

The Possible Ultra Structural Ameliorative Effect of Taurine in Rat's Liver Treated with Monosodium Glutamate (MSG)

Ibrahim M. A. ElAgouza¹, Dalia E. El Nashar^{*,2} and Saad S. Eissa³

¹Faculty of Science, Zoology Department, Cairo University, Egypt

²Health Radiation Research Department, National Center for Radiation Research and Technology (NCRRT). Atomic Energy Authority, Cairo, Egypt

³Surgical Pathology and National Cancer Institute, Cairo University, Egypt

Abstract: Monosodium glutamate (MSG) is a food additive with a wide range of biological effects but its high dose and long period of using can cause a toxic effect on liver. Taurine is a derivative that produces antioxidant in many tissues including liver. The current research concerned with biochemical, histopathological and ultra structural changes in rats liver treated with MSG and throw light on the possible ameliorative effect of taurine – 150 male albino rats were divided into five group each containing 30 rats (controls, taurine, MSG, therapeutic and protective group). Serum AST, ALT, ALP and bilirubin (direct and total) were measured, as well as histopathological and ultrastructural changes in liver tissues were examined in all groups. The rats received MSG orally in a dose 400 mg/Kg. B. wt for 8 weeks (MSG group), while taurine induced orally in a daily dose at 1000 mg/Kg. B. wt alone (Taurine group) or after MSG (therapeutic) or before MSG (protective) for the same period, in addition to frank control group received orally 2 ml saline per day during the experiment. The results showed marked and gradual increase in the serum levels of AST, ALT, ALP and bilirubin (direct and total) in MSG group as compared to control group. The histopathological structure of the same group showed hepatic and local hemorrhage as well as necrosis of hepatocytes with pyknotic nuclei after 4 weeks of the treatment, while after 8 weeks portal edema, thickening in wall of bile duct and portal infiltration were observed. Ultra structure examination in the same group after 8 weeks showed hyaline degeneration; rough endoplasmic reticulum with irregular nuclear membrane and cytoplasm rich in glycogen granules, Interestingly, the results showed a significant restore of hepatic injury as evidenced by attenuation of hepatocellular enzyme and bilirubin release and reduction of hepatocyte necrosis and then portal infiltration in therapeutic group reduction of hepatocyte necrosis and then portal infiltration therapeutic group. While complete normalization in plasma ALT, AST, ALK and bilirubin levels with complete restore of histological and ultra structural arrangement of hepatocyte were observed in protective group.

In conclusion, the results confirm the hepatotoxic effect of MSG and improve the hepatoprotective effect of taurine, especially when it administrated as a protective substance than therapeutic. Also, we advice to add a suitable dose of taurine to any food containing MSG as a food additive.

INTRODUCTION

Monosodium glutamate is a substance widely used as flavoring agent in the whole world. It is the sodium salts of glutamic acid and a form of glutamate. It is absorbed very quickly from gastrointestinal tract and could spike blood plasma levels of glutamate [1].

In addition, MSG causes edema-degeneration –necrosis in liver [2] and induced the oxidative stress and hepatotoxicity in rats [3]. Moreover, MSG treatment of mice induces obesity and diabetes with steatosis and steatohepatitis and they suggested that MSG should be potentially withdrawn from the food chain [4].

Taurine, one of the known amino acids has many important and beneficial effects on the human body. It has been

implicated in many physiological functions, pharmacological actions and pathological conditions [5]. Other physiological roles attributed to taurine, includes membrane stabilization, antioxidation, osmoregulation, neuromodulation, detoxification and regulation of calcium homeostasis [6].

Taurine fulfills a role in promoting hepatic stellate cell's apoptosis. In the case of hepatic fibrosis; it mitigates the liver injury, decrease the expression of transforming growth factor beta-1 and relieves hepatic fibrosis [7]. In addition, it was effective in the prevention of lipopolysaccharide-induced hepatotoxicity and pro-oxidant status [8], reduces oxidative stress and prevents progression of hepatic fibrosis in CCl₄-induced hepatic damage rats as well as inhibits transformation of the hepatic stellate cells [9], protects the integrity of the hepatic tissues by stabilizing the reactive oxygen species mediated lipid peroxidation and protein carbonyl formation and it acts as antioxidant in tamoxifen-induced hepatotoxicity [10], plays a beneficial role for the prevention of cisplatin hepatotoxicity [11] and protects the liver against ethanol-induced oxidative injury *via* its effects on the sulfur-amino acid metabolism [12].

*Address correspondence to this author at the Health Radiation Research Department, National Center for Radiation Research and Technology (NCRRT). Atomic Energy Authority, Cairo, Egypt; E-mail: daliaessamy@yahoo.com

The target of the present investigation is concerned with the possible biochemical, pathological and Ultra structural (E/M) changes that may occur in rats' liver under the effect of MSG (food additive) and shed light on the possible protective and therapeutic effects of taurine.

MATERIALS AND METHODS

Materials

One hundred and fifty male albino rats (*rattus*) were used in the present studies. The weights of rats ranged from 120-130grams and their ages ranged from 2-3 months and obtained from the Egyptian Organization for Biological product and Vaccines.

Animals were maintained on a standard pellet diet and water, for 7 days, the diet consists of about 20% protein, 5% fibers, 3.5% fats and 6.5% ash and supplied with vitamins mixtures. Feed and water were available at the time of experimental period. The animals were housed in suitable cages (each cage contain 10 rats) and kept at optimal conditions of temperature and humidity.

Chemicals

The chemical used monosodium glutamate (MSG) from Fluca Co. with purity >98% NT and taurine supplied by Sigma with a 99% purity. Both drugs were orally administered.

Experimental Animal Grouping

The animals were divided into 5 equal groups each contain 30 male rats:

1-Control Group

Animals of this group received orally saline daily, parallel to the drug treated groups.

2-Monosodium Glutamate (MSG) Treated Group

The rats received a daily dose of MSG (400mg.kgB.wt./d.) for eight weeks.

3-Taurine Treated Group

The rats received orally a daily dose of taurine (1000mg.kgB.wt./d.) for eight weeks.

4-Therapeutic Group

Animals of this group received oral dose of MSG (400mg.kgB.wt./d.) then treated orally after one hour with taurine in a dose of (1000mg.kgB.wt./d.) for eight weeks.

5-Protected Group

Animals of this group received oral dose of taurine in a dose of (1000mg.kgB.wt./d.) then after one hour they administered with oral dose of MSG (400mg.kgB.wt./d.) for eight weeks.

Blood Samples

Blood sampling was made after 2, 4, 6 and 8 weeks. Samples were drawn, after overnight fasting (using a special

micropipettes introduced through the inner contuse of rats orbit into the cavernus sins and the head of the rat was tilted down) in clean, and sterile serum tubes. Blood samples were centrifuged for 10 minutes, at 3000 r. p. m. within an hour of the blood collection and the sera were obtained to measure the liver functions:

Serum aspartate-amino transferase (AST) and alanine-amino transferase (ALT) were determined calorimetrically [13], using commercial kits purchased from the Biomerieux, France.

Serum alkaline phosphatase activity (ALP) activity was determined calorimetrically [14], using commercial kits purchased from the Biomerieux, France.

Serum bilirubin (Direct and Total) was measured [15]. The reagents used were kits purchased from the Boehringer, Germany.

Histopathological Studies

After scarification and dissection, liver was removed immediately and fixed in 10% formal saline and neutral buffered formalin for 7 days, then the tissue was washed and dehydrated in ascending grads of ethyl alcohol cleared in benzene and impregnated in paraffin for 1.5 hour in the oven at 55°C. Serial section 5µm was cut and stained with Hematoxylin and Eosin (H&E) as described by [16].

Electron Microscopic Study

The liver from all groups were examined by E. M.

RESULTS

Biochemical, histopathological and electron microscopic examination were done for liver to evaluate the possible therapeutic and protective effect of taurine against the hepatotoxic effect of MSG.

The Table 1 showed that all liver enzymes measured in this work (AST, ALT, ALP) and bilirubin (Direct and Total) were non significantly changed ($P>0.05$) from control in taurine treated group, while in MSG group the latter enzymes and bilirubin (D and T) were significant, ($P<0.01$) elevated after 2, 4, 6 and 8 weeks, but these elevations were markedly decreased, especially in protective than in therapeutic group after 8 weeks.

Supporting, in MSG group, Fig. (2 and 3) showed cellular and nuclear pleomorphism and apparent vacuolation in the cytoplasm of the hepatocytes. Areas of necrosis and hemorrhage involved most of the hepatic parenchyma were noticed. The hepatocytes in numerous hepatic lobules showed variation in nuclear size, some have small nuclei, while in others the cells were large with longer nuclei. The blood vessels are congested with an increase in Kupffer cells. At the peripheral zones of the hepatic lobules the hepatocytes were markedly vacuolated.

In Fig. (4a), liver of rat in therapeutic group showed, extensive diffuse fatty degeneration, and Fig. (4b), in protective group, showed mild congestion and absence of necrosis with limited fatty change of the hepatocytes.

Table 1. The Level of ALT, AST, ALP, and Bilirubin (Direct & Total) in the Different Experimental Groups at the Different Periods of the Study (Liver Function)

Experimental Groups	Parameters	Periods of Study (Weeks)				F-ratio
		2 weeks	4 weeks	6 weeks	8 weeks	
Control	ALT (U/L)	10.2 ± 0.24 a	10.1 ± 0.23 a	10.3 ± 0.21 a	10.4 ± 0.26 a	0.286 NS
	AST (U/L)	11.2 ± 0.24 a	11.0 ± 0.25 a	11.3 ± 0.21 a	11.4 ± 0.26 a	0.332 NS
	ALP (U/L)	112 ± 7.13 a	113.2 ± 7.2 a	117.9 ± 7.3 a	123.6 ± 7.2 b	0.53 NS
	D.bilirub. (mg/dl)	0.185 ± 0.014	0.175 ± 0.014	0.192 ± 0.013	0.196 ± 0.015	0.412 NS
	T.bilirub. (mg/dl)	0.7 ± 0.05 b	0.67 ± 0.05 b	0.69 ± 0.03 b	0.76 ± 0.06 b	0.586 NS
TAURINE (1000mg/kg/d)	ALT (U/L)	9.1 ± 0.45 a	8.6 ± 0.45 a	8.9 ± 0.50 a	9.9 ± 0.34 a	1.56 NS
	AST (U/L)	10.1 ± 0.45 a	9.7 ± 0.42 a	9.9 ± 0.50 a	10.9 ± 0.34 a	1.448 NS
	ALP (U/L)	107.1 ± 8.40 a	100.1 ± 8.09 a	100.8 ± 8.57 a	113.5 ± 6.7 ab	0.62 NS
	D.bilirub. (mg/dl)	0.192 ± 0.015	0.173 ± 0.016	0.175 ± 0.016	0.187 ± 0.013	0.347 NS
	T.bilirub. (mg/dl)	0.60 ± 0.04 a b	0.56 ± 0.03 ab	0.56 ± 0.04 a	0.57 ± 0.03 a	0.303 NS
MSG (400 mg.kg./d)	ALT (U/L)	12.6 ± 0.70 Ab	21.2 ± 2.04 BC	31.3 ± 1.57 Cc	50.0 ± 3.08 Dd	62.15 **
	AST (U/L)	13.6 ± 0.70 Ab	22.2 ± 2.04 BC	32.3 ± 1.57 Cc	51.0 ± 3.08 Dd	62.15 **
	ALP (U/L)	162.2 ± 3.8 Ad	197.5 ± 4.8 Bd	227.5 ± 4.8 B d	258.5 ± 6.7 Dc	64.59 **
	D.bilirub. (mg/dl)	0.564 ± 0.014	0.574 ± 0.012	0.591 ± 0.011	0.675 ± 0.01	18.24 **
	T.bilirub. (mg/dl)	1.589 ± 0.04	1.671 ± 0.03 ab	1.782 ± 0.04 a	1.801 ± 0.029 a	7.99 **
MSG (400 mg/kg/d) Then TAURINE (1000 mg/kg/d) [Therapeutic]	ALT (U/L)	14.2 ± 0.55 A c	17.2 ± 0.55 A b	22.5 ± 0.6 Bd	30.5 ± 1.96 C b	42.13 **
	AST (U/L)	15.2 ± 0.55 Ac	18.2 ± 0.55 A b	23.5 ± 0.6 Bb	31.5 ± 1.96 C b	42.13 **
	ALP (U/L)	133 ± 3.2 A b c	155 ± 4.1 BC b	148 ± 3.05 Bb	158.6 ± 3.0 C c	11.39 **
	D.bilirub. (mg/dl)	0.67 ± 0.011	0.659 ± 0.011	0.62 ± 0.011	0.60 ± 0.013	8.10 **
	T.bilirub. (mg/dl)	0.95 ± 0.03 a	0.93 ± 0.02 a	0.88 ± 0.03 a	0.80 ± 0.03 a	5.76 **
TAURINE (1000mg//kg/d) Then MSG (400 mg/kg/d) [protective]	ALT (U/L)	9.3 ± 0.36 a	9.2 ± 0.32 a	9.2 ± 0.32 a	9.6 ± 0.33 a	0.32 **
	AST (U/L)	10.3 ± 0.36 a	10.2 ± 0.32 a	10.2 ± 0.32 a	10.6 ± 0.33 a	0.31 **
	ALP (U/L)	107.2 ± 8.09 a	101.2 ± 5.96 a	101 ± 6.17 a	113.5 ± 6.7 a b	0.76 **
	D.bilirub. (mg/dl)	0.173 ± 0.015	0.173 ± 0.015	0.167 ± 0.014	0.186 ± 0.014	0.27 **
	T.bilirub. (mg/dl)	0.58 ± 0.04 a b	0.55 ± 0.05 a b	0.56 ± 0.03 a	0.52 ± 0.03 a	0.39 **
F-ratio	ALT (U/L)	22.38 **	42.58 **	215.60 **	144.16 **	
	AST (U/L)	22.38 **	42.09 **	215.60 **	144.31 **	
	ALP (U/L)	10.652 **	35.72 **	56.09 **	102.46 **	
	D.bilirub. (mg/dl)	1.57 N S	0.57 N S	0.602 **	0.76 N S	
	T.bilirub. (mg/dl)	1.87 N S	1.46 N S	1.71 N S	3.94 **	

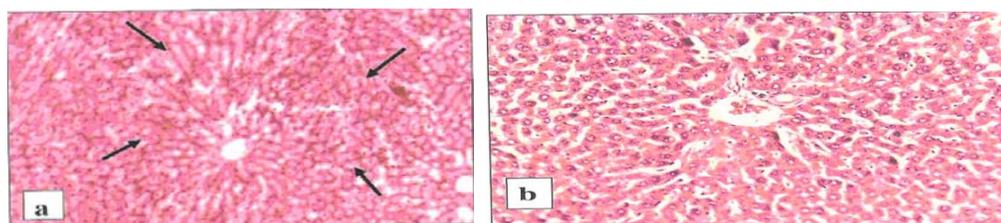


Fig. (1). a) Liver of control, untreated rat showing the normal histological structure of hepatic lobule. (H and E X 200). **b)** Liver of rat treated with taurine 1000 mg/kg body weight/day showing normal architecture of hepatic lobule. (H and E X 400).

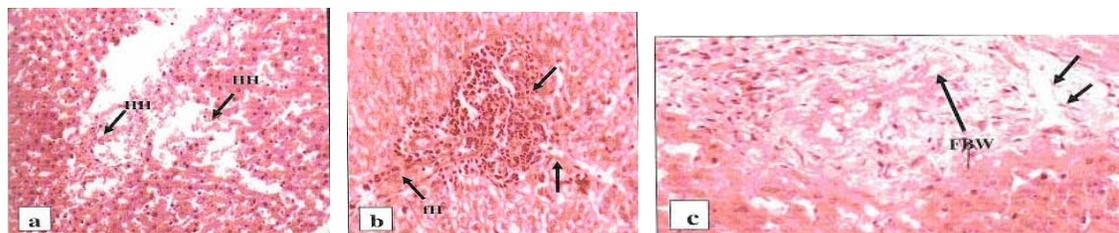


Fig. (2). a) Liver of rat treated with MSG 400mg/kg body weight/day for one month showing hepatic haemorrhage (HH) dispersed the hepatocytes from each other. (H and E X 200). **b)** Liver of rat treated with MSG 400mg/kg body weight/day for one month showing focal haemorrhage (FH) as well as necrosis of hepatocytes with pyknotic nuclei. (H and E X 200). **c)** Liver of rat treated with MSG 400mg/kg body weight/day for one month showing fibrosis of bile duct wall (FBW). (H and E X 200).

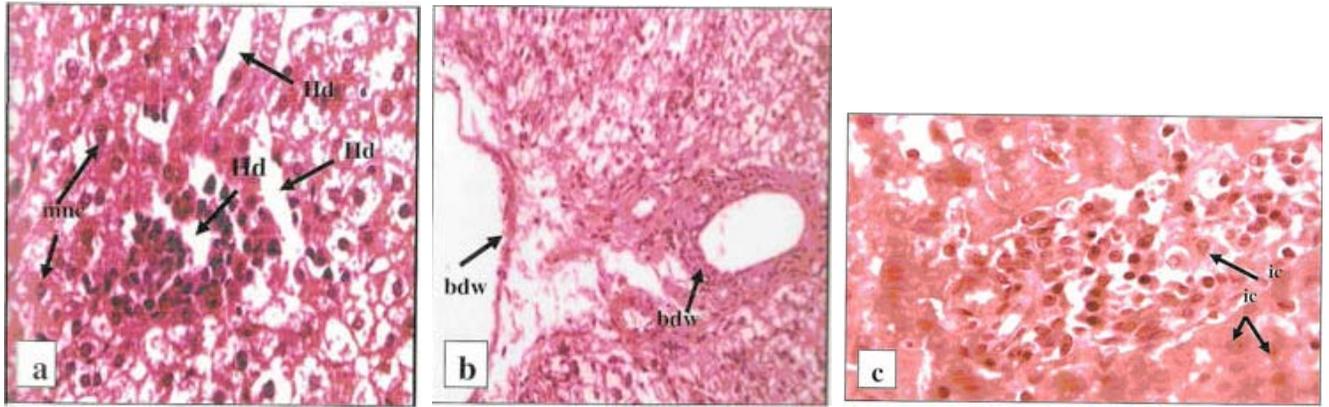


Fig. (3). a) Liver of rat treated with MSG 400mg/kg body weight/day for two months showing hydrobic degeneration (Hd) of hepatocytes as well as focal hepatic necrosis replaced by mononuclear cells infiltration (mnc). (H and E X 400). b) Liver of rat treated with MSG 400mg/kg body weight/day for two months showing portal edema and thickening in the wall of bile duct (bdw). (H and E X 200). c) Liver of rat treated with MSG 400mg/kg body weight/day for two months showing portal infiltration with mononuclear inflammatory cells (ic). (H and E X 400).

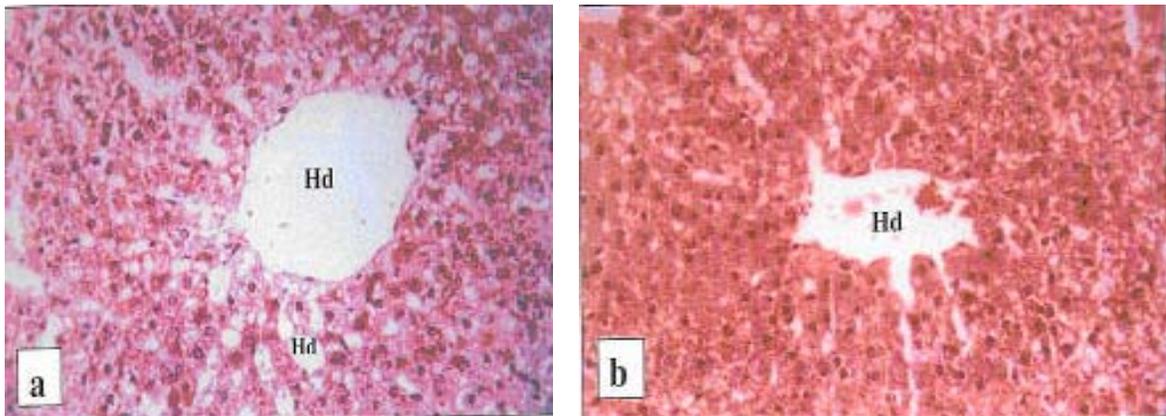


Fig. (4). a) Liver of rat treated with MSG 400mg/kg body weight/day then with taurine 1000 mg/kg body weight/day for two months (therapeutic) showing hydrobic degeneration (Hd) of hepatocytes. (H and E X 200). b) Liver of rat treated with taurine 1000 mg/kg body weight/day then with MSG 400mg/kg body weight/day for two months(protective) showing slight hydrobic degeneration (Hd) of few hepatocytes. (H and E X 200).

The ultrastructural examination of (Fig. 5) shows normal hepatocytes with round nucleus, different shapes of mitochondria, and clear cytoplasm with normal endoplasmic reticulum (E. R.).

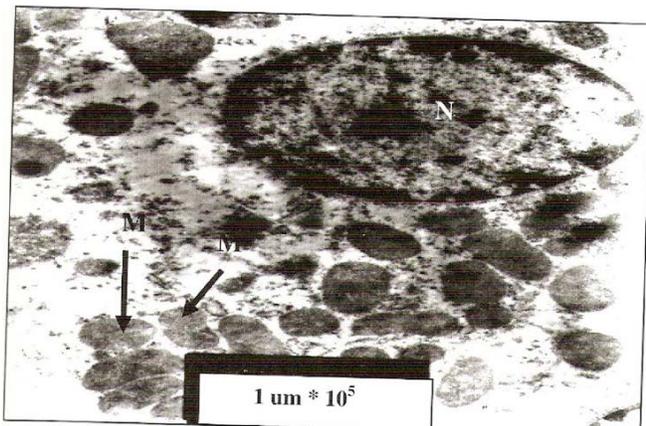


Fig. (5). Part of normal hepatocyte with round nucleus (N). there were different shapes of mitochondria (M).

At the same time, the treatment for 8 weeks lead to different ultra structural changes has been occurred, such as the ultra structural examination in MSG group after 8 weeks showed areas of hyaline degeneration (fatty) of the cytoplasm. Few organelles and few E. R, cytoplasm were rich in glycogen granules, while membranes were slightly irregular with normal cytoplasm and few degenerated areas. The same group showed rounded nucleus, rounded mitochondria, and few cistern of rough E. R. and stage of mitotic figure, where nuclear membrane lost (Fig. 6a-e).

While the hepatocyte examined by EM in therapeutic groups after 8 weeks showed mitochondria and endoplasmic reticulum, and hyaline degeneration of cytoplasm (Fig. 7a), normal nucleus and mitochondria (Fig. 7b) and the reticulate cell showed few organelle and degenerated cytoplasm, normal nucleus, mitochondria and rough E. R. (Fig. 7c), small areas of hyaline degeneration (possible dissolved glycogen), rounded nucleus mitochondria (Fig. 7d), fatty degeneration in between the mitochondria and cistern of rough E. R. (Fig. 7e).

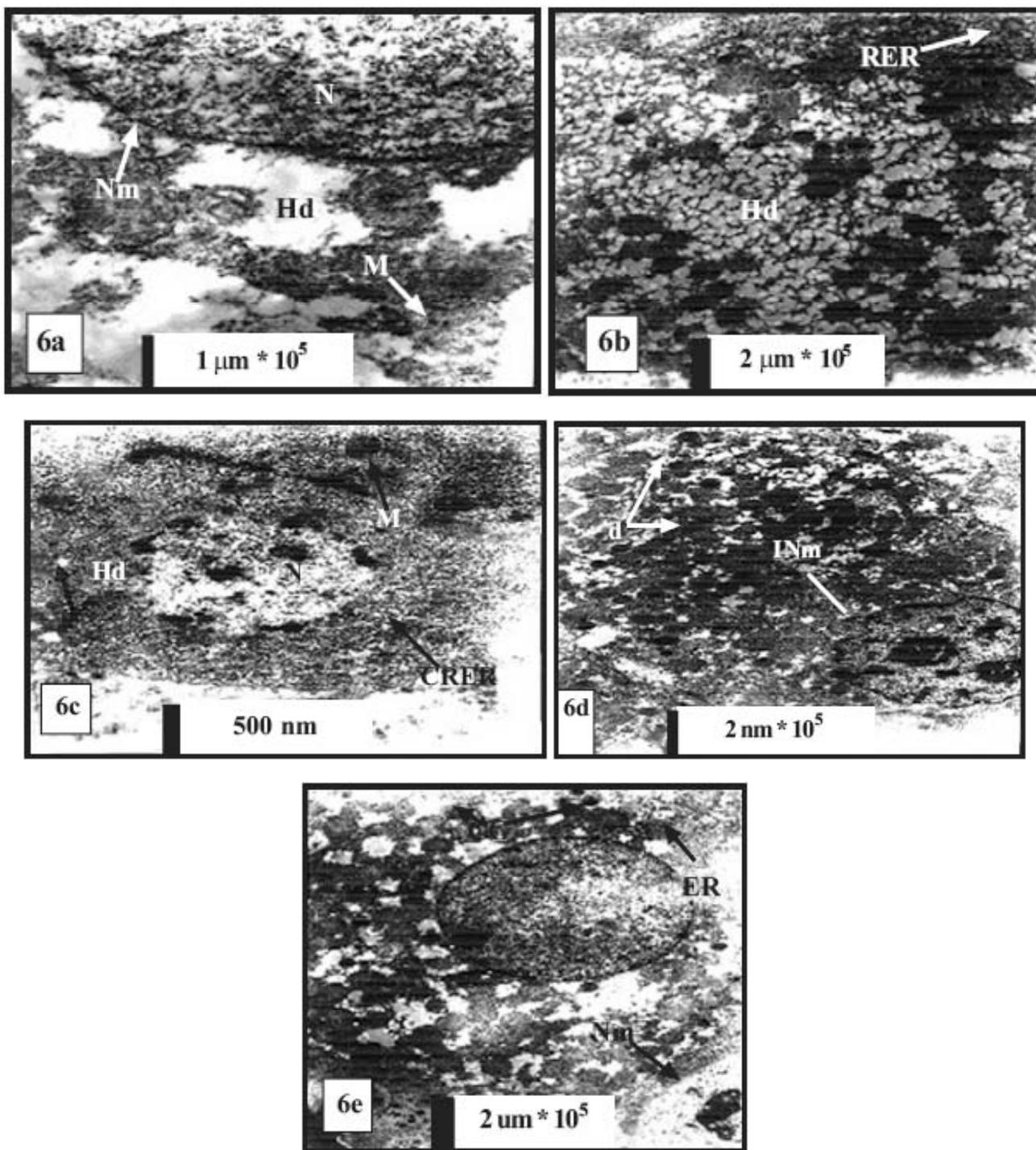


Fig. (6). (a). Part of hepatocyte of liver of a rat treated with MSG 400mg/kg body weight/day for two months shows nucleus (N) with typical nuclear membrane (Nm). The cytoplasm shows rounded mitochondria with typical mitochondria cristea (M). Areas of hyaline degeneration (fatty) of the cytoplasm are appeared (Hd). (b). Part of cytoplasm of liver of a rat treated with MSG 400mg/kg body weight/day for two months shows few organelles. Hyaline degeneration of the cytoplasm (Hd) and few Rough Endoplasmic Reticulum (R.E.R) are appeared. (c). hepatocyte of liver of a rat treated with MSG 400mg/kg body weight/day for two months shows round nucleus (N) rounded mitochondria (M), areas of hyaline or fatty degeneration(Hd) and few cistern of rough endoplasmic reticulum (CRER).(d). Part of hepatocyte of a rat treated with MSG 400mg/kg body weight/day for two months shows slightly irregular nuclear membrane (INm). Normal cytoplasm with few degenerated areas appeared (d). (e). Part of hepatocyte of a rat treated with MSG 400mg/kg body weight/day for two months in stage of mitotic figure where nuclear membrane lost (Nm). Also the cytoplasm rich in glycogen granules (GG) and few Endoplasmic Reticulum (ER).

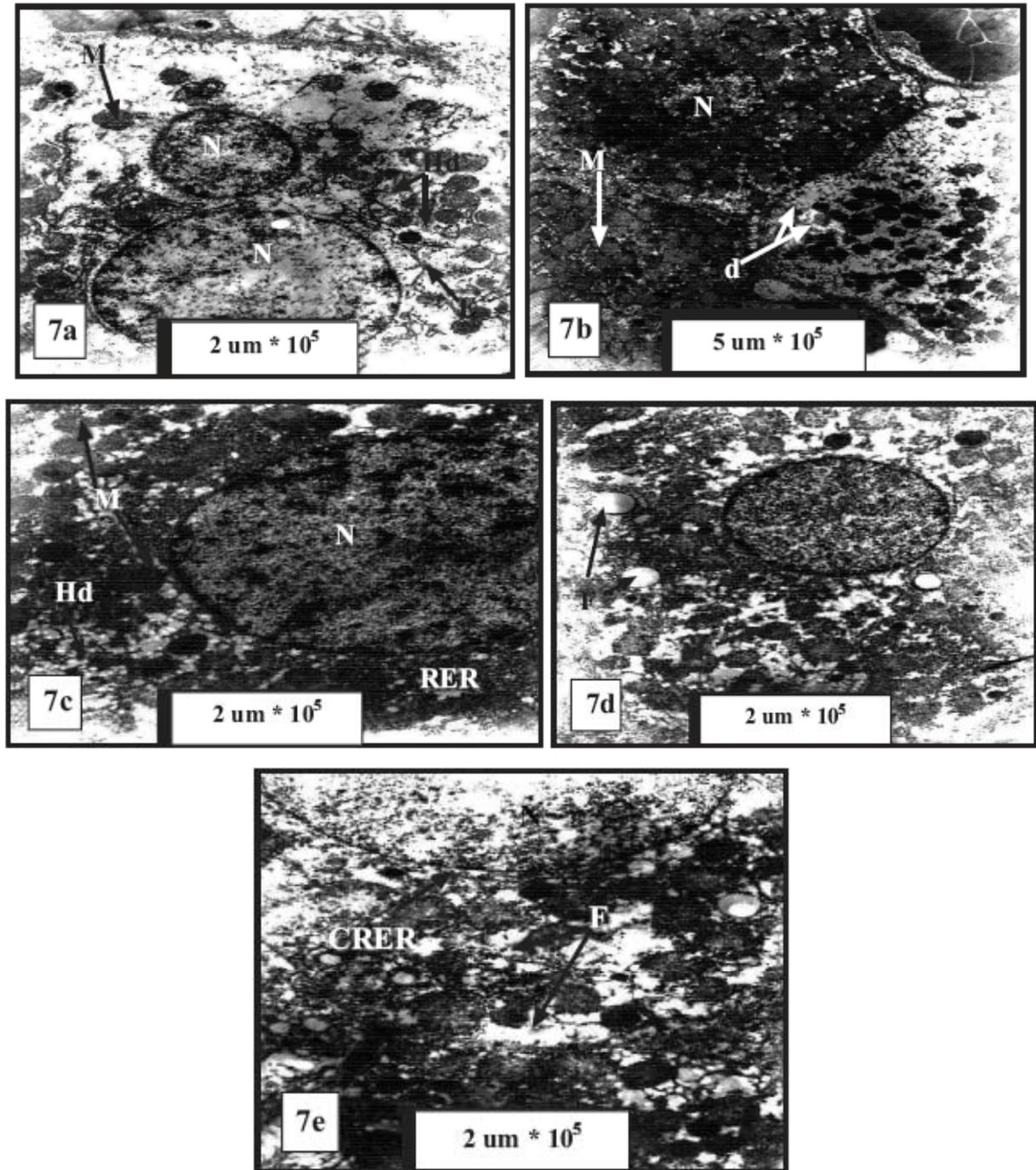


Fig. (7). (a). Part of binucleated hepatocyte N(normal nuclei) of a rat treated with MSG 400mg/kg body weight/day then with taurine 1000 mg/kg body weight/day for two months shows few mitochondria (M) There was Endoplasmic Reticulum (ER) hyaline degeneration of cytoplasm (Hd). (b). Three adjacent hepatocyte of a rat treated with MSG 400mg/kg body weight/day then with taurine 1000 mg/kg body weight/day for two months.two normal shows normal nucleus (N) and mitochondria (M). The cell shows few organelle and degenerated cytoplasm (d). (c). Part of hepatocyte of a rat treated with MSG 400mg/kg body weight/day then with taurine 1000 mg/kg body weight/day for two months shows normal nucleus (N), mitochondria (M) and Rough Endoplasmic Reticulum (R.E.R). There are small areas of hyaline degeneration (Hd), possible dissolved glycogen. (d). Hepatocyte of a rat treated with MSG 400mg/kg body weight/day then with taurine 1000 mg/kg body weight/day for two months shows rounded nucleus (N), fat globules at cytoplasm (F). Normal rounded mitochondria (M). (e). Hepatocyte of a rat treated with MSG 400mg/kg body weight/day then with taurine 1000 mg/kg body weight/day for two months shows normal nucleus (N). Fatty degeneration (F) in between the mitochondria and cistern of rough endoplasmic reticulum (CRER).

Interestingly, the ultra structural examination of the protective group after two months showed large rounded

nucleus, normal mitochondria with normal cristae and normal rough E. R. (Fig. 8a), few lysosomes (Fig. 8b), cyto-

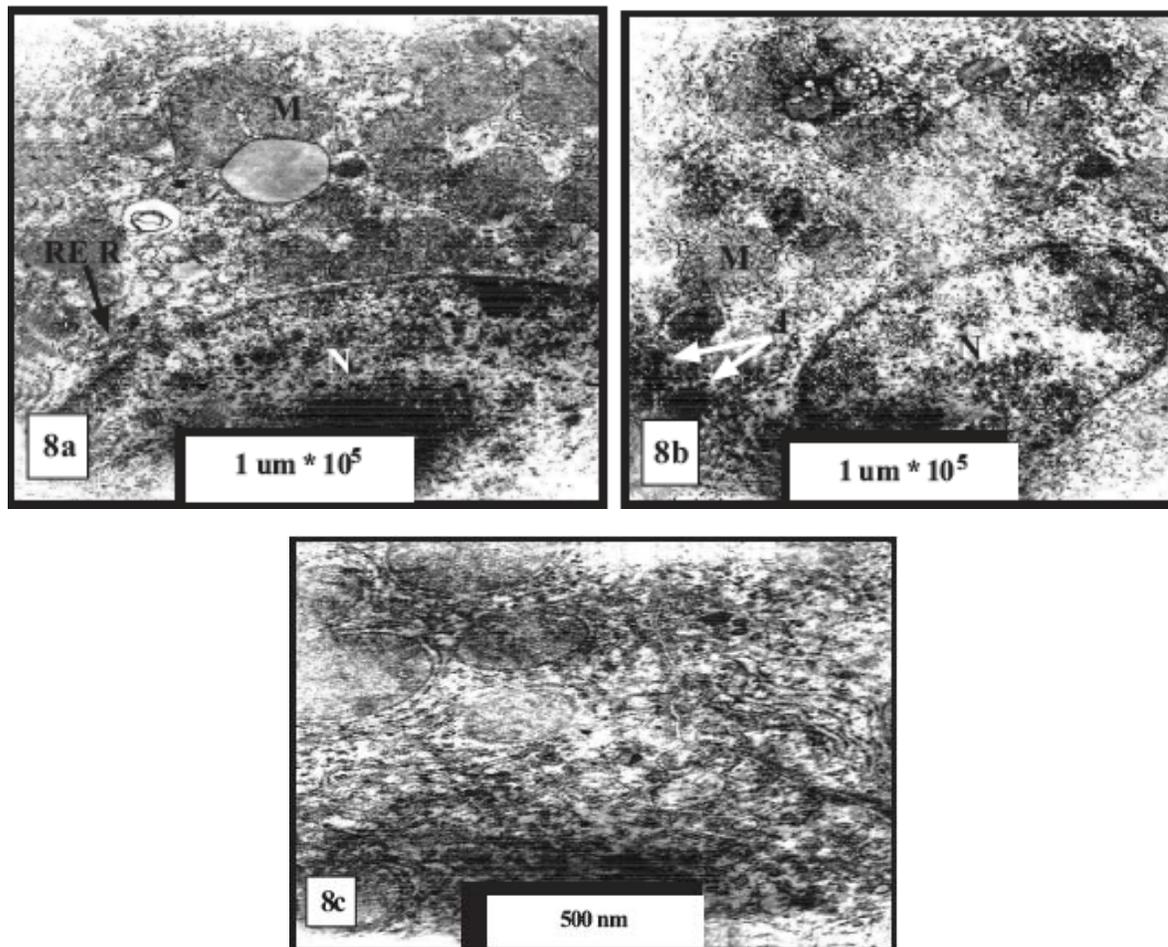


Fig. (8). (a). Part of hepatocyte of rat treated with taurine 1000 mg/kg body weight/day then with MSG 400mg/kg body weight/day for two months with large rounded nucleus (N). There were normal mitochondria with normal cristae (M). Also rough endoplasmic reticulum (R.E.R) was normal. (b). Part of hepatocyte of rat treated with taurine 1000 mg/kg body weight/day then with MSG 400mg/kg body weight/day for two months shows normal hepatocyte, normal nucleus (N), normal mitochondria (M) and few lysosomes (L). (c). Part of hepatocyte of rat treated with taurine 1000 mg/kg body weight/day then with MSG 400mg/kg body weight/day for two months shows normal hepatocyte. The cytoplasm appear normal.

plasm with normal appearance and normal appearance of cistern of rough E. R. (Fig. 8c).

DISCUSSION

MSG is widely used as a food additive to improve the taste of food, because it potentiates the activity of gustatory nerve [17]. In particular, this nerve mediate sweet taste [18].

Regarding to hepatotoxic effect of MSG the result showed gradual elevation ($P < 0.01$) in the serum levels of liver enzymes and bilirubin, which exhibited a value four times higher than normal after 8 weeks. This confirms the cumulative adverse effect of MSG. Moreover, hydropic degeneration of hepatocyte, focal hepatic necrosis, portal edema, focal hemorrhage, necrosis and fibrosis of bile ducts all these histopathological disfigurement were also observed Fig. (2-4) when compared with control and rats treated with taurine only (Fig. 1). Concomitantly, the ultra structural examination for the same group showed hyaline degeneration of

the cytoplasm, few endoplasmic reticulum and irregular nuclear membrane which may lost in some figures after 8 weeks.

Confirming our observation, I noticed that MSG administration initially attacked the peripheral hepatocytes in the central lobules of the liver tissues leading to hepatocellular degeneration [19]. Recently, calcium level increased in liver by administrating MSG to adult male mice [20]. The authors revealed this condition to the increase in oxidative stress effect. An increased intracellular concentration of Ca^{+2} could theoretically act either to enhance lipid peroxidation or to stimulate degeneration of phospholipids. Similarly hepatocellular damage with elevation of all serum liver enzymes was observed [21-23] after ingesting the rat with MSG. Concomitantly, it was attributed with the histopathological and ultrastructural change in rats live treated with MSG to the accumulation of highly reactive and cytotoxic intermediate, which may cause centrilobular necrosis of hepatocyte [2, 24-26]. In conclusion, reactive oxygen species, lipid peroxi-

dation and disordered Ca^{+2} homeostasis are the mechanisms that may have contributory role in the development of liver injury after MSG excess.

On the other hand, administration of taurine before MSG (Therapeutically) resulted in a significant restore of hepatic injury as evidenced by attenuation and hepatocellular enzymes and bilirubin release and reduction of hepatocyte necrosis with slight hydropic degeneration of few hepatocyte.

Same observations were postulated that therapeutic administration of taurine after the hepatotoxic substance elevated the intracellular taurine, that increase the ability to protect against oxidant-induced hepatotoxicity through its cytoprotective effect [27]. The most interesting point in this work is the normal restore of liver enzymes and bilirubin release accompanied with its normal histological and ultrastructural architecture and hepatocyte in the group of rats treated with taurine before MSG (protectively) [28]. In conclusion, when taurine was used protectively (12 h before the oxidant treatment) noticed normalization in plasma level of AST, ALT and ALK and disappearance of hepatocyte necrosis [28]. Lastly, taurine can protect the hepatocyte against MSG, through different mechanisms included cell membrane stabilization [9, 24], antioxidation and detoxification [27-30], osmoregulation [29, 31-33], neuromodulation [34] and regulation of Ca^{+2} homeostasis [27, 35, 36].

In conclusion, on the basis of these findings we advise to add a suitable dose of taurine to any food containing MSG as a food addictive, because there is no doubt that taurine is considered as powerful hepatoprotective substance that can protect the hepatocyte through different mechanisms.

REFERENCES

- [1] Stegink L, Filer L, Baker G. Plasma amino acid concentrations in normal adults ingesting aspartame and monosodium L-glutamate as part of a soup/beverage meal. *Metabolism* 1987; 36: 1073-79.
- [2] Ortiz G, Bitzer-Quintero O, Zarate C, *et al.* *Biomed Pharmacother* 2006; 60: 86-91.
- [3] Onyema O, Farombi E, Emerole G, Ukoha A, Onyeze G. Effect of vitamin E on monosodium glutamate induced hepatotoxicity And oxidative stress in rats. *Indian J Biochem Biophys* 2006; 43(1): 20-4.
- [4] Nakanishi Y, Tsuneyama K, Fujimoto M, *et al.* Monosodium glutamate: a viillian and promoter of liver inflammation and dysplasia. *J Autoimmun* 2008; 30: 42-50.
- [5] Nakashima T, Shima T, Mitsuyoshi H, *et al.* Taurine in the liver. The function of taurine conjugated with bile acids. *Adv Exp Med Biol* 1996; 403: 85-92.
- [6] Birdsall T. Therapeutic applications of taurine. *Altern Med Rev* 1998; 3: 128-36.
- [7] Liang J, Zhang X, Yang G, *et al.* Observation of the promotion effect taurine on hepatic stellate cell's apoptosis in rat hepatic fibrosis model. *Sichuan Da Xue Xue Bao Yi Xue Ban* 2005; 36: 365-7.
- [8] Balkan J, Paldar F, Dogru-Abbasoglu S, Aykac-Toker, Uysal M. The effect of taurine or betaine pretreatment on hepatotoxicity and prooxidant status induced by lipopolysaccharide treatment in the liver of rats. *Eur J Gastroenterol Hepatol* 2005; 17(9): 917-21.
- [9] Miyazaki T, Karube M, Matsuzaki Y, *et al.* Taurine inhibits oxidative damage and prevents fibrosis in carbon tetrachloride-induced hepatic fibrosis. *J Hepatol* 2005; 43: 117-25.
- [10] Tabassum H, Rehman H, Banerjee B, Raisuddin S, Parvez S. Attenuation of tamoxifen-induced hepatotoxicity by taurine in mice. *Clin Chem Acta* 2006; 23: 24-30.
- [11] Liao Y, Lu X, Lu C, Li G, Jin Y, Tang H. Selection of agents for prevention of cisplatin-induced hepatotoxicity. *Pharmacol Res* 2008; 57: 125-31.
- [12] Kim S, Jung Y, Kwondo Y, Kim Y. Alleviation of acute ethanol-induced liver injury and impaired metabolomics of S-containing substances by betaine supplementation. *Biochem Biophys Res Commun* 2008; 368: 893-8.
- [13] Retman S, Frankel S, Amer J. Determination of SGOT and SGPT. *Clin Pathol* 1957; 28: 56-60.
- [14] Wootton J. Determination of alkaline phosphatase in different animals by P-nitrophenyl phosphate. *Acta Biochem* 1974; 39: 72-84.
- [15] Jendrassik L, Grof A. Measuring of bilirubin. *Biochemistry* 1938; 7297: 81-5.
- [16] Bancroft J, Stevens A. Theory and practice of histological techniques. Churchill Livingstone Edinburgh-London and New York, 1977; p. 98.
- [17] Schiffman S, Gill J. Psychophysical and neurophysical taste responses to glutamate and purinergic compounds. In: Kawamura Y, Kare M, Eds. *ABasic Taste. Umami*; New York: Marcel Dekker Inc., 1987; pp. 271-288.
- [18] Nishijo H, Ono T, Norgren R. Parabranchial gustatory neural responses to monosodium glutamate ingested by awake rats. *Physiol Behav* 1991; 49: 965-71.
- [19] Andrew O, Eweka. M. Histological studies of the effects of MSG on kidney of adult Wistar rats. *Internet J Health* 2007; 7: 211-5.
- [20] Cairoli P, Pieraccini S, Sironi M, Morelli C, Speranza G, Manitto P. Studies on umami taste. Synthesis of new guanosine5'-phosphate derivatives and their synergistic effect with monosodium glutamate. *J Agric Food Chem* 2008; 56: 1043-50.
- [21] Barogi S, Baracca A, Cavazzoni M, Parenti CG, Lenaz G. Effect of the oxidative stress induced by adriamycin on rat hepatocyte bioenergetics during ageing. *Mech Ageing Dev* 2000; 113: 1-21.
- [22] Pritsos C, Ma J. Basal and drug-induced antioxidant enzyme activities correlate with age-dependent doxorubicin oxidativetoxicity. *Chem Biol Interact* 2000; 127: 1-11.
- [23] Nocito A, Dahm F, Jochum W, *et al.* Serotonin mediates oxidative stress and mitochondrial toxicity in a murine model of non alcoholic steatohepatitis. *Gastroenterology* 2007; 133(2): 608-18.
- [24] Eoin W, Jiang H, Paul R, Qiong D, Elaine K, David B. Role of taurine in preventing acetaminophen-induced hepatic injury in the rat. *Am J Physiol Gastrointest Liver Physiol* 2001; 280: 1274-9.
- [25] Diniz Y, Faine L, Galdardi C, *et al.* Monosodium glutamate in standard and high-fiber diets: metabolic syndrome and oxidative stress in rats. *Nutrition* 2005; 21: 749-55.
- [26] Matyskova R, Maletinska L, Maixnerova J, Pirnik Z, Kiss A, Zelezna B. Comparison of the obesity phenotypes related to monosodium glutamate effect on arcuate nucleus and/or the high fat diet feeding in C57B1/6 and NMR1 mice. *Physiol Res Rev* 2007 [Epub ahead of print].
- [27] Schemmer P, Liang R, Kinctus M, *et al.* Taurine improves graft survival after experimental liver transplantation. *Liver Transplant* 2005; 11: 950-9.
- [28] Metayer S, Seiliez I, Collin A, *et al.* Mechanisms through which sulfur amino acids control protein metabolism and oxidative status. *J Nutr Biochem* 2008; 19: 207-15.
- [29] Huxtable R. Physiological actions of taurine. *Physiol Rev* 1992; 72: 101-63.
- [30] Klamt F, Shacter E. Taurine Chloramine, an oxidant derived from neutrophils induces apoptosis in Human B lymphoma cells through mitochondrial damage. *J Biol Chem* 2005; 280: 21346-52.
- [31] Lambert H. Regulation of the cellular content of the organic osmolyte taurine in mammalian cells. *Neurochem Res* 2004; 29: 27-63.
- [32] Franco R, Rodriguez R, Pasantes -Morales H. Mechanisms of ATP potentiation of hyposmotic taurine release in Swiss 3T3 fibroblasts. *Pflugers Arch* 2004; 449: 159-69.
- [33] Friis B, firiborg R, Schneder L, Nielsen B, Lambert H, Christensen S. Cell Shrinkage as a signal to apoptosis in NIH3T3 fibroblasts. *J Physiol* 2005; 567: 427-43.

- [34] Selina N, sapronov S, Bul'on V, Kuznetsova N. The neuro-protector effect of a new taurine derivative on a model of compression spinal cord trauma in rats. *EKSP Klin Farmakol* 1999; 68: 45-8.
- [35] Zhao P, Huang Y, Cheng J. Taurine antagonizes calcium overload induced by glutamate or chemical hypoxia in cultured rat hippocampal neurosis. *Neurosci Lett* 1999; 268: 25-8.
- [36] Foos T, Wu J. The role of taurine in the central nervous system and the modulation of intracellular calcium homostasis. *Neurochem Res* 2002; 27: 21-6.

Received: June 12, 2009

Revised: October 28, 2009

Accepted: November 22, 2009

© ElAgouza *et al.*; Licensee *Bentham Open*.

This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>), which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.