pathway. RAS expression was also markedly decreased after TGF- β down-regulation, suggesting that the growth-inhibitory effect could be *via* the TGF- β pathway. Gu *et al.* postulate that downregulation of the PI3K/Akt pathway after E6 silencing comprise the link that indirectly affects TGF- β expression, but this requires further investigation [53].

Recently, López *et al.* characterized a self-renewing subpopulation of CSC among four well-known human cancer-derived cell lines (HeLa, SiHa, Ca Ski, and C-41) and found that these express the CSC markers characteristic of the female reproductive system including CD44, ITGB1 (CD29), PSCA, NTSE (CD73), ENG (CD105), MYC (c-Myc), PCGF4 (BMI-1), and ABCG2. Other epithelial CSC markers found included ITGB6, ALCAM (CD166), and MET (c-Met)[44]. Interestingly, components of the double-strand break DNA repair machinery and genes involved in the metabolism of reactive oxygen species were also up-regulated and indeed, dose-dependent radiation assays indicated that CSC-enriched populations exhibit increased resistance to ionizing radiation. They found that CSC enriched as spheroids could generate reproducible tumor phenotypes in immunodeficient nu-nu mice and could be propagated serially. Injection of 1×10^3 dissociated cells

from spheroids induced tumors in the majority of animals, whereas injection of monolayer-grown 1×10^5 cells remained nontumorigenic. Sphere-derived CSC highly expressed CD49f. Interestingly, CD49f (ITGA6), an alpha-6 integrin, has been postulated as the primary receptor protein in natural HR-HPV infection [54-57]; thus, it was postulated that adult reserve cells with CD49f expression could be preferentially targeted by HR-HPV types during cervical carcinogenesis. In addition, EMT transition-associated markers were found highly expressed in spheroid cells [44].

CERVICAL CANCER AND HUMAN PAPILLOMAVIRUS INFECTIONS

Cancer of the uterine cervix is a leading cause of death in women. It is the second most frequent type of cancer in women worldwide, preceded only by breast cancer [58]; GLOBOCAN 2008 reported that each year, ~530,000 [59] women worldwide are diagnosed with invasive CC and more than one half of these die from this disease. Eighty percent of these deaths occur in developing countries, and CC remains a cause of significant morbidity and mortality. It is therefore essential to develop a deeper understanding of the biology of this disease in order to find more effective therapies, because the majority of patients with CC receive standard radiotherapy and chemotherapy. However, clinical outcomes vary significantly and are difficult to predict. CC is the result of a multistep process that involves the transformation of the normal cervical epithelium into a pre-neoplastic cervical intraepithelial neoplasm that can subsequently transform into invasive cancer [60]. Many studies have shown the tight association between development of CC and infection with HR-HPV [61]. Cell cycle, apoptosis, and proliferation are influenced by molecular interactions of HPV early gene products, particularly E6 and E7 with cellular proteins. These oncogenic proteins have the ability to bind many host regulatory proteins, among the most important of them leading to degradation and functional inactivation of the p53 tumor suppressor and retinoblastoma gene proteins [62]. In the case of the targets for viral infection, it has been suggested that both the basal and stem cells of the cervical epithelium could constitute target for infections by HPV.

Persistence infections with a HR-HPV are a prerequisite for cervical carcinoma development. It was recently found that entry of these viruses is dependent upon a specific proteolytic event that occurs prior to viral endocytosis. Specifically, a proprotein convertase, furin or proprotein convertase 5/6, must cleave the minor capsid protein for infection to proceed [63].

The entry of HPV *in vitro* is initiated by binding to a cell surface receptor in contrast to the *in vivo* situation where the basement membrane has recently been identified as the primary site of virus binding. Binding of HPV triggers conformational changes, which affect both capsid proteins L1 and L2, and such changes are a prerequisite for interaction with the elusive uptake receptor. Furthermore, the productive entry of HPV is a process that occurs slowly and asynchronously and it is characterized by an unusually extended residence on the cell surface [64].

It has been shown that *in vivo* HPV virions bind initially to the base membrane prior to transfer to the basal keratinocyte cell surface [65]. Glycosaminoglycans,

frequently found in the extracellular matrix and on the surface of most cells, especially heparan sulfate, have been suggested as initial attachment receptors for HPV [66, 67].

The fact that HR-HPV could target both basal and stem cells of the cervical epithelium, together with the fact that the entry of HPV DNA to the cell nucleus has been shown to be dependent of the cell cycle, particularly the pass through mitosis has fundamental implications. We propose that the targets for transforming events, caused by HR-HPV infection in the uterine cervical epithelium are stem cells; this is in contrast to the majority of infections where the target of productive infections would be the basal cells, progenitors of normal stem cells and with transiently capacity for replication, that is extended by viral factors interacting with their targets (Fig. 1).

Indeed, infection by HPV has been shown to be dependent on CDK1 activity, consistent with the idea that the final delivery of the HPV genome into the nucleus will depend ultimately on nuclear envelope breakdown [68]. CDK1 activity is correlated with nuclear envelope breakdown and its inhibition prevents this to occur. Another possibility is that nuclear entry of HPV is dependent on the reorganization of the cytoplasmic microtubules to support the formation of the mitotic spindle [69].

In addition, Roberts *et al.* have observed that the columnar epithelium adjacent to the TZ shows a marked increase in sensitivity to infection when disrupted by N-9, a non-ionic, membrane- active surfactant, widely used as spermicidal and known to disrupt the normal architecture of animal and human genital epithelium [65, 70]. This supports the idea that cervical stem cells particularly those of the TZ could be targets for HR-HPV induced transformation events.

CONCLUSIONS

Although limited, the evidence that supports the existence of CSC populations in cervical tumors is relatively convincing. These findings serve as a basis from which to examine the functional parameters of the cancer stem/progenitor cell population and its contribution to the development and progression of CC. However, there remains a need to identify definitive markers for additional selective CSC isolation and enrichment, as well as the need to improve the precision and reproducibility of markers. Transgenic animals and xenograft model systems for human cells engraftment should also be implemented and optimized in order to examine the hallmark characteristics of putative cervical cancer stem cells.

PERSPECTIVES

A complete understanding of the nature of human cervical cancer stem cells that have the potential to contribute significantly to CC development is needed, as well as their relation with HR-HPV infections. This knowledge will certainly allow us to better develop novel strategies for more clinically effective diagnosis of cervical cancer and its treatment.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflicts of interest.

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