

Apoptosis Induction by the 5' NCR of Infectious Bursal Disease Virus

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Abstract: Virus invasion and replication can induce apoptosis and play an important role in the life cycle of viruses. To illustrate the relationship between apoptosis induction and the non-coding region (NCR) of infectious bursal disease virus (IBDV), the 5' NCR of Harbin-1 (a very virulent IBDV) segment A was sequentially deleted using PCR and exchanged with the corresponding region of the mild virulent strain Cj801. IBDV mutants were recovered using a reverse genetics system and named HΔ35, HΔ74, HΔ94, and HCA. Apoptosis in chicken embryo bursal cells was then determined by flow cytometry. The results revealed that the IBDV mutants could induce apoptosis, but with varying capability. As the length of the deleted sequence increased, the capacity to induce apoptosis decreased. However, there was no marked difference in apoptosis induction between HΔ35 and HΔ74. The above results indicate that the NCR may increase the ability to induce apoptosis. Quantitative nested real-time PCR was performed to illustrate the relationship between replication and apoptosis. The result showed an obvious positive correlation for HΔ35 and HCA but an independent process for HΔ74 and HΔ94.

Keywords: Apoptosis induction, flow cytometry assay, NCR deletion of IBDV, quantitative real time PCR.

Infectious bursal disease virus (IBDV) is a non-enveloped virus with a small double-stranded segmented RNA genome belonging to the family Birnaviridae [1]. IBDV is classified into serotypes I and II based on serological response. In chickens, serotype I is pathogenic and causes lymphocyte depletion in the bursa of Fabricius (BF), while serotype II is non-pathogenic [2]. Nieper *et al.* showed that the virus particles for serotype I bind to the lymphocytes in BF, spleen, and thymus like those of serotype II, suggesting that the tropism of IBDV for lymphocyte cells is not decided by the existence of a specific receptor but by another regulating factor, possibly related to the regulation of replication [3]. The regulation of replication is governed by multiple factors such as the 5' non-coding region (NCR), 3' NCR, and stability of mRNA.

Different NCRs of viruses might play important roles in regulation. An internal ribosome entry site (IRES) was detected in the 5' NCR of picornavirus. The IRES was necessary for cap-dependent virus translation, and the predicted tertiary structure suggested a possible binding site for human 18S RNA [4]. The conservative sequences in 5' and 3' NCRs of eight influenza viruses were necessary for nuclease activity [5]. Furthermore, an enhancer and enhancer-regulating region and binding sites for the CCAAT-binding transcription factor NF-Y were identified inside the NCR [6]. Mundt *et al.* sequenced and cloned the 5' and 3' NCRs of both segments of several serotype I strains and one serotype II strain [7]. Their secondary structure showed significant differences between the two serotypes, among which the 5' NCR in serotype I had a Y-shaped stem-

loop structure, but serotype II had a stem-loop structure shaped like a "1"; moreover, inverted terminal repeats were detected in the 5' and 3' NCRs and were possibly related to the formation and maintenance of a hairpin-loop structure. The 5' NCRs of both serotypes had a promoter consisting of conservative 32-nucleotide and IRES region. The initial two bases were GG in the 5' NCR; VPg can bind one or two Gs covalently and prime the synthesis of sense and antisense strands using protein as the primer when there is no genome template [8]. If the structural difference is related to virulence and the capacity to induce apoptosis, this relation has not yet been reported.

IBDV infection results in lymphoid depletion and marked atrophy of the BF, suggesting the induction of apoptosis [9]. Apoptosis processes affect the pathogenesis of IBD. The induction of apoptosis has been reported in peripheral blood lymphocytes [10, 11], Vero cells, and chicken embryo (CE) cells [12]. The apoptotic process is possibly due to the interaction between IBDV infected and non-infected cells, suggesting that infection is not a prerequisite for the induction of IBDV apoptosis [13]. VP5 and VP2 have been demonstrated to be apoptosis inducers [14, 15]. It is hypothesized that VP4 may be involved in an apoptotic signal during virus replication; however, Rodriguez-Lecompte *et al.* demonstrated that VP4 of IBDV serotypes I and II did not induce apoptosis in the chicken B-lymphocyte cell line DT-40 [16].

Apoptosis is related to virulence because apoptosis can lead to immunosuppression. To date, research about virulence and apoptosis has focused on VP1, VP2, VP3, VP4, and VP5. There have been fewer reports of the function of NCRs and the relationship between NCRs and virulence in regard to apoptosis. Furthermore, research findings have differed largely in regard to the relationship between viral replication and apoptosis. Tham *et al.* suggested that apoptosis and virus replication are independent processes

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[12]. However, Jungmann *et al.* found a correlation between the proportion of apoptotic cells and IBDV replication [13]. In 2002, the full-length nucleotide sequence of the genome for the very virulent IBDV (vvIBDV) Harbin-1 was obtained, and 5' and 3' end sequences were analyzed; the result showed that the 5' NCR of segment A contained a promoter and IRES region. However, the function of other regions in the 5' NCR is still unknown. In this study, we examined the relationship between the 5' NCR of segment A and apoptosis. We also investigated the role of the 5' NCR in IBDV replication efficiency by deleting the promoter and the interspace between the promoter and the IRES, and exchanging the 5' NCR with the corresponding region of the mild virulence strain Cj801.

MATERIALS AND METHODS

Viruses and Cells

The vvIBDV strain Harbin-1 was provided by the Harbin Veterinary Research Institute of the Chinese Academy of Agricultural Sciences. When specific-pathogen-free (SPF) chickens were inoculated with Harbin-1 at a dose of 2×10^3 EID₅₀, 60% mortality resulted. In contrast, the cell-adaptable strain Cj801 caused 0% mortality when given to SPF chickens at a dose of 2×10^3 EID₅₀. Primary bursal cells were derived from 18-day-old embryonated SPF eggs (Merial, Beijing, China) and grown in Dulbecco's modified Eagle's medium (DMEM, Sigma Aldrich, St. Louis, MO, USA) supplemented with 10% fetal calf serum (FCS) and maintained with DMEM containing 5% FBS. Transfection experiments were performed on primary bursal cells.

Polyadenylation of dsRNA and Amplification of 3' Ends

Isolation of viral RNA was performed with a MiniBest viral RNA/DNA extraction kit according to the manufacturer's protocol (Takara Biotechnology, Dalian, China). The 3' termini of both strands of segments A and B were polyadenylated following the protocol described by Mundt [7]. The polyadenylation reaction mixture (50 μ l), consisting of 1 μ g viral RNA, 50 mM Tris-Cl (pH 7.9), 10 mM MgCl₂, 250 mM NaCl, 0.25 mM adenosine triphosphate (ATP), 2.5 mM MnCl₂, 250 μ g bovine serum albumin (BSA), and 1 unit poly(A) polymerase (Invitrogen, Carlsbad, CA, USA), was added and incubated for 20 minutes at 37°C. Next, poly(A) RNA was extracted using equal volumes of phenol and chloroform and recovered by ethanol. Reverse

transcription was performed using Oligo (dT) as the primer following the protocol given for Invitrogen's 3'-RACE kit with minor alteration. PCR was performed using P1 (segment A) and P2 (segment B) as the primers (Table 1). The PCR profile was as follows: 1 cycle of 94°C for 4 min and 30 cycles of 94°C for 30 s, 55°C for 30 s, and 72°C for 10 s.

Amplification of the 5' Ends

Amplification of the 5' ends followed the method described by Frohman [17], with minor modifications, using Invitrogen's 5'-RACE reaction kit. Briefly, purified viral RNA was denatured at 98°C for 5 min, and then the first strand of cDNA was synthesized using reverse transcriptase superscript II (Invitrogen) with the P3 and P6 primers (Table 1). cDNA was purified by column chromatography (Invitrogen) and tailed by poly(C) using terminal deoxynucleotidyl transferase (TdT, Invitrogen). The cDNA was then amplified with P4/P5 (segment A) and P7 (segment B) primers and an anchor primer (provided as part of the kit). The PCR profile was as follows: segment A: 1 cycle of 94°C for 5 min and 35 cycles of 94°C for 30 s, 58°C for 30 s, and 72°C for 30 s; segment B: 35 cycles of 94°C for 30 s, 63°C for 30 s, and 72°C for 30 s.

Cloning of NCR PCR Fragments and Sequencing

PCR fragments were recovered by glassmilk after electrophoresis in 1.5% agarose gels and ligated into the pGEM-T vector (Promega, Madison, WI, USA). A positive clone was obtained using X-gal and IPTG: α complementation as described by Sambrook [18] and identified by PCR after transforming *Escherichia coli* strain DH5 α . Both strands of respective inserts were sequenced by Takara Biotechnology in Dalian of China and Shanghai Sangon Biological Engineering Technology and Services Co., Ltd. in China.

Construction of Full-Length cDNA Clones Using Fusion PCR

Segments A and B of the Harbin-1 clone included six clones, including the coding region (CR) of segments A and B, designated as pGEM-T-HA and pGEM-T-HB, respectively, and the NCRs of segments A (pGEM-T-H5'A and pGEM-T-H3'A) and B (pGEM-T-H5'B and pGEM-T-H3'B) (Table 2) [19, 20]. All recombinant plasmids were based on pGEM-T (Promega). There was an area of partial overlap between the CR clone and the NCR clone for

Table1. Primers Used for Cloning of 5' and 3' Non-Coding Regions

Primer	Sequence	Position	Terminal
P1	CCCAATGCTCCAACACAGAGAC	+ ^a (nt3092-3113)	3'segA
P2	CGCCAAAAGGAGAGCCGCTAA	+ (nt3178-3196)	3'segB
P3	TTGAGCCAGGGAAACCAGGG	- ^b (nt310-330)	5'segA
P4	CTGACCTGAGAGTGTGCTTC	- (nt232-251)	5'segA
P5	GATCTTGCAGGTCGTCATCGC	- (nt130-150)	5'segA
P6	GTCGGTCTCATACTCCTCATTC	- (nt318-340)	5'segB
P7	AGGGCTGGCAAGCGGATCCT	- (nt248-268)	5'segB

^a Sense.

^b Antisense.

segments A and B. The overlapped area lacked an appropriate restriction site; thus fusion PCR was used to ligate the NCR and CR to obtain a full-length cDNA clone for segments A and B. Oligonucleotides HACR1, HACR2, HANCR1, HANCR2, HBCR1, HBCR2, HBNCR1, and HBNCR2 were used for segments A and B (Table 3). For transcription *in vitro*, the EcoRI site and T7 promoter were introduced into the 5' end of the oligonucleotides, the XbaI site at the 3' end in segment A, and the XhoI site at the 3' end in segment B. The fusion PCR product of segment A was ligated into the pMD-18T-simple vector (Takara Biotechnology) to obtain full-length cDNA of the segment A clone, which was named pRHA.

Because 52 bps at the 3' extreme end of the 3' non-coding region of segment B were truncated [19], a series of oligonucleotides including HBRA1, HBRA2, and HBRB to HBRE (Table 3) was designed to fill in the 3' extreme end by

use of consecutive PCR and the primer walking method. The fusion PCR product was ligated into the pMD-18T-simple vector (Takara Biotechnology) to obtain the full-length clone of segment B, named pRHB, under the control of the T7 promoter.

Table 2. Plasmids Used for Fusion PCR and their Characteristics

Plasmid	Main Feature
pGEM-T-HA	Harbin-1A segment coding sequence
pGEM-T-HB	Harbin-1B segment coding sequence
pGEM-T-H5'A	Harbin-1A segment5' non-coding sequence
pGEM-T-H3'A	Harbin-1A segment3' non-coding sequence
pGEM-T-H5'B	Harbin-1B segment5' non-coding sequence
pGEM-T-H3'B	Harbin-1B segment3' non-coding sequence

Table 3. Oligonucleotides Used for Amplification of Serotype I Strain of Harbin-1 Sequences

Oligonucleotide Orientation Position Name			
<u>CTCCTCCTTCTACAACGCTATCAT</u> XbaI	sense	71-924	HACR1
<u>GAATCTAGAGGGGACCCGCGAACG</u> EcoRI	antisense	3246-3261	HACR2
<u>GGAATTCTAATACGACTCACTATAGGGGGGATACGATCGGTCTG</u> <u>GATCTTGCAGGTTTCGTATCGC</u>	sense	1-16	HANCR1
<u>TGGCTACTAGGGGCGATGC</u>	antisense	128-149	HANCR2
<u>CTTCTTGAGTGGTTCATC</u> EcoRI	sense	42-60	HBCR1
<u>GGAATTCTAATACGACTCACTATAGGATACGATGGGTCTGACCCCT</u> <u>GTCATCATGGTGGCAGAATCAT</u> EcoRI	antisense	2756-2775	HBCR2
<u>GGAATTCTAATACGACTCACTATAGGATACGATGG</u> <u>GGTCTGGGGTTAGTGTCTTCTTGTGAGTGGTTCATC</u>	sense	1-21	HBNCR1
<u>GGCCGGGATACGGGGTCTGGGGTTAGTGTCT</u> <u>GCAGGCGAAGGCCGGGATACGGGGTC</u>	antisense	98-120	HBNCR2
<u>CCGAGGCGAAGGCCGGGGAT</u> <u>GCCCCGCAGGCGAAGG</u> XhoI	sense	1-11	HBRA1
<u>CTCGAGGGGGCCCCGCAGGCGAAG</u> EcoRI	antisense	2756-2794	HBRA2
<u>GGAATTCTAATACGACTCACTATAGACCCGGGACAGGCT</u> EcoRI	antisense	2776-2808	HBRB
<u>GGAATTCTAATACGACTCACTATAGTCTCTCTTCTACAATG</u> EcoRI	antisense	2791-2817	HBRC
<u>GGAATTCTAATACGACTCACTATAGTCATTGATGGTCAG</u> EcoRI	antisense	2799-2819	HBRD
<u>GGAATTCTAATACGACTCACTATAGGATACGATCGGTCTGACCCGG</u> XbaI	antisense	2807-2823	HBRE
<u>GAATCTAGAGGGGACCCGCGAACG</u> TAACCGTCTCAGCTTACCC	antisense	2808-2827	HBRF
TCAGGATTTGGGATCAGCTC	sense	35	PD35
CCAACAGCGAGATAACC	sense	74	PD74
GGCGACCGTAACGACAG	sense	94	PD94
	sense	1	PCA
	antisense	3176-3199	PDA
	sense	625-644	outside 1
	antisense	1250-1267	outside 2
	sense	1023-1040	inside 1
	antisense	1216-1232	inside 2

The restriction sites used are given in bold text and the appropriate restriction enzymes are named. Underlined nucleotides are virus-specific. The T7 promoter is shown in italics. The positions where the primers bind (nucleotide number) are in accordance with the sequence of strain P2 (7).

Construction of the 5' NCR Mutant

A series of oligonucleotides was designed for deletion of the 5' NCR in segment A. Deletion mutants were obtained using PCR and sequenced by the Takara Biotechnology and Sangon companies. In addition, the NCR of Harbin-1 segment A was exchanged with the attenuated IBDV strain Cj-801 NCR (data not show) using fusion PCR. Table 3 lists the oligonucleotides used for the deletion.

Transcription and Transfection of Synthetic RNAs

Plasmids pMD-18HA and pUC18-CA and their mutant clones pMD18 Δ 35, pMD18 Δ 74, pUC18 Δ 94, and pMD18-RHB were digested with XbaI and XhoI, respectively, and used as templates for transcription *in vitro* with T7 RNA polymerase (Promega). Briefly, restriction enzyme cleavage assays were adjusted to 0.5% sodium dodecyl sulfate (SDS) and incubated with proteinase K (0.5 mg/ml) for 1 h at 37°C. The linearized DNA templates (1 μ g) were recovered after ethanol precipitation and were added to transcription reaction mixtures (50 ml) containing 40 mM Tris-HCl (pH 7.9), 10 mM NaCl, 6 mM MgCl₂, 2 mM spermidine, 0.5 mM ATP, 0.5 mM cytidine triphosphate (CTP), 0.5 mM uridine triphosphate (UTP), 0.1 mM guanosine triphosphate (GTP), 0.25 mM cap analog [m⁷G(5')ppp (5')G] (Promega), 120 units ribonuclease inhibitor (Takara Biotechnology), and 150 units T7 RNA polymerase (Promega), and incubated at 37°C for 1 h. Synthetic RNA transcripts were purified by phenol-chloroform extraction and ethanol precipitation. As controls, the transcription products were treated with either DNase or RNase (Promega) before the purification step.

Primary bursal cells of chicken were prepared according to standard protocols and maintained at 37°C and 5% CO₂ atmosphere in DMEM supplemented with 5% FCS. After primary bursal cells were grown to 80% confluency in 35-mm dishes, the cells were washed twice with DMEM (serum-free) and incubated at 37°C for 10 min after each wash in a CO₂ incubator. Simultaneously, 60 μ l DMEM was incubated with 6 μ l of Lipofectin reagent (Invitrogen) for 60 min in a polystyrene tube at room temperature to form a Lipofectin-DMEM mixture. Synthetic RNA transcripts of both segments resuspended in 30 μ l of water treated with diethylpyrocarbonate (DEPC) were added to the DMEM-Lipofectin mixture, mixed gently, and incubated on ice for 5 min. After removing the DMEM from the monolayer cells in the 35-mm dishes and replacing it with 800 μ l of fresh DMEM, the nucleic acid-containing mixture was added drop-wise to the cells and swirled gently. After 2 h of incubation at 37°C, the mixture was replaced with DMEM containing 5% FCS (without rinsing the cells) and further incubated at 37°C for desired time intervals.

Virus Recovery from cRNA and Identification

Two days after transfection, cells were frozen-thawed three times and centrifuged at 700 \times g to remove cellular debris. The supernatant was passaged four times in the primary bursal cells. To screen the recombinant virus from a large quantity of samples, antigen-capture enzyme-linked immunosorbent assay (AC-ELISA) was performed using a monoclonal antibody, Mab6 [21]. If the optical density (OD) value of a sample was greater than the mean OD plus three

times the standard deviation of the negative control sample, then the sample was considered to be positive and was stored at -86°C for future use.

Part of the hypervariable region was amplified using nested real-time PRC (RT-PCR) to determine virus presence. If the target band appeared on the gel (209 bp), a positive result was obtained. Table 3 lists the primers including outside 1, outside 2, inside 1, and inside 2.

Research on the Apoptosis Induced by Recombinant IBDV Induction

Sub-G₀/G₁ DNA Peak Technique

Following the procedure described by Lam [22], 18 million primary bursal cells (3 \times 10⁶/ml) were inoculated with 100 pfu virus mutants. To quantify the number of apoptotic cells, infected and control bursal cells were collected at different time intervals, incubated with propidium iodide, and subjected to flow cytometry. Floating bursal cells and attached cells were removed from Petri dishes by trypsin treatment and collected in one tube by centrifugation; they were then washed with phosphate buffered saline (PBS) once and centrifuged to remove the PBS. The cells were fixed by 1 ml ice-cold 70% ethanol and kept for 1 h at 4°C in ethanol. Ethanol was removed by centrifugation for 10 min at 300 \times g. The pellet was resuspended in PBS for 5 min. After the cells were filtered once with a 400-mesh screen, PBS was removed by centrifugation at 1000 rpm for 5 min. Next, 1 \times 10⁶ cells were incubated for 30 min at room temperature with 1 ml propidium iodide (50 μ g/ml in PBS supplemented with 1% BSA and 40 μ g/ml RNase A). The DNA content of the 1 \times 10⁶ cells was analyzed by flow cytometry. In the flow cytometry assay, propidium iodide produces red fluorescence with sub-ion fluorescence excitation. The wave length for excitation is 488 nm and the wave length for emission is greater than 630 nm. The front-scatter light plot was analyzed against the side-scatter light plot. Compared with cells in the G₀/G₁ phase of the cell cycle, apoptotic cells appeared as a population of cells with a lower DNA content (sub-G₀/G₁ DNA peak technique) [23]. Cells to the left of the G₀/G₁ peak were therefore considered apoptotic. Each experiment was repeated at least three times.

Measurement of the Apoptosis Index

Measurement of the apoptosis index was performed using the procedure described by Duke and Cohen with minor modifications [24]. First, 5 μ l of ethidium bromide and acridine orange were added to 100 μ l of the abovementioned cells (10⁶/ml). The mixture was mixed well and incubated for 20-30 min at room temperature; 50-60 μ l of the mixture were then added to a slide, sealed with a cover glass, and observed with a fluorescent microscope using excitation filters BG12 and BV and a 515 nm or SP3 interdictio filter. Each sample was observed in ten visual fields and the mean value was obtained. The apoptosis index is represented as the number of apoptotic cells per 100 bursal cells.

Plaque Assay of Recombinant Virus [25]

The titer of virus was determined using a plaque assay and expressed as PFU/ml.

Characterization of IBDV Mutants in the Cell Culture

To assay the replication of IBDV mutants, quantitative nested RT-PCR was performed. Eighteen million primary bursal cells were inoculated with IBDV mutants and Harbin-1 at the same titer and incubated for 5 days at 37°C in a 5% CO₂ incubator. The cells were collected, frozen and thawed three times, and the resulting supernatant was used for isolating viral RNA. Viral RNA was extracted with a viral RNA/DNA extraction kit (Takara) following the manufacturer's instructions. Conventional RT-PCR was performed; the first PCR was amplified with outside 1 and outside 2 primers (Table 3). The second PCR was performed with inside 1 and inside 2 primers (Table 3). The second PCR products were detected by electrophoresis through a 1% agarose gel stained with ethidium bromide and visualized under UV light. If there was no primer dimer and non-specific bands appeared, then the first PCR products could be used as the template for the quantitative nested RT-PCR. Quantitative RT-PCR was performed by the SYBR Green I method and determined by absolute quantification using the inside 1 and inside 2 primers (Table 3). The quantitative nested RT-PCR protocol was 94°C for 5 min; 40 cycles of 94°C for 30 s, 60°C for 30 s, 72°C for 30 s; and 72°C for 10 min. The cycle threshold (Ct) values were measured in triplicate. Regression analysis of the mean values of three replicates for the log₁₀ diluted pRHA was used to generate standard curves. The coefficient of variance for three replicates was calculated for the IBDV specific reactions. The reproducibility of the assay was tested by analysis of variance (ANOVA) comparing repeated runs of samples, and mean values generated at individual time points were compared using Student's *t*-test using SPSS 11.5 software.

RESULTS

Identification of the Recombinant Plasmid

The mutant clones were designated pUC-CA, pMD-18HA, pMD-18Δ35, pMD-18Δ74, and pMD-18Δ94 (Table 4). The abovementioned plasmids were digested by EcoRI and XbaI to produce the expected bands of 3.2 and 2.7 kb, respectively, and mutant clones of segment A were obtained.

Identification of Full-Length Segment A

The ligation between coding and non-coding regions was only identified by sequencing because the NCRs were so

short that they could not be identified by restriction enzyme digestion. The part linking the deleted NCR and CR was sequenced, and the results are given in Fig. (1).

RNA Synthesis *In Vitro* and Recovery of Virus

The deletion mutant clones for full-length cDNA of segments A and B were linearized by XbaI and XhoI, respectively. The linearized plasmid was recovered and used as the template for transcription. Plus-sense RNA was synthesized under T7 RNA polymerase, with 1.5 kb and 1.2 kb for segments A and B, respectively.

Four virus mutants were screened from 30 samples by AC-ELISA. Their OD values were greater than the mean OD value plus three times the standard deviation of the negative control sample. The recovered virus mutants were designated rHarbin-1, HCA, HΔ35, HΔ74, and HΔ94. The results indicated that the deletion mutant virus could be recovered without reference to partial or full deletion of the NCR.

Bursal Cells Infected with IBDV Mutants Undergo Apoptosis

Apoptotic cells appeared to the left of the G₀/G₁ cell cycle because fragmentation of genomic DNA resulted in the production of hypodiploid DNA. In infected cell cultures, the proportion of cells with lower DNA content increased considerably, indicating the induction of apoptosis. Apoptotic cells show typical hypodiploid peaks in histograms; namely, peaks appear to the left of the diploid peak (G₁) and have characteristics of typical apoptotic cells at 48 h post infection (p.i.). The more apoptotic cells, the more evident the hypodiploid peak is. The percentages of apoptotic cells were as follows: 15.6% (rHarbin-1), 16.8% (HCA), 13.8% (HΔ35), 12.5% (HΔ74), 11.7% (HΔ94), and 6.0% (CK) at 48 h p.i. (Fig. 2). The number of apoptotic bursal cells in infected groups increased at 4-48 h p.i., but increased markedly at 8-48 h p.i. (Fig. 3). There were significant differences in the percentages of apoptosis cells among the infected groups, with HCA inducing a higher percentage of apoptosis than the other virus mutants.

The results indicated that the ability to induce apoptosis decreased as the deletion length of the NCR increased. Virus mutants with NCR deletion had a decreased ability to induce apoptosis relative to rHarbin-1 and HCA in which the NCRs remained intact; however, HCA had a slightly stronger ability to induce apoptosis than HA. The virus mutant HΔ94, in which the NCR was totally deleted, had the least ability to induce apoptosis in the infected group. There was no marked difference in the abilities of HΔ35 and HΔ74 to induce apoptosis.

Determination of the Apoptotic Index of Bursal Cells

Infected and uninfected cells were stained with acridine orange and ethidium bromide. Apoptotic cells fluoresce orange, while normal cells fluoresce green under a

Table 4. The Characteristics of 5' NCR Harbin-1 Mutants

Name	Characteristics
pMD-18HA	Contains full-length 5' NCR
pUC-CA	Contains full-length 5' NCR of IBDV CJ-801; other sequence is the same as Harbin-1
pMD-18Δ35	Deleted promoter-like region in Harbin-1 NCR
pMD-18Δ74	Deleted region between promoter-like region and internal ribosome entry site in 5' NCR of Harbin-1
pMD-18Δ94	Deleted promoter-like region and ribosome binding site

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ABBREVIATIONS

CR	=	Coding region
DMEM	=	Dulbecco's modified Eagle's medium
IBDV	=	Infectious bursal disease virus
IRES	=	Internal ribosome entry site
NCR	=	Non-coding region
PCR	=	Polymerase chain reaction
TdT	=	Deoxynucleotidyl transferase
SPF	=	Specific-pathogen-free
vvIBDV	=	Very virulent IBDV

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